## A National Epidemic of Multiple Metallo-B-Lactamase Clones (VIM-2, -5, -6, -11 and new VIM-18) M CASTANHEIRA, JM BELL, RN JONES, D MATHAI, JD TURNIDGE



**JMI** Laboratories North Liberty, IA, USA www.jmilabs.com 319.665.3370, fax 319.665.3371 ronald-jones@jmilabs.com

# High Carbapenem Resistance Among Pseudomonas aeruginosa from India: JMI Laboratories, North Liberty, IA, USA; Women's and Children's Hosp. Adelaide, Australia; Christian Medical College, Vellore, India

## AMENDED ABSTRACT

**Objectives:** We assessed the prevalence of metallo-B-lactamase (MBL)-encoding genes among carbapenem-resistant *P. aeruginosa* isolates recovered in India during 2006. In addition, class 1 integrons harbouring MBL genes were amplified and compared, and MBL-positive isolates were molecular typed to evaluate clonal dissemination.

Methods: A total of 282 *P. aeruginosa* were consecutively collected in 10 Indian medical centers and tested by CLSI broth microdilution methods. Isolates resistant to imipenem or meropenem (MIC,  $\geq 8$ mg/L) were screened for MBL-encoding genes by real-time PCR. Positive samples were tested with primers targeting the class 1 integron conserved structures anchoring in the MBL genes. Amplicons generated were sequenced on both strands and MBL isolates ribotyped for possible clonality.

**Results:** Among the 282 isolates, 96 (34%) showed carbapenem resistance and MBL genes were detected in 53 strains (19% of total; 55% of carbapenem-resistant). MBL-producing *P. aeruginosa* were detected in 9 of 10 hospitals. Five *blavim* genes were found, including a new variant named *bla*<sub>VIM-18</sub>. This new MßL gene showed a 12-bp deletion (position 428) when compared to  $bla_{VIM-2}$  and was carried as a single gene cassette in a class 1 integron. VIM-2 producing isolates were most common (38 strains) and were detected in 8 medical centers. VIM-6 was identified in 12 isolates from 4 sites. VIM-5, VIM-11 and VIM-18 were found only once. Two medical centers had 3 distinct MBL types and other 2 had 2 MBL types. Wide genetic diversity was noted among MBL-carrying *P. aeruginosa* with 21 and 10 ribotypes seen among VIM-2- and -6-producers, respectively. Seven clones were found in  $\geq 1$  participant hospital and 6 clones were noted within institutions.  $bla_{VIM-6}$ -carrying integrons of 3.5 and 5 Kb (5 and 7 isolates, respectively) were detected and 2 sites had both  $bla_{VIM-6}$ integron types.

				lonality	
Site	CARB-R <sup>a</sup> / MßL (%)	MßL types (No. of isolates)	Intra	Inter	No. of clones
Kolkatta	30/22	VIM-2 (3), -5 (1), 18 (1)	0	2	2
Indore	42/42	VIM-2 (3), -6 (5)	1 2 3		3
New Delhi	34/18	VIM-2 (8)	1 3 4		4
Trivandrum	11/11	VIM-6 (2)	1	0	1
Hyderabad	51/17	VIM-2 (3), -6 (2)	1	3	4
Chennai <sup>b</sup>	46/0	None	-	-	-
Kochi	30/20	VIM-2 (5)	0	2	2
Manipal	30/20	VIM-2 (10)	2	3	5
Mumbai	30/22	VIM-2 (3), -6 (2), -11 (1)	0	3	3
Rajkot	25/25	VIM-2 (4)	0	1	1
a. CARB-R: o b. Clones of al. (2007) f	carbapenem-res non-MBL CARE rom a 2003 isol	sistant 3-R <i>P. aeruginosa</i> were docum ate.	nented ar	nd VIM-2	by Toleman et

**Conclusions:** MBL-producing isolates have recently been reported in India, but limited data exists on the prevalence and characterization of MBL-encoding genes. In this study, we show that MBL-producing P. aeruginosa are epidemic in India with a great diversity of MBLtypes (5 VIM types, including new VIM-18). In addition, different MBL-carrying integrons were observed, suggesting widespread dissemination of MBL-carrying mobile elements. Carbapenem resistance rates among *P. aeruginosa* were elevated and were principally caused by MBL production (55%).

## INTRODUCTION

Metallo-B-lactamases (MBLs) constitute one of the most important carbapenem resistance mechanisms found in Pseudomonas aeruginosa. These enzymes can hydrolyze the vast majority of available B-lactam agents available for clinical use and are not inhibited by B-lactamase inhibitors currently marketed or in development. Additionally, the acquired genes encoding these enzymes are carried in mobile genetic structures that usually harbor other resistance elements and facilitate the dissemination of the MBL genes.

Among the six types of MBLs reported to date (IMP, VIM, SPM-1, GIM-1, SIM-1 and AIM), IMP- and VIM-variants are the most prevalent and have been described in numerous geographic locations. Among the VIMtypes, VIM-2 appears to be the most dominant genotype and has been described in 23 countries, including India. Only small numbers of isolates, however, have been evaluated and limited data on the prevalence and characterization of MBL-encoding genes is available from this country.

In this study, initiated to characterize MBL-producing *P. aeruginosa* isolates collected during 2006, we describe a high prevalence of VIMtype enzymes and the dissemination of these genes in numerous Indian medical centers.

Bacterial isolates. During 2006, 10 medical centers located in India were recruited to participate in the SENTRY Antimicrobial Surveillance Program. *P. aeruginosa* isolates were consecutively collected from bloodstream, respiratory tract, skin and soft tissue, and urinary tract infections according to defined protocols. Only clinically significant isolates were included in the study; one per patient episode. Species identification was confirmed by standard biochemical tests and use of Vitek System (bioMérieux; Hazelwood, Missouri, USA), when necessary

Susceptibility testing. All isolates were susceptibility tested against more than 25 antimicrobials by the broth microdilution procedure described by the Clinical and Laboratory Standards Institute (CLSI, 2006) using validated panels manufactured by TREK Diagnostics (Cleveland, Ohio, USA). Interpretations of susceptibility testing results were by CLSI (2008) breakpoint criteria. Escherichia coli ATCC 25922 and P. aeruginosa ATCC 27853 were concurrently tested for quality assurance.

MBL detection. *P. aeruginosa* isolates non-susceptible to imipenem and meropenem (MIC,  $\geq 8$  mg/L) were tested with multiplex PCR using MBL generic primers in a Real-time platform, recently described. Amplicons obtained were sequenced on both strands. The nucleotide sequences and deduced amino acid sequences were analyzed using Lasergene software package (DNASTAR, Madison, WI) and compared with the sequences available through the internet using BLAST (http://www.ncbi. nlm.nih.gov/blast/).

Class 1 integron characterization. *bla*<sub>VIM-6</sub> and *bla*<sub>VIM-18</sub>-carrying integrons were further analyzed. Primers designed in the 5' and 3' conserved sequence (CS) regions of class 1 integrons were used in combination with the MBL primers to determine the size and structure of the integron. One integron of each type was fully sequenced. Restriction patterns generated with Dral and Xbal were used to confirm the integron arrays in the bla<sub>VIM-6</sub> remaining strains.

Molecular Typing. MBL-producing isolates were ribotyped using the Riboprinter Microbial Characterization System<sup>®</sup> (Qualicon, Wilmington, Delaware). Overnight cultures were treated with lysis buffer and placed into the automated system. In brief, this automated process includes bacterial cell lysis, cleavage of DNA using the restriction enzime *Pvull*, size separation using gel electrophoresis and modified Southern blotting. Results were analyzed by the Riboprinter and isolates were considered to have the same ribotype if the similarity coefficient was  $\geq 0.93$ .

## MATERIALS AND METHODS

## RESULTS

Ninety-six of 282 (34.0%) P. aeruginosa isolates collected in 2006 from medical center sites participating on the SENTRY Program (India) showed elevated MIC values for carbapenems (MIC,  $\geq 8$  mg/L).

MBL genes were detected in 53 (55.2%) carbapenem-resistant strains (18.8% overall). All MBL-carrying strains possessed VIMencoding genes. These isolates were detected in 9 of 10 participating institutions (Table 1).

• Sequencing of the MBL gene showed five  $bla_{VIM}$  variants.  $bla_{VIM-2}$  was the most common, being detected in 38 (71.7% of the MBLproducers) isolates collected from 8 of 10 medical centers sampled.

- 4 Indian hospitals.  $bla_{VIM-5}$  and  $bla_{VIM-11}$  were detected in one isolate each (1.8%).
- in one strain. This MBL gene showed a 12bp deletion (position 428) when compared to  $bla_{VIM-2}$  and was carried as a single gene cassette in a class 1 integron.
- Medical centers located in Kolkata and Indore and Hyderabad had 2 distinct VIMvariants (Figure 1).
- Thirty-one different ribotypes and 12 clones (Figure 2) were identified among the VIMproducing P. aeruginosa. Six of these clones

Table 1. Distribution of carbapenem-resistant Pseudomonas spp. in medical centers located in different Indiparticipating in the SENTRY Program for 2006.									
MßL enzymes	Number and species of VIM-producing <i>Pseudomonas</i> isolates in Indian medical centers (total number of carbapenem-resistant isolates)								
	Kolkatta (30)	Indore (42)	New Delhi (34)	Trivandrum (11)	Hyderabad (51)	Chennai (46)	Kochi (30)	Manipal (30)	Mumba (30)
VIM-2	3 PSA	3 PSA 1 PPU	8 PSA		3 PSA		6 PSA	10 PSA	2 PSA 1 PST
VIM-5	1 PSA								
VIM-6		5 PSA		2 PSA	3 PSA				2 PSA
VIM-11									1 PSA
VIM-18	1 PSA								
PSA with negative MBL result	25	34	26	9	45	46	24	20	25
PSA= <i>P. aeruginosa</i> PST= <i>P. stutzeri</i>									

PPU= P. putida

Figure 1. Geographic dissemination of acquired VIM-producing *Pseudomonas* spp. recovered from medical centers in India. Indicated cities represent the location of the participating medical centers and the VIM-types detected. Locations with VIM-6 underlined had isolates with different *bla*<sub>VIM-6</sub> carrying integrons (3.5 and 5 Kb).



 $bla_{VIM-6}$  was detected in 12 (22.7%) strains from

A new variant, named  $bla_{VIM-18}$ , was identified

Mumbai had 3 VIM-types and the isolates from

had isolates carrying distinct *bla*<sub>VIM</sub> genes. Furthermore, 7 of 12 VIM-producing *P*. aeruginosa clones were detected in multiple institutions (Figure 2), indicating inter-hospital dissemination.

- Integrons carrying  $bla_{VIM-6}$  of 3.5 and 5 Kb (5) and 7 isolates, respectively) were characterized further. Both integron types were identified in Indore and Hyderabad (Figure 1).
- VIM-encoding genes were detected in other Pseudomonas species in three medical centers (Indore, Mumbai and Rajkot). Two P. putida carrying bla<sub>VIM-2</sub> and two P. stutzeri, one harboring  $bla_{VIM-2}$  and the other carrying the  $bla_{VIM-11}$  were detected.

**Figure 2.** Ribotyping patterns of clonal *bla*<sub>VIM</sub>-carrying P. aeruginosa isolated in 2006 from Indian medical centers. Twelve clones were detected, six involving distinct VIM-enzymes and medical centers (each clone is indicated in a different color). Eighteen *blavim*-carrying isolates showing unique ribotypes are not presented here.

City	Isolate	MβL	Ribotype	Ribotype pattern	
Hyderabad	248-35-D	VIM-2	PVU 105-528-S-6		
Manipal	251-59-D	VIM-2	PVU 105-528-S-6		
Rajkot	253-17-D	VIM-2	PVU 105-528-S-6		VIM-2/VIM-5 – 4 medical centers
Rajkot	253-40-D	VIM-2	PVU 105-528-S-6		
Kolkatta	243-40-C	VIM-5	PVU 105-528-S-6		
New Delhi	246-68-D	VIM-2	PVU 105-566-S-6		VIM-2 – 1 medical center
New Delhi	246-71-D	VIM-2	PVU 105-566-S-6		
Mumbai	252-16-D	VIM-11	PVU 105-1600-S-6		
Kochi	250-14.2-C	VIM-2	PVU 105-1600-S-6		VINA 2/VINA 11 A modical contors
Manipal	251-18-C	VIM-2	PVU 105-1600-S-6		
New Delhi	246-78-D	VIM-2	PVU 105-1600-S-6		
New Delhi	246-81-D	VIM-2	PVU 127-114-S-4		VINA 2/VINA 6 2 modical contars
Hyderabad	248-77-C	VIM-6	PVU 127-114-S-4		VIIVI-2/VIIVI-6 – 2 medical centers
Kolkatta	243-46-D	VIM-2	PVU 222-134-S-4		VIM 2VIM 6 - 2 modical contors
Mumbai	252-09-C	VIM-6	PVU 222-134-S-4		
Hyderabad	248-26-C	VIM-2	PVU 252-45-S-6		
Indore	244-47-D	VIM-2	PVU 252-45-S-6		
Kolkatta	243-43-D	VIM-2	PVU 252-45-S-6		VIM-2/VIM-6 – 4 medical centers
New Delhi	246-73-D	VIM-2	PVU 252-45-S-6		
Hyderabad	248-11-A	VIM-6	PVU 252-45-S-6		
Manipal	251-22-C	VIM-2	PVU 258-147-S-6		VIDA 2/VIDA C 2 modical contant
Mumbai	252-05-C	VIM-6	PVU 258-147-S-6		VIIVI-2/VIIVI-6 – 2 medical centers
Hyderabad	248-30-D	VIM-2	PVU 258-179-S-8		
Indore	244-46-A	VIM-2	PVU 258-179-S-8		
Kochi	250-04-A	VIM-2	PVU 258-179-S-8		VIM-2 – 5 medical centers
Kolkatta	243-13-D	VIM-2	PVU 258-179-S-8		
New Delhi	246-66-D	VIM-2	PVU 258-179-S-8		
Trivandrum	247-03-D	VIM-6	PVU 258-348-S-6		VINA 6 1 modical contar
Trivandrum	247-06-D	VIM-6	PVU 258-348-S-6		
Manipal	251-05-A	VIM-2	PVU 258-350-S-3		
Manipal	251-54-D	VIM-2	PVU 258-350-S-3		viivi-2 – 1 medical center
Manipal	251-06-C	VIM-2	PVU 258-350-S-4		VINA 2 1 modical contar
Manipal	251-42-C	VIM-2	PVU 258-350-S-4		viivi-2 – 1 medical center
Indore	244-24-D	VIM-6	PVU 258-352-S-2		VINA 6 1 modical contar
Indore	244-29-D	VIM-6	PVU 258-352-S-2		viivi-6 – 1 medical center



an cities
Rajkot (25)
4 PSA 1 PPU
1 PST
21

## CONCLUSIONS

- A high prevalence of MBL-producing isolates was observed in Indian medical centers (approximately 20% of all *P. aeruginosa*), with only one medical center having no MBLproducing strains.
- P. aeruginosa isolates from India were found to harbor distinct blavetypes; interestingly, the amino acid sequences of VIM-2, VIM-6, VIM-11 and VIM-18 are very similar, suggesting that this enzymes could be derived from a unique ancestor.
- Despite genetic diversity recognized among the MBL-producing isolates, inter-hospital dissemination was observed with several clones presenting distinct VIM-types.
- This initial nationwide MBL survey for India suggests a complex dissemination pattern and evolution of VIM-enzymes. The high prevalence and continuous dissemination of these resistance determinants in this region is a serious therapeutic dilemma that compromises the utility of carbapenem-class agents and many other classes of *B*-lactams.

## **SELECTED REFERENCES**

- Clinical and Laboratory Standards Institute. (2006). M7-A7, Methods for dilution antimicrobial susceptibility tests for bacteria that grow aerobically; approved standard - seventh edition. Wayne, PA: CLSI.
- Clinical and Laboratory Standards Institute. (2008). M100-S18, Performance standards for antimicrobial susceptibility testing, 18th informational supplement. Wayne, PA: CLSI.
- Poirel L, Naas T, Nicolas D, Collet L, Bellais S, Cavallo JD, Nordmann P (2000). Characterization of VIM-2, a carbapenem-hydrolyzing metallo-B-lactamase and its plasmid- and integron-borne gene from a Pseudomonas aeruginosa clinical isolate in France. Antimicrob Agents Chemother 44: 891-897.
- Poirel L, Pitout JD, Nordmann P (2007). Carbapenemases: Molecular diversity and clinical consequences. Future Microbiol 2: 501-512.
- Toleman MA, Vinodh H, Sekar U, Kamat V, Walsh TR (2007). *bla*<sub>VIM-2</sub>-harboring integrons isolated in India, Russia, and the United States arise from an ancestral class 1 integron predating the formation of the 3' conserved sequence. Antimicrob Agents Chemother 51: 2636-2638.