Frequency and Antimicrobial Susceptibility of Bacteria Isolated from Patients Hospitalized with Pneumonia in United States Medical Centers (2017–2018)

ATS 2019 Poster #8470

INTRODUCTION

- The initial antimicrobial management of patients with pneumonia is determined mostly by the understanding of causative pathogens, and there is very little current information regarding the frequency and antimicrobial susceptibility of organisms causing healthcare-associated pneumonia
- Although Staphylococcus aureus is a significant cause of pneumonia in hospitalized patients, the importance of gram-negative organisms such as Pseudomonas aeruginosa and Enterobacterales species, mainly Klebsiella pneumoniae, Enterobacter spp., and Escherichia coli, has increased substantially in recent years
- Ceftazidime-avibactam is approved by the United States Food and Drug Administration (US FDA) and by the European Medicines Agency (EMA) to treat hospital-acquired bacterial pneumonia (HABP), including ventilatorassociated bacterial pneumonia (VABP)
- Ceftazidime-avibactam is also approved to treat complicated intra-abdominal infections (cIAIs) in combination with metronidazole, as well as complicated urinary tract infections, including pyelonephritis
- We evaluated the frequency and antimicrobial susceptibility of gram-negative bacteria isolated from patients hospitalized with pneumonia in US medical centers and assessed the activity and spectrum of 2 recently approved β-lactamase inhibitor combinations, ceftazidime-avibactam and ceftolozane-tazobactam, and many other antimicrobial agents currently used to treat pneumonia

MATERIALS AND METHODS

Bacterial isolates

- A total of 7,787 clinical isolates were consecutively collected from patients hospitalized with pneumonia (1/patient) in 66 US medical centers in 2017–2018
- Only isolates determined to be significant by local criteria as the reported probable cause of infection were included in the program

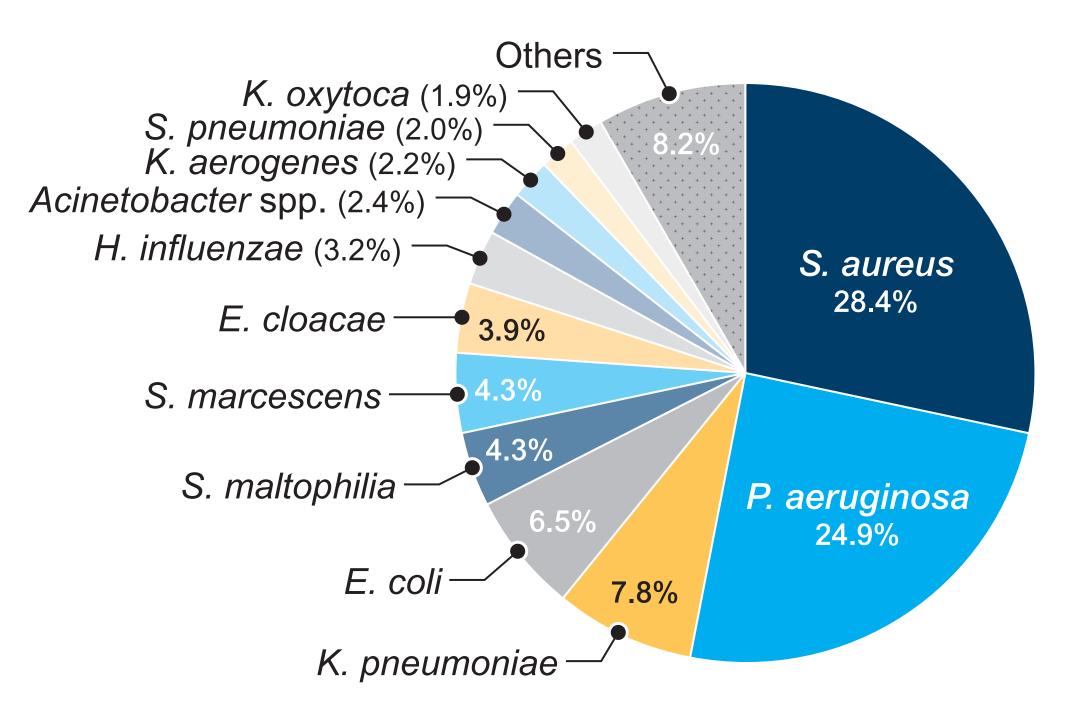
Resistant subsets

- Carbapenem-resistant Enterobacterales (CRE) isolates were defined as displaying imipenem, meropenem, and/or doripenem MIC values at ≥4 mg/L (CLSI, 2019)
- Imipenem was not applied to *Proteus mirabilis* or indole-positive Proteeae due to the intrinsically elevated MIC values
- Multidrug-resistant (MDR) and extensively drug-resistant (XDR) Enterobacterales and P. aeruginosa strains were classified according to recommended guidelines (Magiorakos et al., 2012) as follows:
- MDR = nonsusceptible (NS; CLSI breakpoints) to at least 3 antimicrobial classes
- XDR = susceptible (S) to 2 or fewer antimicrobial classes

Susceptibility testing

- The broth microdilution test method was conducted according to CLSI
- Avibactam was provided by Allergan (Irvine, California, USA) and combined with ceftazidime (avibactam at fixed concentration of 4 mg/L) for susceptibility testing
- Ceftolozane stock solution was obtained from ThermoFisher Scientific (Cleveland, Ohio, USA) and combined with tazobactam (acquired from United States Pharmacopeia [USP]) at fixed concentration of 4 mg/L for susceptibility testing
- All other compounds were obtained from USP or Sigma-Aldrich (St. Louis, Missouri, USA)

Figure 1 Frequency of organisms isolated from patients hospitalized with pneumonia in US medical centers (2017–2018)



JMI Laboratories, North Liberty, Iowa, USA

Screening for β-lactamase-encoding genes

- Enterobacterales isolates displaying MIC values $\geq 2 \text{ mg/L}$ for at least 2 β -lactams (ie, ceftazidime, ceftriaxone, aztreonam, or cefepime) and all CRE isolates were tested for β-lactamase-encoding genes using next-generation sequencing
- Libraries were normalized using the bead-based normalization procedure (Illumina) and sequenced on MiSeq
- FASTQ files were assembled using SPAdes Assembler and subjected to a proprietary software (JMI Laboratories) for screening of β -lactamase genes

RESULTS

- The most common organisms were S. aureus (28.4%), P. aeruginosa (24.9%), K. pneumoniae (7.8%), E. coli (6.5%), Stenotrophomonas maltophilia (4.3%), Serratia marcescens (4.3%), and Enterobacter cloacae (3.9%); overall, 68.1% of the organisms were gram-negative and 31.9% were gram-positive (Figure 1)
- All S. aureus isolates were susceptible (S) to dalbavancin (MIC₉₀, 0.06 mg/L), linezolid, teicoplanin, and vancomycin; 99.9%S to tigecycline; 95.8%S to ceftaroline; and 55.9%S to oxacillin (Table 1)
- Ceftazidime-avibactam and ceftolozane-tazobactam were very active against *P. aeruginosa* isolates and exhibited identical coverage against these organisms (95.7%S; Table 2)
- Ceftazidime-avibactam retained activity against *P. aeruginosa* isolates nonsusceptible to meropenem (86.6%S), MDR (83.1%S), XDR (77.2%S), and nonsusceptible to ceftazidime, cefepime, meropenem, and piperacillintazobactam (68.4%S; Table 2 and Figure 2)
- Ceftazidime-avibactam exhibited potent activity against Enterobacterales (MIC_{50/90}, 0.12/0.5 mg/L; 99.9%S), including extended-spectrum β-lactamase (ESBL)-producing strains (MIC_{50/90}, 0.25/0.5 mg/L; 100.0%S), CRE (MIC_{50/90}, 1/2 mg/L; 96.3%S), MDR (MIC_{50/90}, 0.25/1 mg/L; 99.3%S), and XĎŘ isolates (MIC_{50/90}, 1/2 mg/L; 96.1%S; ŤaĎle 3 and Figure 3)

Table 1 Antimicrobial susceptibility of *S. aureus* isolates (n=2,080) from patients hospitalized with pneumonia in US medical centers

Antimicrobial	MIC ₅₀ ^a	MIC _{on} a	%S ^a	%R ^a
Dalbavancin	0.03	0.06	100.0	
Ceftaroline	0.25	1	95.8	0.0
Clindamycin	0.06	>2	80.6	19.2
Levofloxacin	0.25	>4	62.2	37.7
Linezolid	1	2	100.0	0.0
Oxacillin	1	>2	55.9	44.1
Teicoplanin	0.5	0.5	100.0	0.0
Tigecycline	0.12	0.12	99.9 ^b	
TMP-SMX	≤0.5	≤0.5	98.5	1.5
Vancomycin	1	1	100.0	0.0

TMP-SMX, trimethoprim-sulfamethoxazole.

^a MIC₅₀ and MIC₉₀ values in mg/L; criteria as published by CLSI (2019).

^b US FDA breakpoints were applied.

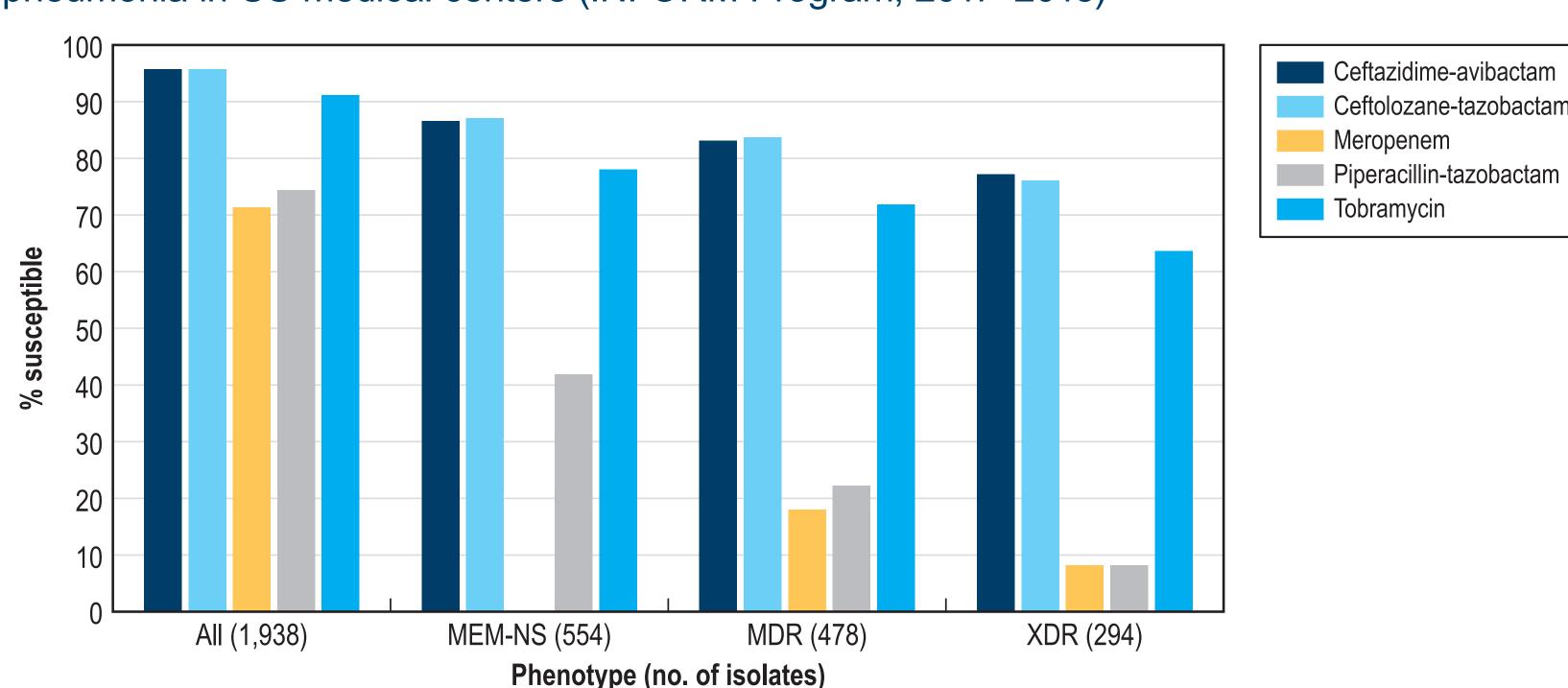


Figure 2 Antimicrobial susceptibility of *P. aeruginosa* isolated from patients hospitalized with pneumonia in US medical centers (INFORM Program, 2017–2018)

Abbreviations: MEM-NS, meropenem-nonsusceptible; MDR, multidrug-resistant; XDR, extensively drug-resistant.

Helio S. Sader, Mariana Castanheira, Cecilia G. Carvalhaes, Robert K. Flamm

- The most active agents against *Enterobacterales* were ceftazidime-avibactam (99.9%S), amikacin (98.7%S), meropenem (97.7%S), and tigecycline (94.8%S), but only ceftazidime-avibactam and tigecycline retained good activity (≥90%S) against CRE (96.3% and 92.6%S, respectively; Figure 3)
- The most active agents against MDR Enterobacterales were ceftazidime-avibactam (99.3%S) and amikacin (90.4%S), whereas ceftolozane-tazobactam and meropenem were active against only 62.6% and 81.1% of these organisms, respectively (Figure 3)
- Only 2 Enterobacterales isolates (0.1%) were ceftazidime-avibactam resistant; 2 S. marcescens isolates from New York, NY, 1 with KPC-3, OXA-10, OXA-9, SRT-like, TEM-1, and alterations in *ompC* and *ompF* isolated in 2017 and 1 with NDM-1 and alterations in *ompC* isolated in 2018
- Among ESBL-producing *Enterobacterales* (excluding carbapenemase-producing) isolates, susceptibility to ceftazidime-avibactam, ceftolozane-tazobactam, and meropenem was 100.0%, 85.5%, and 99.6%, respectively (Table 3 and Figure 3)
- The most common ESBLs found among Enterobacterales isolates were CTX-M-15 (62.0% of ESBL), OXA-1/OXA-30 (37.6%), CTX-M-27 (11.4%), and SHV-12 (8.9%); 49 of 51 (96.1%) carbapenemase-producing Enterobacterales displayed a KPC-2 and/or a KPC3 (data not shown)

Table 2 Antimicrobial susceptibility of *P. aeruginosa* isolates from patients hospitalized with pneumonia in US medical centers

Antimicrobial	MIC ₅₀ ^a	MIC ₉₀ ^a	%S ^a	%Rª
P. aeruginosa (1,938)				
Ceftazidime-avibactam	2	8	95.7	4.3
Ceftolozane-tazobactam	1	2	95.7	2.8
Piperacillin-tazobactam	8	128	74.4	14.1
Meropenem	0.5	16	71.3	20.7
Ceftazidime	2	32	79.2	15.7
Cefepime	4	16	78.8	9.1
Levofloxacin	1	16	70.4	20.1
Gentamicin	2	16	78.7	11.4
Amikacin	4	16	92.5	4.1
Colistin	0.5	1	99.9	0.1
β-lactam-nonsusceptible <i>P. aeruginosa</i> (2	212) ^b			
Ceftazidime-avibactam	8	32	68.4	31.6
Ceftolozane-tazobactam	4	>32	70.3	20.9
Colistin	0.5	1	99.5	0.5
Levofloxacin	8	>16	27.0	60.7
Amikacin	8	>32	74.1	18.4
Tobramycin	2	>16	67.9	25.0

^a MIC₅₀ and MIC₀₀ values in mg/L; criteria as published by CLSI (2019).

^b β-lactam-nonsusceptible defined as nonsusceptible to ceftazidime, cefepime, meropenem, and piperacillin-tazobactam.

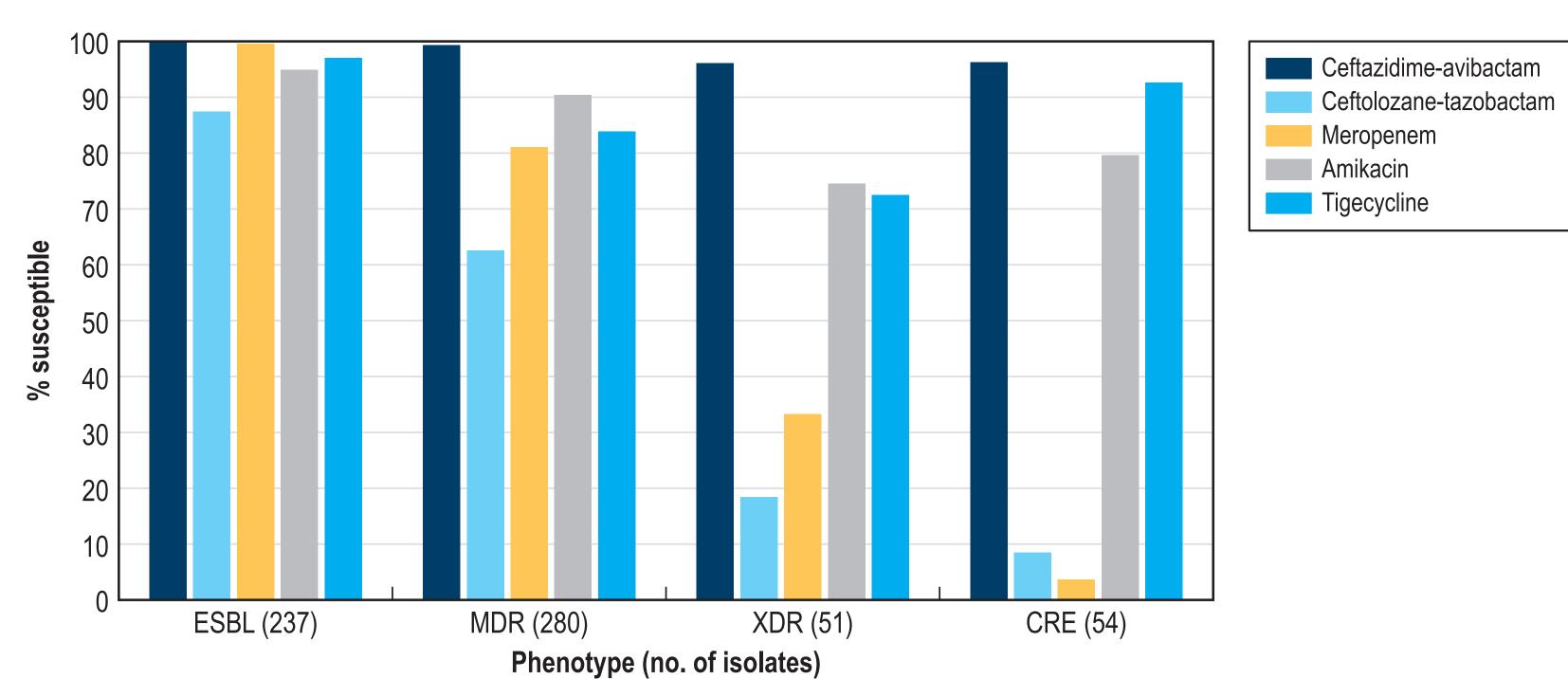


Figure 3 Antimicrobial susceptibility of *Enterobacterales* isolated from patients hospitalized with pneumonia in US medical centers (INFORM Program, 2017–2018)

Abbreviations: ESBL, extended-spectrum β -lactamases (excluding carbapenemase-producing strains); MDR, multidrug-resistant; XDR, extensively drug-resistant, CRE, carbapenem-resistant Enterobacterales.

Contact Information: Helio S. Sader, MD, PhD JMI Laboratories 345 Beaver Kreek Centre, Suite A North Liberty, IA 52317 Phone: (319) 665-3370 Fax: (319) 665-3371 Email: helio-sader@jmilabs.com

Table 3 Antimicrobial susceptibility of *Enterobacterales* isolates from patients hospitalized with pneumonia in US medical centers

Antimicrobial	MIC ₅₀ ^a	MIC ₉₀ a	%S ^a	%R ^a
Enterobacterales (2,370)				
Ceftazidime-avibactam	0.12	0.5	99.9	0.1
Ceftolozane-tazobactam	0.25	2	91.0	6.8
Piperacillin-tazobactam	2	64	87.0	7.0
Meropenem	0.03	0.06	97.7	2.0
Ceftriaxone	0.12	>8	76.4	21.6
Ceftazidime	0.25	32	82.0	16.3
Cefepime	≤0.12	8	86.9	9.3
Levofloxacin	0.06	16	79.5	17.6
Tigecycline	0.5	2	94.8 ^b	0.6 ^b
Gentamicin	0.5	2	91.7	7.4
Amikacin	2	4	98.7	0.2
Colistin	0.12	>8		
ESBL-producing Enterobacterales (excluding o	carbapenemases; 237)	'		
Ceftazidime-avibactam	0.25	0.5	100.0	0.0
Ceftolozane-tazobactam	0.5	8	85.5	12.3
Piperacillin-tazobactam	8	64	80.6	9.3
Meropenem	0.03	0.06	99.6	0.4
Ceftriaxone	>8	>8	1.3	96.6
Ceftazidime	32	>32	17.3	73.0
Cefepime	>16	>16	10.5	70.0
Levofloxacin	8	>16	24.5	67.9
Tigecycline	0.25	1	97.0 ^b	0.0 ^b
Gentamicin	1	>16	59.5	37.6
Amikacin	2	8	94.9	0.4
Colistin	0.12	0.25	98.2 ^c	

ESBL, extended-spectrum β -lactamase.

^a MIC_{50} and MIC_{90} values in mg/L; criteria as published by CLSI (2019).

^b US FDA breakpoints were applied. ^c Percentage of wild type based on ECV (CLSI M100, 2019).

CONCLUSIONS

- Ceftazidime-avibactam demonstrated potent activity against a large US collection of contemporary Enterobacterales (n=2,370) and *P. aeruginosa* (n=1,938) isolates from patients with pneumonia, including organisms resistant to most currently available agents, such as CRE and meropenem-nonsusceptible *P. aeruginosa*
- Ceftazidime-avibactam and ceftolozane-tazobactam showed similar coverage (%S) against *P. aeruginosa* (95.7%S), including against MDR (83.1%S vs. 83.7%S) and XDR (77.2%S vs. 76.1%S) isolates, respectively
- Ceftolozane-tazobactam was less active than ceftazidime-avibactam against Enterobacterales in general and exhibited limited activity against resistant subsets
- Ceftazidime-avibactam represents a valuable option for treating patients hospitalized with pneumonia caused by gram-negative organisms in US medical centers

ACKNOWLEDGEMENTS

The authors would like to thank all participants of the International Network for Optimal Resistance Monitoring (INFORM) Program for providing bacterial

This study was supported by Allergan. Allergan was involved in the design and decision to present these results, and JMI Laboratories received compensation for services related to preparing the poster. Allergan had no involvement in the collection, analysis, or interpretation of data.

REFERENCES

- 1. AVYCAZ[®] (2016). AVYCAZ[®] (ceftazidime-avibactam). Allergan USA, Inc. Available at https://www.allergan.com/assets/pdf/avycaz pi. Castanheira M, Mendes RE, Jones RN, Sader HS (2016). Changes in the frequencies of beta-lactamase genes among Enterobacteriaceae isolates in U.S. Hospitals, 2012 to 2014: Activity of ceftazidime-avibactam tested against beta-Lactamase-producing isolates. Antimicrob Agents Chemother 60: 4770-4777.
- Clinical and Laboratory Standards Institute (2018). M100Ed28E. Performance standards for antimicrobial susceptibility testing: 28th informational supplement. Wayne, PA: CLSI.
- 4. Clinical and Laboratory Standards Institute (2018). M07Ed11E. Methods for dilution antimicrobial susceptibility tests for bacteria that grow aerobically; approved standard: eleventh edition. Wayne, PA: CLSI.
- . Magiorakos AP, Srinivasan A, Carey RB, et al. (2012). Multidrug-resistant, extensively drug-resistant and pandrug-resistant bacteria: an international expert proposal for interim standard definitions for acquired resistance. Clin Microbiol Infect 18: 268-281
- Sader HS, Castanheira M, Mendes RE, Flamm RK (2018) Frequency and antimicrobial susceptibility of Gram-negative bacteria isolated from patients with pneumonia hospitalized in ICUs of US medical centres (2015–17). J Antimicrob Chemother 73:3053-9



To obtain a PDF of this poster: Scan the QR code

Visit http://www.allergancongressposters.com/597180

Charges may apply. No personal information is stored.