In Vitro Activity of Cefiderocol and Comparator Agents against Molecularly Characterized Carbapenem-resistant **Enterobacterales Clinical Isolates Causing Infection** in United States Hospitals (2020)

R.E. Mendes¹, TB Doyle¹, D. Shortridge¹, H.S. Sader¹, J.M. Streit¹, M. Castanheira¹ ¹JMI Laboratories, North Liberty, Iowa, USA

Introduction

- Cefiderocol represents a new addition to the antimicrobial armamentarium for the treatment of serious infections caused by Gram-negative bacteria.
- This molecule is a novel injectable siderophore cephalosporin, which hijacks the bacterial iron transport machinery to facilitate cell entry and achieve high periplasmic concentrations.
- This siderophore cephalosporin has broad *in vitro* activity against Gram-negative bacteria, including multidrug resistant (MDR) organisms like carbapenem resistant Enterobacterales (CRE), carbapenem resistant Pseudomonas aeruginosa and Acinetobacter baumannii.
- In addition, cefiderocol remains stable to hydrolysis in the presence of serine β -lactamases (ESBLs, KPC and OXA-type carbapenemases) and metallo-β-lactamases.
- Cefiderocol was approved by the Food and Drug Administration (FDA) for the treatment of complicated urinary tract infections and nosocomial pneumonia based on clinical trials demonstrating noninferiority to comparator agents.
- The activities of cefiderocol and comparator agents were analyzed against Enterobacterales, including molecularly characterized CRE, as a part of the SENTRY Antimicrobial Surveillance Program for 2020 in the USA.

Materials and Methods

Bacterial organisms

- This study comprises a collection of 4,043 *Enterobacterales* collected from clinical specimens from patients hospitalized in 31 medical centers in 9 US Census Divisions. Only consecutive isolates (1 per patient infection episode) responsible for documented infections according to local criteria were included.
- Bacterial identification was confirmed by standard algorithms supported by MALDI-TOF (Bruker Daltonics, Bremen, Germany).

Susceptibility testing

- Isolates were tested for susceptibility by broth microdilution following the Clinical and Laboratory Standards Institute (CLSI) M07 (2018) guidelines.
- Frozen-form broth microdilution panels were manufactured by JMI Laboratories (North Liberty, IA, USA) and contained cation-adjusted Mueller-Hinton broth (CA-MHB) for comparator agents. Cefiderocol susceptibility testing used broth microdilution panels containing iron-depleted media as per the CLSI guidelines.
- Quality assurance was performed by sterility checks, colony counts, and testing CLSIrecommended quality control reference strains. MIC interpretations were performed using CLSI breakpoints for comparators and FDA/CLSI breakpoints for cefiderocol $(\leq 4/8 \geq 16 \mu g/mL$ for susceptible, intermediate, and resistant). Imipenem-relebactam MIC interpretations used FDA breakpoints.
- Isolates displaying MIC values $\geq 4 \mu g/mL$ for imipenem (excluded for *Proteus mirabilis*, *P. penneri* and indole-positive *Proteus*) or meropenem were subjected to genome sequencing and screening of β -lactamase genes.

Screening of β-lactamase genes

- Selected isolates had total genomic DNA extracted by the fully automated Thermo Scientific[™] KingFisher[™] Flex Magnetic Particle Processor (Cleveland, OH, USA), which was used as input material for library construction.
- DNA libraries were prepared using the Nextera[™] library construction protocol (Illumina, San Diego, CA, USA) following the manufacturer's instructions and were sequenced on MiSeq Sequencer platforms at JMI Laboratories.
- FASTQ format sequencing files for each sample set were assembled independently using de novo assembler SPAdes 3.11.0. An in-house software was applied to align the assembled sequences against a comprehensive in-house database containing known β-lactamase genes. This database was originally based on the NCBI Bacterial Antimicrobial Resistance Reference Gene Database (https://www.ncbi.nlm.nih.gov /bioproject/PRJNA313047).

Results

- A total of 36 (0.9%) CRE isolates were detected, which were represented by 8 Enterobacterales species and most isolates carried bla_{KPC} (75.0%; 27/36) (Table 1).
- A small number of *Enterobacterales* (11.1%; 4/36) carried carbapenemase genes other than bla_{KPC} , as follows: 1 each of $bla_{\text{NDM-1}}$, $bla_{\text{NDM-5}}$, $bla_{\text{OXA-232}}$, and $bla_{\text{SME-2}}$. – Five (13.9%) Enterobacterales isolates did not carry any known carbapenemases.
- Cefiderocol inhibited all but 8 *Enterobacterales* at $\leq 4 \mu g/mL$ (99.8% susceptible) (Table 1 and Figure 1).
- A single isolate (non-CRE) showed a resistant phenotype to cefiderocol (MIC, 32 µg/mL).
- Among CRE isolates, all but 1 K. pneumoniae were inhibited by cefiderocol at $\leq 4 \,\mu g/mL (97.2\% \, susceptible).$
- This single CRE isolate showed a cefiderocol MIC of 8 µg/mL and carried CTX-M-15, OXA-1, SHV-168, and TEM-1.
- In general, cefiderocol, imipenem-relebactam, meropenem-vaborbactam, ceftazidimeavibactam, piperacillin-tazobactam and carbapenems were active (92.2% susceptible) against non-CRE isolates (Table 2).
- Cefiderocol (MIC_{50/90}, 0.5/4 µg/mL; 97.2% susceptible) and ceftazidime-avibactam (MIC_{50/90}, 1/8 µg/mL; 94.4% susceptible) were the most active agents against CRE (Table 2)
- All newest agents (i.e. cefiderocol, imipenem-relebactam, meropenemvaborbactam, ceftazidime-avibactam) were active (100% susceptible) against the KPC subset.
- Finally, cefiderocol (MIC, 0.5-8 µg/mL) was the most active agent against noncarbapenemase CRE and CRE isolates carrying genes other than bla_{kpc} (Table 2).

Conclusions

- The cefiderocol activity was potent and consistent across *Enterobacterales* with different phenotypes and genotypes.
- Cefiderocol also demonstrated an in vitro activity advantage against noncarbapenemase CRE and isolates carrying genes other than *bla*_{KPC}, where approved β -lactam/ β -lactamase inhibitor combinations showed limited activity.
- These data confirm cefiderocol as an important option for the treatment of serious infections caused by Enterobacterales and resistant subsets in patients hospitalized in US medical centers.

Acknowledgements

This research and poster presentation were sponsored by Shionogi & Co., LTD.

References

CLSI. M07Ed11. Methods for dilution antimicrobial susceptibility tests for bacteria that grow aerobically; approved standard: Eleventh Edition. Wayne, PA, Clinical and Laboratory Standards Institute, 2018.

CLSI. M100Ed31. Performance standards for antimcirobial susceptibility testing: 31st Informational Supplement. Wayne, PA, Clinical and Laboratory Standards Institute, 2021.

FDA Susceptibility Test Interpretive Criteria: https://www.fda.gov/drugs/development -resources/antibacterial-susceptibility-test-interpretive-criteria. Accessed August, 2021. Figure 1. Cumulative MIC distribution of cefiderocol against CRE and non-CRE subsets from the USA



Table 1. MIC distribution of cefiderocol obtained against *Enterobacterales* and resistant subsets from the USA

Organism/		No. and cumulative % of isolates inhibited at MIC (µg/mL) of:														
Organism group (no. of isolates)	≤0.004	0.008	0.015	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	MIC ₅₀	MIC ₉₀
Enterobacterales (4,053)	131 (3.2)	337 (11.5)	569 (25.6)	585 (40.0)	649 (56.0)	735 (74.2)	565 (88.1)	253 (94.3)	134 (97.7)	63 (99.2)	24 (99.8)	7 (>99.9)	0 (>99.9)	1 (100.0)	0.06	0.5
Non-CRE ^a (4,017)	131 (3.3)	336 (11.6)	569 (25.8)	582 (40.3)	648 (56.4)	732 (74.6)	561 (88.6)	243 (94.6)	129 (97.9)	59 (99.3)	20 (99.8)	6 (>99.9)	0 (>99.9)	1 (100.0)	0.06	0.5
CRE ^a (36)		1 (2.8)	0 (2.8)	3 (11.1)	1 (13.9)	3 (22.2)	4 (33.3)	10 (61.1)	5 (75.0)	4 (86.1)	4 (97.2)	1 (100.0)			0.5	4
KPC ^b (27)		1 (3.7)	0 (3.7)	3 (14.8)	1 (18.5)	3 (29.6)	4 (44.4)	6 (66.7)	5 (85.2)	3 (96.3)	1 (100.0)				0.5	2
Other ^c (4)								2 (50.0)	0 (50.0)	0 (50.0)	2 (100.0)				-	-
No-carbapenemasesd (5)								2 (40.0)	0 (40.0)	1 (60.0)	1 (80.0)	1 (100.0)			-	-

^a Non-CRE, includes isolates with imipenem and/or meropenem MIC $\leq 2 \mu g/mL$; CRE, defined as isolates with imipenem (excluded for *P. mirabilis, P. penneri* and indole-positive *Proteus*) and/or meropenem MIC $\geq 4 \mu g/mL$ and includes *Citrobacter* freundii (3), Enterobacter cloacae (3), Escherichia coli (2), Klebsiella aerogenes (2), K. oxytoca (4), K. pneumoniae (18), Serratia marcescens (2), Raoultella ornithinolytica (2)

^b Includes 12 isolates carrying *bla*_{KPC-2} and 15 *bla*_{KPC-3} ^c Includes 1 isolate each of *bla*_{NDM-1} (MIC, 4 μg/mL), *bla*_{NDM-5} (MIC, 4 μg/mL), *bla*_{OXA-232} (MIC, 0.5 μg/mL), and *bla*_{SME-2} (MIC, 0.5 μg/mL) ^d Includes 5 CRE with no known carbapenemase genes detected

Table 2. Antimicrobial activity of cefiderocol tested against *Enterobacterales* and resistant subsets from the USA

	MIC (µg/mL)			CLSI ^a			Autimic robiol octout		MIC (µĮ	g/mL)	CLSI ^a				
Antimicropial agent	50 %	90%	Range	% S	%	% R	Antimicropial agent	50 %	90%	Range	% S	%	% R		
Non-CRE (4,017)						Ceftazidime-avibactam	1	8	≤0.015 to >32	94.4		5.6			
Cefiderocol ^b	0.06	0.5	≤0.004 to 32	99.8	0.1	<0.1	Meropenem	8	>32	1 to >32	2.8	13.9	83.3		
Imipenem-relebactam b	0.12	0.5	≤0.03 to 8	99.9	0.1	0.0	Imipenem	8	>8	4 to >8	0.0	0.0	100.0		
Meropenem-vaborbactam	0.03	0.06	≤0.015 to 0.5	100.0	0.0	0.0	KPC (27)								
Ceftazidime-avibactam	0.12	0.25	≤0.015 to 8	100.0		0.0	Cefiderocol ^b	0.5	2	0.008 to 4	100.0	0.0	0.0		
Piperacillin-tazobactam	2	16	≤0.06 to >128	92.2	3.9	3.8	Imipenem-relebactam ^b	0.12	0.25	0.06 to 1	100.0	0.0	0.0		
Aztreonam	0.12	>16	≤0.03 to >16	84.5	1.7	13.7	Meropenem-vaborbactam	0.03	1	≤0.015 to 2	100.0	0.0	0.0		
Ceftriaxone	≤0.06	>8	≤0.06 to >8	81.7	0.7	17.5	Ceftazidime-avibactam	0.5	2	≤0.015 to 4	100.0	—	0.0		
Ceftazidime	0.25	16	0.03 to >32	85.2	2.0	12.7	Meropenem	8	>32	1 to >32	3.7	14.8	81.5		
Cefepime	0.06	8	≤0.03 to >32	88.3	2.4	9.3	Imipenem	8	>8	4 to >8	0.0	0.0	100.0		
Meropenem	0.03	0.06	≤0.015 to 2	99.9	0.1	0.0	CRE Non-KPC (9)								
Imipenem	≤0.12	1	≤0.12 to 8	94.4	4.7	0.8	Cefiderocol ^b	2		0.5 to 8	88.9	11.1	0.0		
Ciprofloxacin	0.03	>4	≤0.008 to >4	77.9	3.4	18.7	Imipenem-relebactam ^b	4		0.12 to >8	22.2	22.2	55.6		
Levofloxacin	0.06	8	≤0.015 to >32	80.3	3.2	16.5	Meropenem-vaborbactam	8		0.03 to >8	33.3	33.3	33.3		
Amikacin	2	4	≤0.25 to >32	99.6	0.3	0.1	Ceftazidime-avibactam	2		0.25 to >32	77.8		22.2		
Gentamicin	0.5	2	≤0.12 to >16	91.4	0.7	7.9	Meropenem	16		2 to >32	0.0	11.1	88.9		
CRE (36)							Imipenem	>8		4 to >8	0.0	0.0	100.0		
Cefiderocol ^b	0.5	4	0.008 to 8	97.2	2.8	0.0	CRE, carbapenem resistant Enterobacterales; KPC, Klebsiella pneumoniae carbapenemase.								
Imipenem-relebactam ^b	0.12	4	0.06 to >8	80.6	5.6	13.9	 ^a Criteria as published by CLSI (2021) unless otherwise indicated; "—", breakpoint not available. ^b Using EDA breakpoints. Iminenem-relebactam breakpoints applied to organisms other than Morganolla sport. Protocome and the organisms of the sport of the organisms. 								
Meropenem-vaborbactam	0.03	8	≤0.015 to >8	83.3	8.3	8.3	spp., and Providencia spp.								

Mendes RE, Jones RN, Woosley LN, Cattoir V, Castanheira M (2019). Application of nextgeneration sequencing for characterization of surveillance and clinical trial isolates: Analysis of the distribution of β -lactamase resistance genes and lineage background in the United States. Open Forum Infect Dis 6: S69-S78.

Ong'uti S, Czech M, Robilotti E, Holubar M (2021). Cefiderocol: A new cephalosporin stratagem against multidrug resistant Gram-negative bacteria. Clin Infect Dis. In press.

Syed YY (2021). Cefiderocol: A review in serious Gram-negative bacterial infections. Drugs. 24: 1–13.

Contact

R.E. Mendes JMI Laboratories 345 Beaver Kreek Centre, Suite A North Liberty, IA 52317

Phone: 319-665-3370 Fax: (319) 665-3371 Email: rodrigo-mendes@jmilabs.com