# Activity of Investigational Polymyxin-B-Like Compound (SPR206) against Set of Gram-Negative Bacilli Responsible for Human Infections

SJR Arends<sup>1</sup>, PR Rhomberg<sup>1</sup>, T Lister<sup>2</sup>, N Cotroneo,<sup>2</sup> A Rubio<sup>2</sup>, RK Flamm<sup>1</sup>, RE Mendes<sup>1</sup>

<sup>1</sup>JMI Laboratories, North Liberty, Iowa, USA; <sup>2</sup>Spero Therapeutics, Cambridge, Massachusetts, USA

#### Introduction

- Gram-negative bacilli (GNB) are opportunistic organisms that have emerged as important healthcare-associated pathogens, mainly in the immunocompromised patient population
- GNB producing extended-spectrum  $\beta$ -lactamase (ESBL) and/or carbapenemase enzymes that show resistance to many antibiotics have been steadily increasing to alarming levels in hospital and community settings
- SPR206 is a polymyxin derivative compound being clinically developed for treating serious infections caused by GNB (Figure 1)
- This study evaluated the *in vitro* potency of SPR206 and compared its potency to those of polymyxin-B and colistin against a current collection of GNB, including carbapenem-resistant organisms

#### Materials and Methods

#### Bacterial isolates

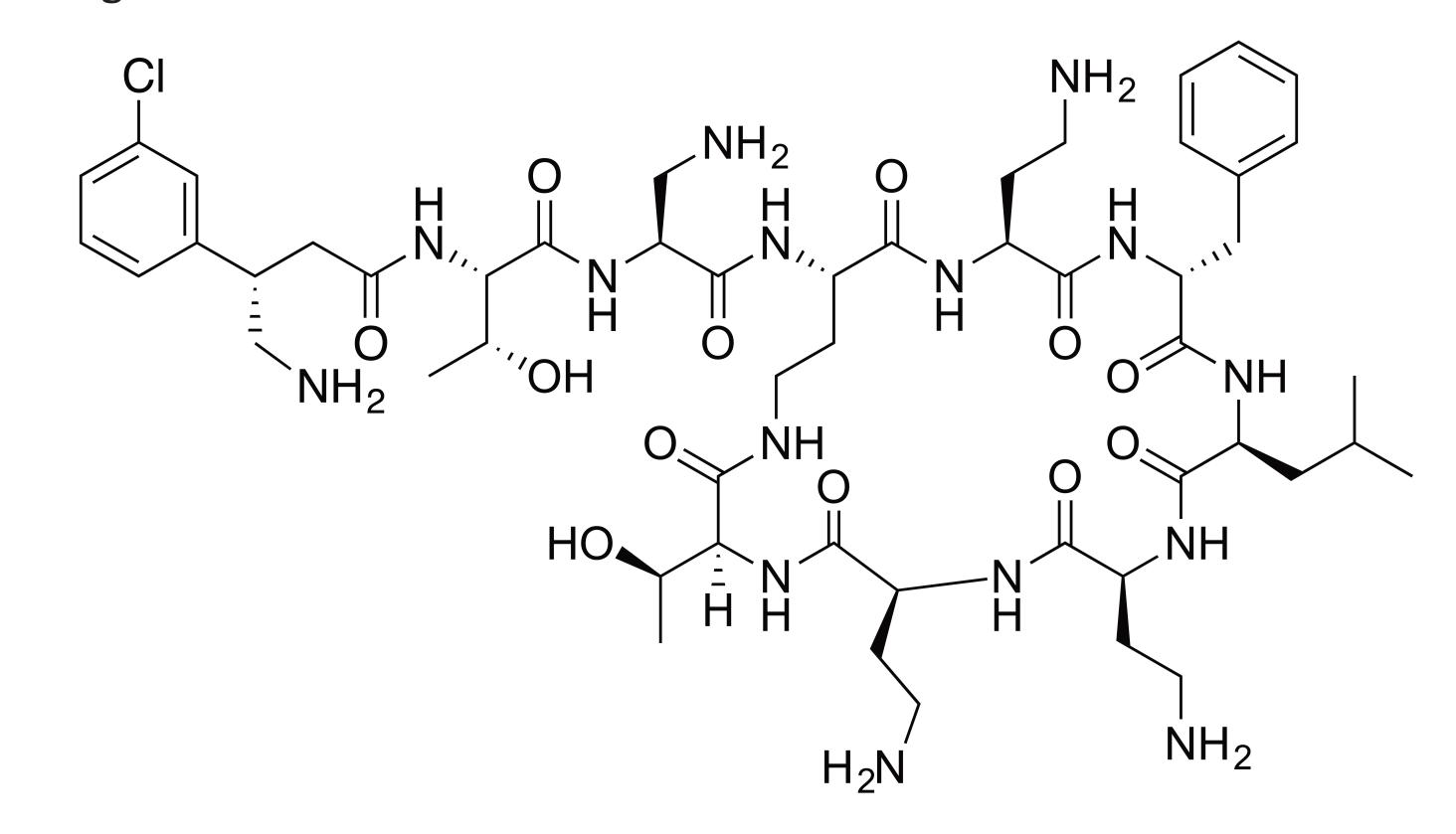
- A total of 930 recent clinical *Enterobacteriaceae* isolates (2016–2017) were randomly selected through the SENTRY Antimicrobial Surveillance Program from 40 medical centers in 21 European (EUR) nations (n=339), 82 medical centers located in North America (NA; n=486), 17 medical centers from 9 Asia-Pacific (APAC) nations (n=68), and 16 medical centers from 11 Latin American (LATAM) nations (n=90) (Table 1)
- Isolates were responsible for bloodstream (30%), urinary tract (26%), pneumonia (20%), skin and skin structure (15%), and other infections (9%)
- An additional set of 53 meropenem-nonsusceptible isolates was also included to investigate the drug activities against carbapenem nonsusceptible organisms (Table 2)
- Isolates were determined to be clinically significant based on local guidelines and submitted to a central monitoring laboratory (JMI Laboratories, North Liberty, Iowa)
- monitoring laboratory (JMI Laboratories, North Liberty, Iowa)
   Bacterial isolate identification was confirmed by standard algorithms supported by matrix-assisted laser desorption ionization-time of flight mass spectrometry (Bruker Daltonics, Bremen, Germany)

#### Antimicrobial susceptibility testing

- Isolates were tested for susceptibility by broth microdilution following guidelines in the Clinical and Laboratory Standards Institute (CLSI) M07 (2018) document against the following antimicrobial agents (ranges)
- SPR206 (0.015 16 mg/L)
- Colistin (0.06 64 mg/L)Polymyxin-B (0.06 64 mg/L)
- Ceftriaxone (0.12 8 mg/L)
- Meropenem (0.12 8 mg/L)
- JMI Laboratories manufactured the frozen-form reference 96-well panels used in this study with Starstead polystyrene panels (product #82.1582.001)

  Ouglity accurrence was performed by consumently testing CLSI recommended quality control reference.
- Quality assurance was performed by concurrently testing CLSI-recommended quality control reference strains (E. coli ATCC 25922, E. coli NCTC 13846, K. pneumonia ATCC 700603, and P. aeruginosa ATCC 27853)
- Breakpoint criteria for comparator agents were from the M100 CLSI (2018) and European Committee on Antimicrobial Susceptibility Testing (EUCAST; 2018) documents
- Isolates were tested for susceptibility by broth microdilution following guidelines in the CLSI M07 (2018) document against SPR206 and select comparator agents

#### Figure 1 Structure of SPR206



## Results

- Against *Enterobacteriaceae*, SPR206 (MIC<sub>50/90</sub>, 0.06/0.12 mg/L) was more potent than colistin and polymyxin-B (MIC<sub>50/90</sub>, 0.25/0.25 mg/L; Table 1, Figure 2)
- SPR206 inhibited 93.2% of all *Enterobacteriaceae* at  $\le$ 0.12 mg/L, while colistin and polymyxin-B inhibited 38.3% and 33.1%, respectively, at  $\le$ 0.12 mg/L (Table 1)
- SPR206 had an MIC $_{100}$  of  $\leq 2$  mg/L against Escherichia, Citrobacter, Salmonella, and Shigella species (Table 1 and data not shown)
- Ceftriaxone displayed a bimodal MIC distribution (MIC $_{50/90}$ ,  $\leq 0.12/>8$  mg/L) against all Enterobacteriaceae isolates and 77.4% were susceptible at the CLSI and EUCAST breakpoints of  $\leq 1$  mg/L
- Meropenem was very active (MIC $_{50/90}$ ,  $\le$ 0.12/ $\le$ 0.12 mg/L) against these isolates and 97.0%/97.2% were susceptible at the CLSI/EUCAST breakpoints, respectively (Table 1)
- Against a CRE challenge set, SPR206 (MIC $_{50/90}$ , 0.06/0.12 mg/L) showed MIC values 4-fold lower than colistin and polymyxin-B (MIC $_{50/90}$ , 0.25/0.5 mg/L; Table 2)
- Isolates included  $bla_{\rm KPC}$ ,  $bla_{\rm NDM}$ ,  $bla_{\rm VIM}$ , and  $bla_{\rm OXA-48}$  genotypes
- MIC results similar to the random selection set are seen in Table 1
- As expected, ceftriaxone (MIC $_{50/90}$ , >8/>8 mg/L) and meropenem (MIC $_{50/90}$ , >8/>8 mg/L) showed little activity against this challenge set (Table 2)
- SPR206 inhibited all randomly selected *P. aeruginosa* at  $\leq 2$  mg/L and showed MIC results (MIC<sub>50/90</sub>, 0.25/0.5 mg/L) 2-fold lower than colistin (MIC<sub>50/90</sub>, 0.5/1 mg/L) and polymyxin B (MIC<sub>50/90</sub>, 0.5/1 mg/L) (Table 1, Figure 3)
- Similar MIC results for the respective compounds were obtained against carbapenem-nonsusceptible *P. aeruginosa* compared with the randomly selected set (Table 2)
- Against Acinetobacter baumannii, SPR206 (MIC $_{50/90}$ , 0.12/0.25 mg/L) was 2- to 8-fold more potent than polymyxin-B (MIC $_{50/90}$ , 0.25/1–2 mg/L) and 4- to 32-fold more potent than colistin (MIC $_{50/90}$ , 0.5/4–8 mg/L) (Tables 1 and 2, Figure 4)
- In addition, SPR206 inhibited 95.7% of randomly selected Acinetobacter spp. or 93.1% of all tested Acinetobacter spp. at  $\leq 2$  mg/L
- SPR206 (MIC $_{50/90}$ , 0.25/4 mg/L) and polymyxin-B (MIC $_{50/90}$ , 0.5/4 mg/L) showed similar MIC values against S. maltophilia, and these compounds had MIC results 4- to 16-fold lower than collistin (MIC $_{50/90}$ , 4/16 mg/L) (Table 1)

## Conclusions

- Overall, SPR206 was highly potent against a contemporary collection of *Enterobacteriaceae* and non-fermentative GNB
- Based on MIC<sub>50/90</sub> results, SPR206 potency was consistently 2- to 4-fold greater than the potency of clinically available in-class comparator agents colistin and polymyxin-B
- Against a challenge set of isolates with increased carbapenem MIC values
   SPR206 MIC results were not adversely affected when compared with the MIC values obtained against randomly selected organisms
- SPR206 MIC values were consistently lower than colistin and polymyxin-B
- These *in vitro* results obtained for SPR206 warrant its further development as an option for treating gram-negative infections

# Acknowledgements

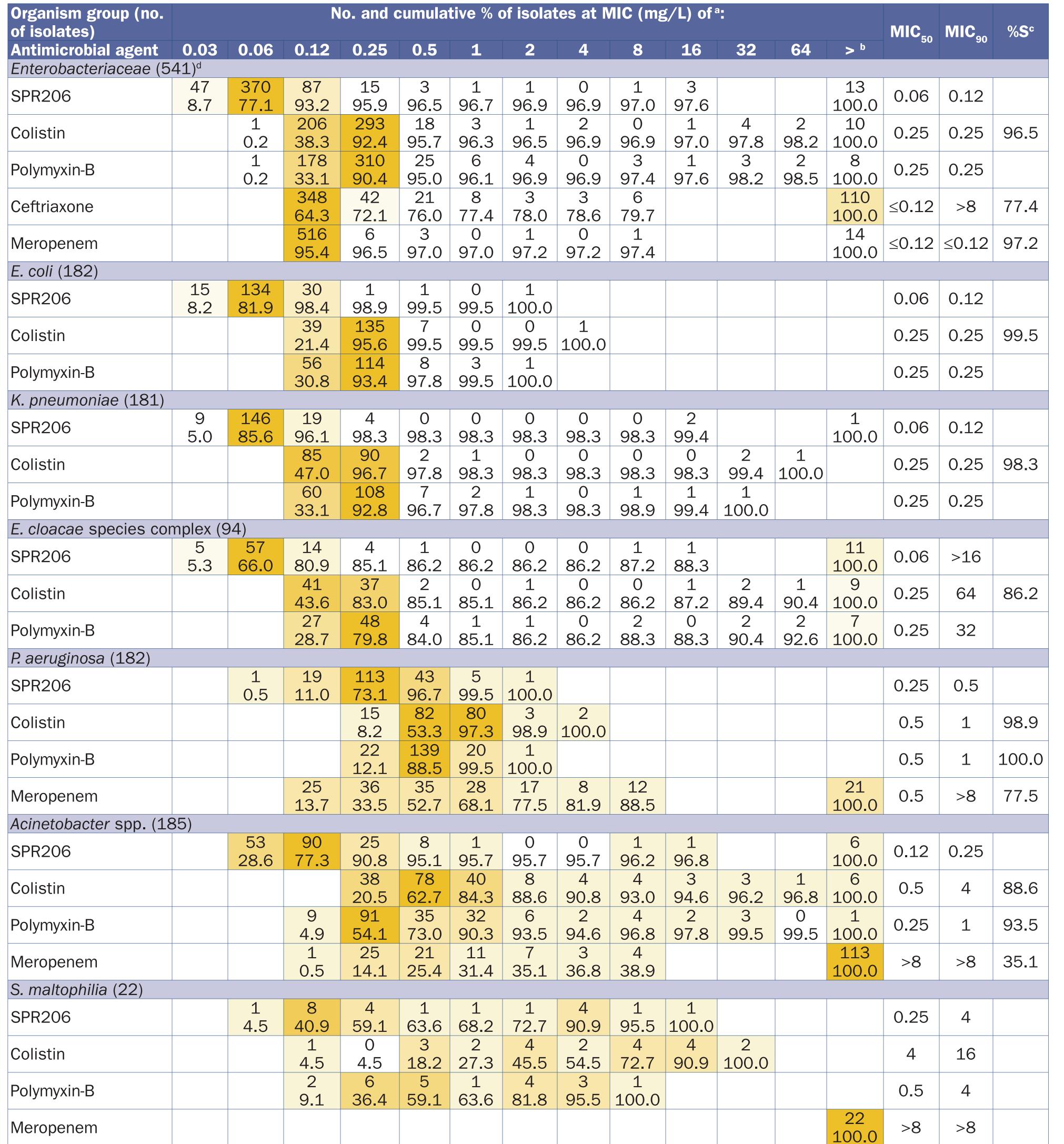
This study was supported by Spero Therapeutics. JMI Laboratories received compensation for services related to preparing this poster.

## References

- Clinical and Laboratory Standards Institute (2018). M100Ed28E. Performance standards for antimicrobial susceptibilities testing: 28th informational supplement. Wayne, PA: CLSI.
- Clinical and Laboratory Standards Institute (2018). M07Ed11E. Methods for dilution antimicrobial susceptibilty tests for bacteria that grow aerobically; approved standard eleventh edition. Wayne, PA: CLSI. EUCAST (2018). Breakpoint tables for interpretation of MIC's and zone diameters. Version 8.0, January 2018.
- Available at: http://www.eucast.org/fileadmin/src/media/PDFs/EUCAST\_files/Breakpoint\_tables/v\_8.0\_Breakpoint\_Tables.pdf. Accessed January 2018.

Vaara M (2013). Novel derivatives of polymyxins. J Antimicrob Chemother 68:1213–1219.

## Table 1 Antimicrobial activity of SPR206 and comparators tested against the main organisms and organism groups for non-challenge isolates



The intensity of shading is proportional to the % of tested isolates within each row that display the indicated MIC value.

Susceptible breakpoints were those from CLSI/EUCAST (2018).

and K. pneumoniae (9 isolates; 1  $bla_{KPC-2}$ , 3  $bla_{KPC-3}$ , 2  $bla_{NDM-1}$ , and 3  $bla_{OXA-48}$ ).

Susceptible breakpoints were those from CLSI/EUCAST (2018).
Organisms include: Citrobacter species (19), Enterobacter aerogenes (22), E. cloacae species complex (94), Escherichia coli (182), Klebsiella oxytoca (19), K. pneumoniae (181), Salmonella species (13), and Shigella species (11)

#### Figure 2 Comparison of colistin (A) or polymyxin-B (B) to SPR206 when tested against 573 Enterobacteriaceae isolates

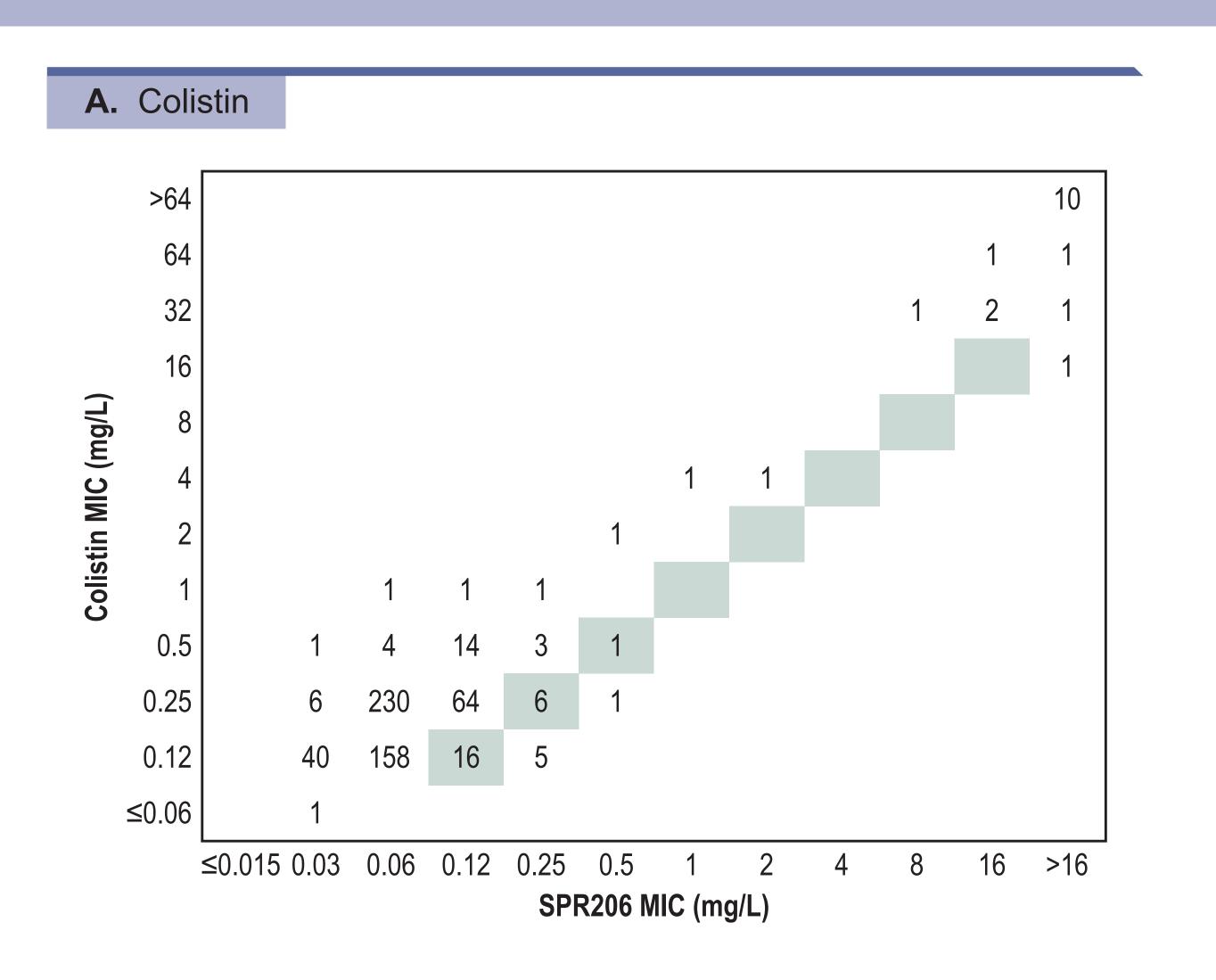
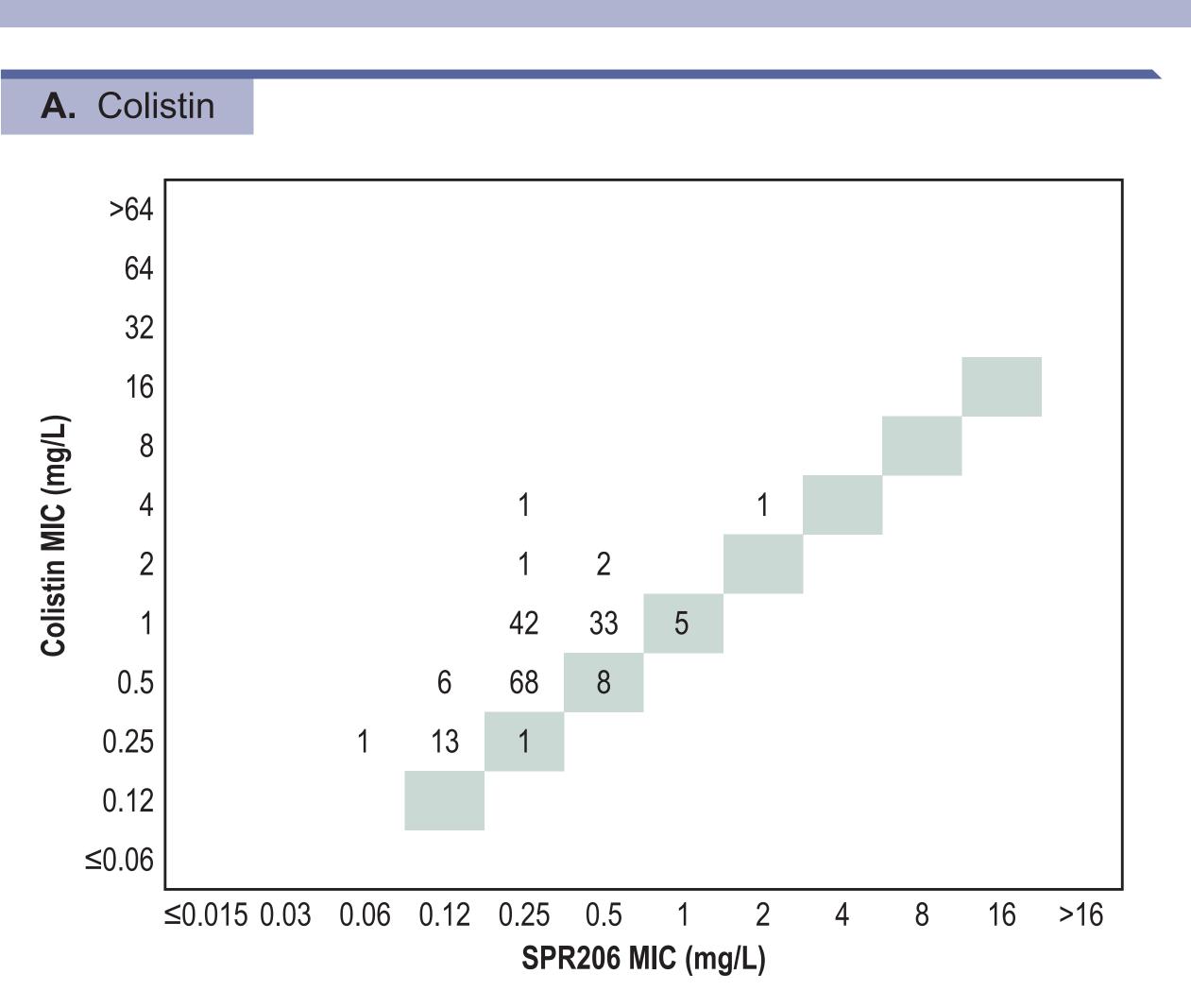


Figure 3 Comparison of colistin to SPR-206 when tested against 182 *Pseudomonas aeruginosa* isolates



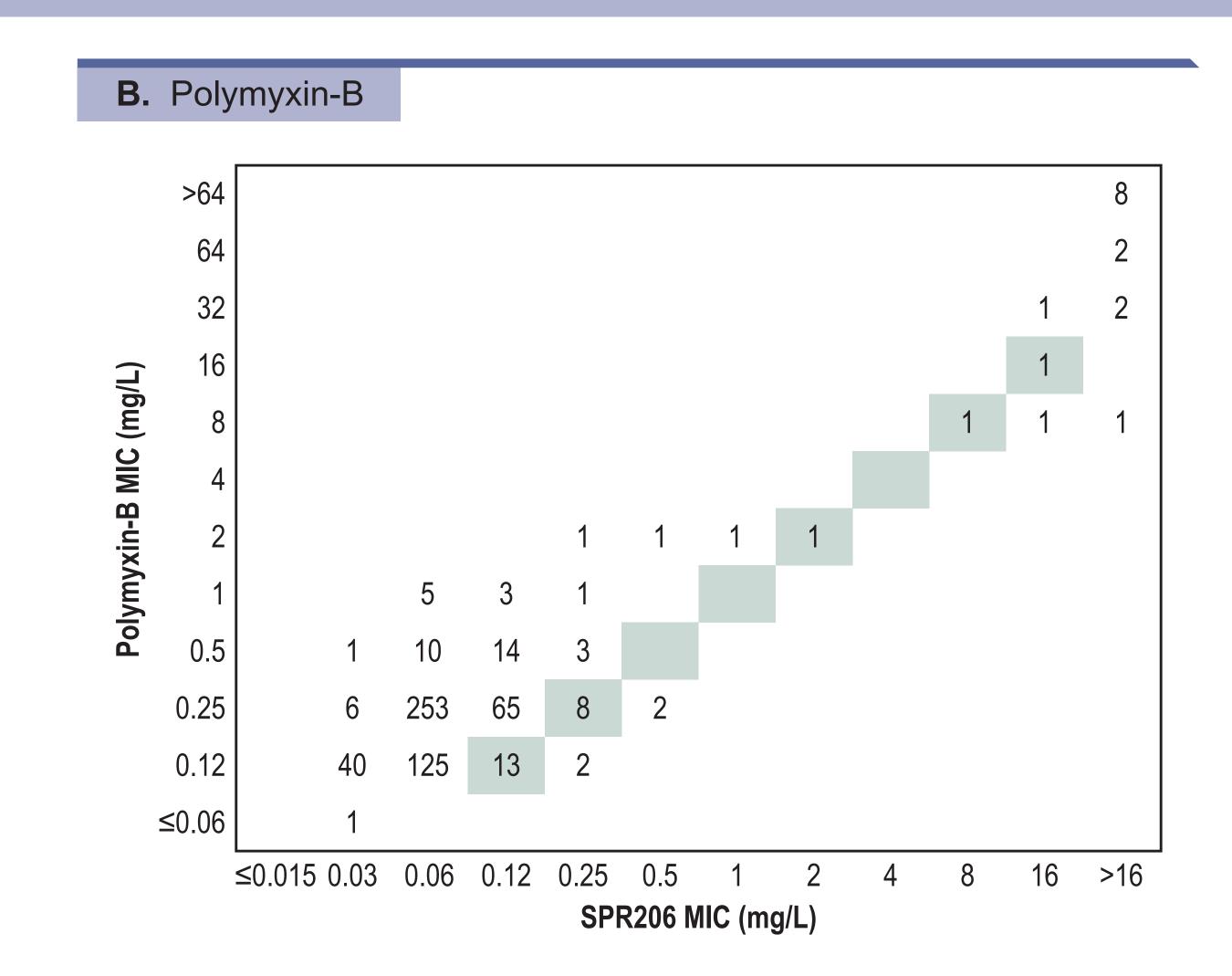
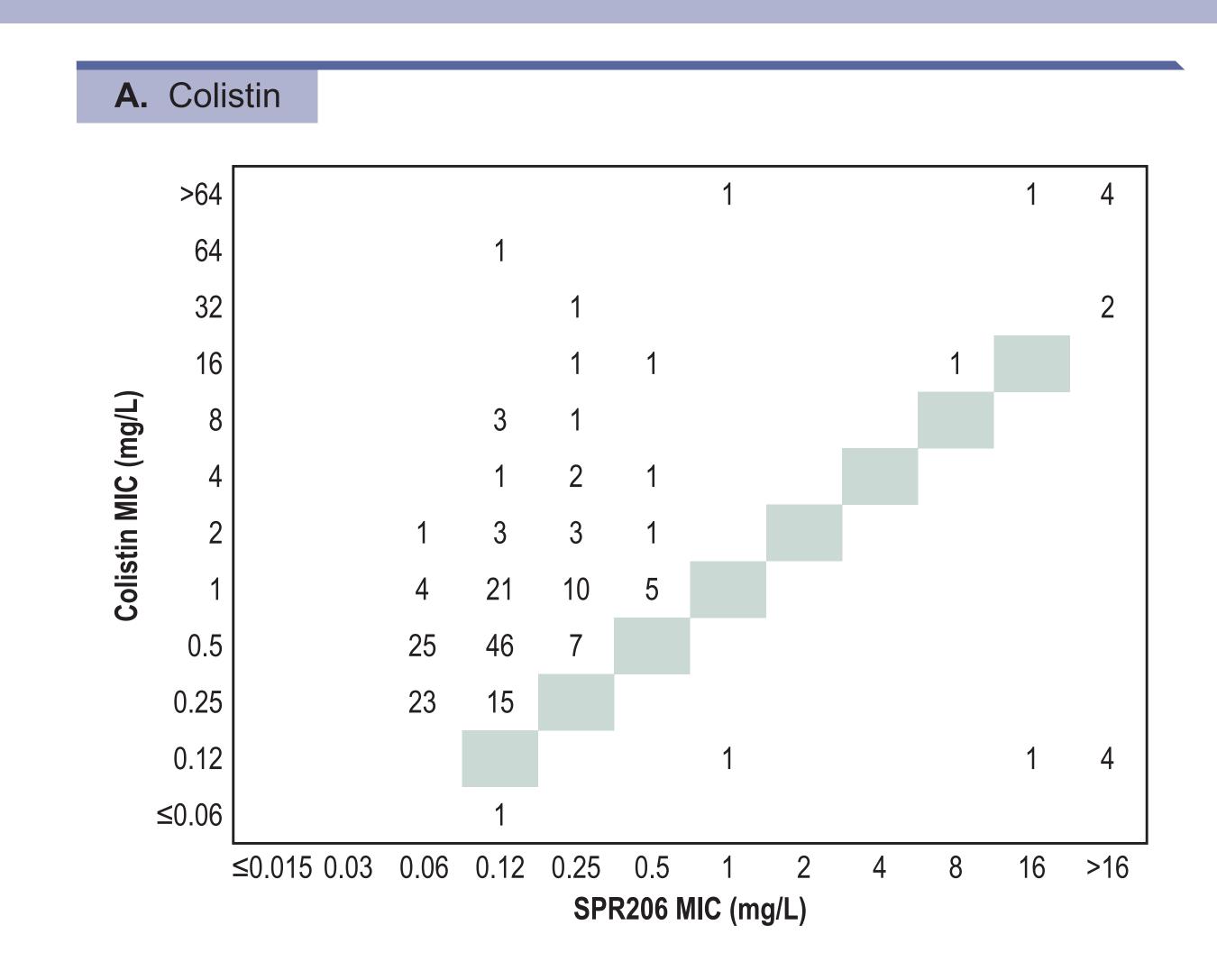
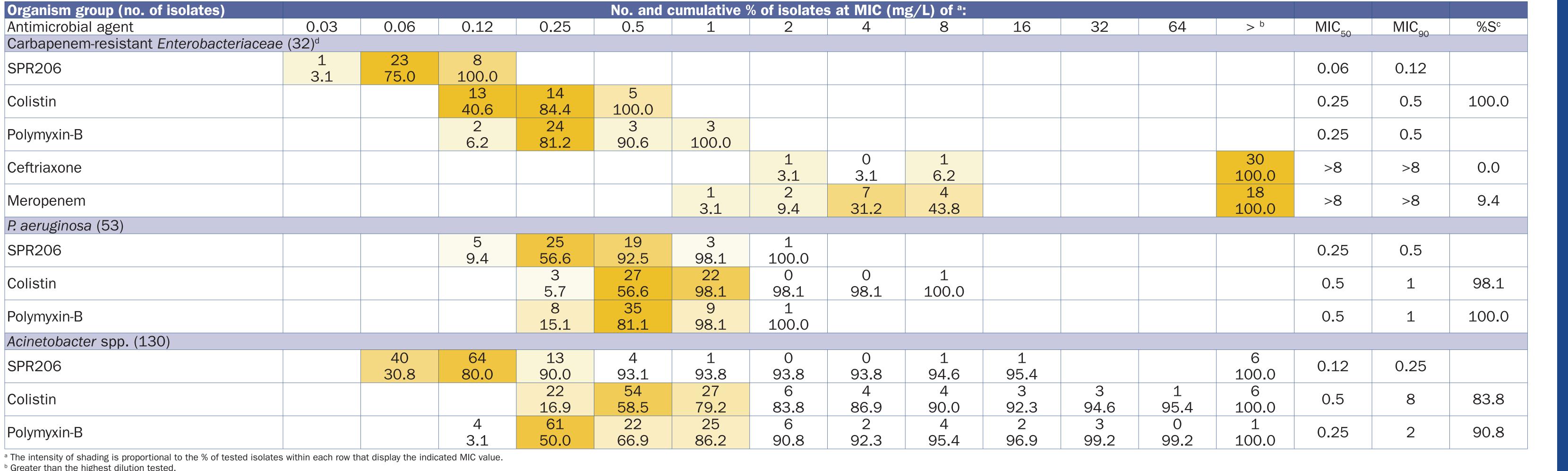


Figure 4 Comparison of colistin to SPR-206 when tested against 185 *Acinetobacter* spp. isolates



#### Table 2 Antimicrobial activity of SPR206 and comparators tested against meropenem-nonsusceptible gram-negative bacilli



<sup>d</sup> Includes Citrobacter freundii species complex (4 isolates; 1  $bla_{KPC-2}$ , 1  $bla_{KPC-2}$ , 1  $bla_{KPC-2}$ , 2  $bla_{KPC-2}$ , 2  $bla_{KPC-2}$ , 2  $bla_{KPC-2}$ , 1  $bla_{NDM-1}$ , 1  $bla_{NDM-1}$ , and 1  $bla_{NDM-1}$ , and 1  $bla_{NDM-1}$ , 2  $bla_{NDM-1}$ , 1  $bla_{NDM-1}$ , 1  $bla_{NDM-1}$ , 3  $bla_{NDM-1}$ , and 1  $bla_{NDM-1}$ , 1  $bla_{NDM-1}$ , 2  $bla_{NDM-1}$ , 2  $bla_{NDM-1}$ , 2  $bla_{NDM-1}$ , 3  $bla_{NDM-1}$ , 3  $bla_{NDM-1}$ , 3  $bla_{NDM-1}$ , 2  $bla_{NDM-1}$ , 3  $bla_{NDM-1}$ , 3  $bla_{NDM-1}$ , 3  $bla_{NDM-1}$ , 4  $bla_{NDM-1}$ , 5  $bla_{NDM-1}$ , 5  $bla_{NDM-1}$ , 6  $bla_{NDM-1}$ , 6  $bla_{NDM-1}$ , 8  $bla_{NDM-1}$ , 8  $bla_{NDM-1}$ , 8  $bla_{NDM-1}$ , 8  $bla_{NDM-1}$ , 9  $bla_{NDM-1}$ , 9  $bla_{NDM-1}$ , 9  $bla_{NDM-1}$ , 9  $bla_{NDM-1}$ , 1  $bla_{NDM-1}$ , 2  $bla_{NDM-1}$ , 1  $bla_{NDM-1}$ , 2  $bla_{NDM-1}$ , 3  $bla_{NDM-1}$ , 2  $bla_{NDM-1}$ , 3  $bla_{NDM-1}$ , 2  $bla_{NDM-1}$ , 3  $bla_{NDM-1}$ , 3  $bla_{NDM-1}$ , 3  $bla_{NDM-1}$ , 4  $bla_{NDM-1}$ , 5  $bla_{NDM-1}$ , 6  $bla_{NDM-1}$ , 6  $bla_{NDM-1}$ , 8  $bla_{NDM-1}$ , 8  $bla_{NDM-1}$ , 9  $bla_{NDM-1}$ , 9  $bla_{NDM-1}$ , 1  $bla_{NDM-1}$ , 1  $bla_{NDM-1}$ , 1  $bla_{NDM-1}$ , 1  $bla_{NDM-1}$ , 2  $bla_{NDM-1}$ , 1  $bla_{NDM-1}$ , 2  $bla_{NDM-1}$ , 3  $bla_{NDM-1}$ , 1  $bla_{NDM-1}$ , 2  $bla_{NDM-1}$ , 2  $bla_{NDM-1}$ , 3  $bla_{NDM-1}$ , 2  $bla_{NDM-1}$ , 3  $bla_{NDM-1}$ , 3  $bla_{NDM-1}$ , 4  $bla_{NDM-1}$ , 4  $bla_{NDM-1}$ , 5  $bla_{NDM-1}$ , 6  $bla_{NDM-1}$ , 6  $bla_{NDM-1}$ , 8  $bla_{NDM-1}$ , 9  $bla_{NDM-1}$ , 1  $bla_{NDM-1}$ , 2  $bla_{NDM-1}$ , 1  $bla_{NDM-1}$ , 2  $bla_{NDM-1}$ , 2  $bla_{NDM-1}$ , 3  $bla_{NDM-1}$ , 3  $bla_{NDM-1}$ , 3  $bla_{NDM-1}$ , 1  $bla_{NDM-1}$ , 3  $bla_{NDM-1}$ 

## Contact

S.J. Ryan Arends, PhD
JMI Laboratories
345 Beaver Kreek Centre, Suite A
North Liberty, IA 52317
Phone: (319) 665-3370
Fax: (319) 665-3371

Email: ryan-arends@jmilabs.com

To obtain a PDF of this poster:

Scan the QR code or visit https://www
.jmilabs.com/data/posters/ASM-Microbe19SPR206-gram-negative-bacilli.pdf

Charges may apply. No personal information is stored.