Tigecycline Activity Tested against Infrequently Recovered Clinical Species of Non-enteric Gram-negative Bacilli

ABSTRACT

Objectives: To assess tigecycline activity and potency against nonenteric Gram-negative bacilli (NEGNB) clinical isolates. NEGNB usually display multidrug-resistance (MDR) phenotype due to upregulated efflux and chromosomal β-lactamases belonging to Classes A, B, C or D, limiting the therapeutic options for treating infections caused by these organisms. Tigecylcine has demonstrated a broad spectrum of activity against Gram-positive and -negative bacterial pathogens, including many MDR isolates.

Methods: A total of 2,996 clinically-significant isolates of NEGNB (23 species) were collected from 142 hospitals in 32 countries as part of the SENTRY Antimicrobial Surveillance Program over a seven year sampling period (2003-2009; North America [32.1%], Europe [26.8%], Asia-Pacific region [23.4%] and Latin America [17.7%]). Isolates were submitted to a coordinator laboratory where species identifications were confirmed using standard algorithms and Vitek 2, and tested for susceptibility against tigecycline and comparators by reference CLSI methods (M07-A8, 2009). CLSI interpretative criteria were applied when available.

Results: Isolates were recovered mostly from bacteremia (58.1%), pneumonia (29.7%) or skin and skin structure infections (10.4%). Tigecycline was most active against *Pasteurella multocida* (MIC₉₀, 0.12 mg/L) and various Acinetobacter spp. (MIC₉₀, 0.5 mg/L). Aeromonas spp., Rhizobium radiobacter, Ralstonia pickettii and Sphingomonas paucimobilis were also very susceptible to tigecycline (MIC₉₀, 0.5) mg/L). In contrast, Burkholderia cepacia, P. fluorescens/putida, *Chryseobacterium* spp. and *Elizabethkingia* spp. (MIC₉₀, 4 - >4 mg/L) generally exhibited elevated tigecycline MIC values. Tigecycline (MIC_{50/90}, 0.5/2 mg/L; 95.5% inhibited at \leq 2 mg/L) and trimethoprim/ sulfamethoxazole (TMP/SMX) (MIC_{50/90}, $\leq 0.5/1$ mg/L; 96.1% susceptible) were the most active compounds tested against Stenotrophomonas maltophilia; while ceftazidime (MIC_{50/90}, 16/>16 mg/L; 45.1% susceptible), levofloxacin (MIC_{50/90}, 1/4 mg/L; 83.3% susceptible) and ticarcillin/clavulanate (MIC_{50/90}, 32/>128 mg/L; 38.7% susceptible) showed more limited activity against this organism.

	MIC (mg/L)		MIC (mg/L)	
Organism (no. tested)	50%	90%	Organism (no. tested)	50%	90%	
Achromobacter xylosoxidans (130)	0.5	2	Ochrobactrum anthropi (14)	0.5	1	
Acinetobacter haemolyticus (13)	0.12	0.5	Pasteurella multocida (53)	0.06	0.12	
A. junii (47)	0.12	0.5	Pseudomonas spp. (83) ^b	0.25	2	
A. Iwoffii (216)	0.25	0.5	P. fluorescens/putida (191)	2	>4	
Aeromonas spp. (211) ^a	0.25	0.5	Ralstonia pickettii (17)	0.25	0.5	
Alcaligenes faecalis (23)	0.5	2	Rhizobium radiobacter (18)	0.12	0.5	
Burkholderia cepacia (198)	1	4	Sphingomonas paucimobilis (32)	0.12	0.5	
Chryseobacterium indologenes (26)	4	>4	Stenotrophomonas maltophilia	0.5	2	
Elizabethkingia meningosepticum (39)	2	>4	(1685)			

Includes A. caviae (34), A. hydrophila (118), A. salmonicida (1), A. sobria (15), A. veronii (6) and Aeromonas

Includes P. mendocina (14), P. oryzihabitans (27) and P. stutzeri (42).

Conclusions: Tigecycline showed potent in vitro activity against many NEGNB for which there are very limited therapeutic options and susceptibility data to guide therapy. Against S. maltophilia, tigecycline activity was comparable to that of TMP/SMX. The results of this study indicated that tigecycline may have an important role in the treatment of infections caused by these species.

INTRODUCTION

Non-enteric Gram-negative bacilli (NEGNB) occur in moist environments and are ubiquitous in nature, particular in soil and water. These bacterial species can contaminate the hospital environment and spread horizontally through fomites or healthcare workers, and frequently cause hospital outbreaks. As nosocomial opportunists, these organisms rarely affect healthy persons, but cause infections in immunocompromised hosts and those with severe underlying medical conditions.

Pseudomonas spp., Acinetobacter baumannii and Stenotrophomonas maltophilia are the most important NEGNB responsible for bacteremia in hospitalized patients. In addition, *P. aeruginosa* and *Burkholderia cepacia* complex are important pathogens in cystic fibrosis patients. Infections caused by NEGNB have significant morbidity and mortality, owing to the compromised nature of the patients rather than the inherent pathogenicity of the organisms. Moreover, many NEGNB species are intrinsically resistant to multiple antimicrobial agents (MDR) and capable of acquiring additional resistance mechanisms, affecting drugs that are usually active.

Given the compromised affected patient population, intrinsic resistance and that susceptibility testing methodologies are not standardized for some of these organisms, appropriate broad-spectrum empiric therapy strategies are crucial. Tigecycline is approved in the United States (USA) and Europe for the treatment of complicated skin and skin structure infections (SSSI) and intra-abdominal infections (IAI). Tigecycline possesses a proven broad-spectrum of activity against numerous bacterial pathogens, including aerobic and anaerobic species. In this study, we evaluated the activity and potency of tigecycline tested against a clinically-significant and worldwide collection of several infrequently recovered NEGNB.

MATERIALS AND METHODS

Bacterial isolates. A total of 2,996 isolates of NEGNB (23 species) were collected from patients in 142 hospitals in 32 countries as part of the SENTRY Antimicrobial Surveillance Program over a seven year sampling period (2003-2009). Isolates were collected in a prevalence mode and mostly from bacteremia (58.1%), pneumonia (29.7%) or SSSI (10.4%) in hospitalized patients in North America (USA; 32.1%), Europe (26.8%), Asia-Pacific region (23.4%) and Latin America (17.7%). Bacterial species identifications were confirmed using standard algorithms and the automated Vitek 2 system (bioMérieux, Hazelwood, Missouri, USA).

Antimicrobial susceptibility testing. All isolates were tested for susceptibility by reference broth microdilution methods using the Clinical Laboratory Standards Institute recommendations (CLSI; M07-A8, 2009). Susceptibility testing was performed by using validated broth microdilution panels manufactured by TREK Diagnostics Systems/Sensititre (Cleveland, Ohio, USA) and freshly prepared Mueller-Hinton broth (MHB), which is necessary for testing tigecycline. Comparator agents included meropenem, ceftazidime, piperacillin/tazobactam, ticarcillin/clavulanate, amikacin, tobramycin, levofloxacin, trimethoprim/sulfamethoxazole (TMP/SMX) and polymyxin B.

Validation of the minimum inhibitory concentration (MIC) values was performed by concurrent testing of CLSIrecommended (M100-S20, 2010) quality control (QC) strains: Enterococcus faecalis ATCC 29212, Staphylococcus aureus ATCC 29213, Escherichia coli ATCC 25922 and P. aeruginosa ATCC 27853.

Categorical interpretation of comparator MIC values were performed according to CLSI (M100-S20, 2010) criteria, when available. Tigecycline breakpoints approved by the USA-FDA for Enterobacteriaceae ($\leq 2 \text{ mg/L}$ for susceptible and $\geq 8 \text{ mg/L}$ for resistant) were applied for comparison purposes only. MIC ranges for tigecycline and comparator agents tested against ATCC QC strains were those published in the CLSI M100-S20 (2010) document.

RE MENDES, HS SADER, MG STILWELL, RN JONES JMI Laboratories, North Liberty, Iowa, USA

RESULTS

- S. maltophilia represented the majority of isolates in this investigation (56.2%). Other organisms tested included non-baumannii Acinetobacter spp. (9.2%), Aeromonas spp. (7.0%), B. cepacia (6.6%), Pseudomonas fluorescens/putida (6.4%) and Achromobacter xylosoxidans (4.3%) among others (Table 1).
- Tigecycline (MIC_{50/90}, 0.5/2 mg/L; 95.5% inhibited at ≤ 2 mg/L) and TMP/SMX (MIC_{50/90}, ≤0.5/1 mg/L; 96.1% susceptible) were very active against S. maltophilia (Tables 1 and 2). Levofloxacin (MIC_{50/90}, 1/4 mg/L; 83.3% susceptible) was two- and four-fold less active than tigecycline and TMP/SMX against S. maltophilia, respectively (Table 2).
- Tigecycline (MIC_{50/90}, 0.12/0.5 mg/L; 99.6% inhibited at $\leq 2 \text{ mg/L}$) was the most active compound tested against non-baumannii Acinetobacter spp., followed by polymyxin B (MIC_{50/90}, ≤0.5/1 mg/L; 98.2% susceptible), meropenem (MIC_{50/90}, 0.25/2 mg/L; 94.6% susceptible) and amikacin (MIC_{50/90}, 2/8 mg/L; 92.0% susceptible; Tables 1 and 2).

SELECTED REFERENCES

- Clinical and Laboratory Standards Institute. (2009). M07-A8, Methods for dilution antimicrobial susceptibility tests for bacteria that grow aerobically; approved standard - seventh edition. Wayne, PA: CLSI.
- 2. Clinical and Laboratory Standards Institute. (2010). M100-S20. Performance standards for antimicrobial susceptibility testing, 20th informational supplement. Wayne, PA: CLSI.
- 3. Daxboeck F, Assadian O, Blacky A, Koller W, Hirschl AM (2004). Resistance of Gram-negative non-fermentative bacilli causing bloodstream infection, Vienna, 1996-2003. Eur J Clin Microbiol Infect Dis 23: 415-6.
- 4. Livermore DM (2005). Tigecycline: What is it, and where should it be used? J Antimicrob Chemother 56: 611-614.
- 5. Livermore DM, Hope R, Brick G, Lillie M, Reynolds R; BSAC Working Parties on Resistance Surveillance (2008). Non-susceptibility trends among Pseudomonas aeruginosa and other non-fermentative Gramnegative bacteria from bacteraemias in the UK and Ireland, 2001-06. J Antimicrob Chemother 62: 55-63.
- 6. McGowan JE Jr. (2006) Resistance in nonfermenting Gram-negative bacteria: Multidrug resistance to the maximum. Am J Infect Control 34: S29-S37.
- . Sader HS and Jones RN (2005). Antimicrobial susceptibility of uncommonly isolated non-enteric Gram-negative bacilli. Int J Antimicrob Agents 25: 95-109.
- 8. Stein GE, Craig WA (2006). Tigecycline: A critical analysis. *Clin Infect Dis* 43: 518-524.
- 9. Tygacil[®] Package Insert (2005). Wyeth Pharmaceuticals Inc. http://www.wyeth.com/content/showlabeling.asp?id=491. (19 February 2010, date last accessed).

ACKNOWLEDGMENT

This study was supported by a grant from Wyeth Pharmaceuticals.

- Tigecycline (MIC_{50/90}, 0.25/0.5 mg/L) and levofloxacin $(MIC_{50/90}, \leq 0.5/\leq 0.5 \text{ mg/L})$ were the most active compounds tested against Aeromonas spp., inhibiting 99.5 and 98.6% of isolates at $\leq 2 \text{ mg/L}$, respectively (Tables 1 and 2).
- Only meropenem (MIC_{50/90}, 2/4 mg/L; 93.9% susceptible) and TMP/SMX (MIC_{50/90}, ≤0.5/2 mg/L 93.9% susceptible) exhibited good coverage (>90% susceptibility) against *B. cepacia*. Tigecycline (MIC_{50/90}, 1/4 mg/L; 84.8% inhibited at ≤ 2 mg/L) and ceftazidime (MIC_{50/90}, 4/16 mg/L; 88.4% susceptible) were less activity against this species (Tables 1 and 2).
- Tigecycline (MIC_{50/90}, 0.5/2 mg/L; 95.4% inhibited at \leq 2 mg/L), meropenem (MIC_{50/90}, 0.25/4 mg/L; 93.8% susceptible), piperacillin/tazobactam (MIC_{50/90}, 1/8 mg/L; 95.4% susceptible) and TMP/SMX (MIC_{50/90}, $\leq 0.5/1$ mg/L; 91.5% susceptible) were the most active compounds tested against A. xylosoxidans (Tables 1 and 2).
- High in vitro potency was noted for tigecycline and most comparator agents when tested against Pasteurella multocida and Pseudomonas spp. (P. mendocina, P. oryzihabitans and P. stutzeri; Table 2).
- Among *Alcaligenes faecalis*, the highest tigecycline MIC value was 4 mg/L, and 91.3% of strains were inhibited at $\leq 2 \text{ mg/L}$ (Table 1).
- The highest MIC value observed for tigecycline when tested against Rhizobium radiobacter was 0.5 mg/L. Furthermore, tigecycline inhibited all tested Ochrobactrum anthropi, Ralstonia pickettii, and Sphingomonas paucimobilis isolates at ≤1 mg/L (Table 1).
- Tigecycline demonstrated limited activity against Chryseobacterium indologenes (MIC_{50/90}, 4/>4 mg/L), *Elizabethkingia meningosepticum* (MIC_{50/90}, 2/>4 mg/L) and *P. fluorescens/putida* (MIC_{50/90}, 2/>4 mg/L; Table 1).

Table 1. MIC distribution of tigecycline tested against non-enteric Gram-negative bacilli clinical isolates collected as part of the SENTRY Antimicrobial Surveillance Program (2003-2009).

		Number (cumulative %) inhibited at tigecycline MIC value (mg/L) of:									
Organism group (number tested; % of total)	≤0.06	0.12	0.25	0.5	1	2	4	>4			
Stenotrophomonas maltophilia (1685; 56.2)	2(0.12)	29(1.8)	211(14.4)	626(51.5)	547(84.0)	195(95.5)	61(99.2)	14(100.0)			
Acinetobacter spp.a (276; 9.2)	42(15.2)	97(50.4)	94(84.4)	27(94.2)	12(98.6)	3(99.6)	0(99.6)	1(100.0)			
Aeromonas spp. ^b (211; 7.0)	1(0.5)	37(18.0)	108(69.2)	52(93.8)	11(99.0)	1(99.5)	1(100.0)	_			
Burkholderia cepacia (198; 6.6)	2(1.0)	2(2.0)	12(8.1)	53(34.8)	64(67.2)	35(84.8)	19(94.4)	11(100.0)			
Pseudomonas fluorescens/putida (191; 6.4)	1(0.5)	9(5.2)	14(12.6)	22(24.1)	44(47.1)	54(75.4)	20(85.9)	27(100.0)			
Achromobacter xylosoxidans (130; 4.3)	4(3.1)	5(6.9)	13(16.9)	67(68.5)	25(87.7)	10(95.4)	6(100.0)	_			
Pseudomonas spp. ^c (83; 2.8)	7(8.4)	25(38.5)	22(65.1)	9(75.9)	7(84.3)	9(95.2)	4(100.0)	_			
Pasteurella multocida (53; 1.8)	44(83.0)	6(94.3)	2(98.1)	0(98.1)	0(98.1)	1(100.0)	_	_			
Elizabethkingia meningosepticum (39; 1.3)	0(0.0)	0(0.0)	0(0.0)	1(2.6)	8(23.1)	17(66.7)	8(87.2)	5(100.0)			
Sphingomonas paucimobilis (32; 1.1)	12(37.5)	8(62.5)	8(87.5)	1(90.3)	3(100.0)	_	_	_			
Chryseobacterium indologenes (26; 0.9)	0(0.0)	0(0.0)	2(7.7)	3(19.2)	5(38.5)	2(46.1)	9(80.8)	5(100.0)			
Alcaligenes faecalis (23; 0.8)	1(4.3)	4(21.7)	4(39.1)	3(52.2)	8(87.0)	1(91.3)	2(100.0)	_			
Rhizobium radiobacter (18; 0.6)	2(11.1)	7(50.0)	6(83.3)	3(100.0)	_	_	_	_			
Ralstonia pickettii (17; 0.6)	0(0.0)	2(11.8)	9(64.7)	5(94.1)	1(100.0)	_	_	_			
Ochrobactrum anthropi (14; 0.5)	0(0.0)	0(0.0)	3(21.4)	7(71.4)	4(100.0)	_	_	_			

ECCMID 2010

JMI Laboratories North Liberty, IA, USA www.jmilabs.com 319.665.3370, 319.665.3371 rodrigo-mendes@jmilabs.com

Activity of tigecycline and comparators tested against most common non-enteric Gram-negative bacilli clinical isolates Table 2. collected as part of the SENTRY Antimicrobial Surveillance Program over a seven year sampling period (2003-2009).

Organism		(mg/L)	%	%	Organism		(mg/L)	%	%	Organism	MIC (%	%
Antimicrobial agent	50%	90%	Susceptible	a Resistant	a Antimicrobial agent	50%	90%	Susceptible	Resistant ^a	^a Antimicrobial agent	50%	90%	Susceptible	Resistant
Stenotrophomonas malto	•	. ,	b	a ab	Achromobacter xylosoxic			b	a ab	Chryseobacterium indolo	genes (2		, s , b	b
Tigecycline	0.5	2	95.5 ^b	0.8 ^b	Tigecycline	0.5	2	95.4 ^b	0.0 ^b	Tigecycline	4	>4	46.1 ^b	19.2 ^b
Meropenem	>8	>8	_c	_	Meropenem	0.25	4	93.8	3.8	Meropenem	>8	>8	19.2	73.1
Ceftazidime	16	>16	45.1	42.1	Ceftazidime	4	16	87.7	4.6	Ceftazidime	4	>16	80.8	11.5
Ticarcillin/clavulanate	32	>128	38.7	24.8	Piperacillin/tazobactam	1	8	95.4	3.8	Piperacillin/tazobactam	4	8	96.2	0.0
Amikacin	>32	>32	_	-	Amikacin	>32	>32	12.3	80.8	Amikacin	>32	>32	11.5	57.7
Levofloxacin	1	4	83.3	8.2	Levofloxacin	2	>4	60.8	16.2	Levofloxacin	≤0.5	>4	73.1	23.1
TMP/SMX ^d	≤0.5	1	96.1	3.9	TMP/SMX	≤0.5	1	91.5	8.5	TMP/SMX	≤0.5	2	96.2	3.8
Polymyxin B	≤1	>4	_	-	Polymyxin B	2	4	86.9	6.9	Polymyxin B	>4	>4	7.7	92.3
Acinetobacter spp. (276)	e		h	h	Pseudomonas spp. (83) ^g			h	h	Alcaligenes faecalis (23)			h	h
Tigecycline	0.12	0.5	99.6 ^b	0.4 ^b	Tigecycline	0.25	2	95.2 ^b	0.0 ^b	Tigecycline	0.5	2	91.3 ^b	0.0 ^b
Meropenem	0.25	2	94.6	2.5	Meropenem	≤0.12	2	91.6	3.6	Meropenem	≤0.12	>8	82.6	13.0
Ceftazidime	4	>16	79.7	15.2	Ceftazidime	≤1	4	94.0	4.8	Ceftazidime	2	>16	78.3	21.7
Piperacillin/tazobactam	≤0.5	>64	83.7	13.8	Piperacillin/tazobactam	2	16	95.2	2.4	Piperacillin/tazobactam	≤0.5	>64	82.6	13.0
Ampicillin/sulbactam	≤2	>16	83.0	11.6	Amikacin	≤4	≤4	97.6	2.4	Amikacin	≤4	>32	78.3	21.7
Amikacin	2	8	92.0	5.8	Levofloxacin	≤0.5	4	88.0	8.4	Levofloxacin	≤0.5	>4	69.6	13.0
Levofloxacin	≤0.5	4	88.8	7.2	TMP/SMX	≤0.5	>2	74.7	25.3	TMP/SMX	≤0.5	>2	87.0	13.0
TMP/SMX	≤0.5	>2	84.8	15.2	Polymyxin B	≤1	≤1	95.2	4.8	Polymyxin B	≤0.5	4	87.0	4.3
Polymyxin B	≤0.5	1	98.2	1.8	Pasteurella multocida (53	3)				Rhizobium radiobacter (1	8)			
Aeromonas spp. (211) ^f					Tigecycline	≤0.06	0.12	100.0 ^b	0.0 ^b	Tigecycline	0.12	0.5	100.0 ^b	0.0 ^b
Tigecycline	0.25	0.5	99.5 ^b	0.0 ^b	Meropenem	≤0.12	≤0.12	100.0	0.0	Meropenem	≤0.12	0.25	100.0	0.0
Meropenem	0.25	1	97.2	2.8	Ceftazidime	≤1	2	100.0	0.0	Ceftazidime	16	>16	44.4	16.7
Ceftazidime	≤1	8	91.9	5.2	Piperacillin/tazobactam	≤0.5	≤0.5	100.0	0.0	Piperacillin/tazobactam	8	16	94.4	0.0
Piperacillin/tazobactam	8	>64	63.8	16.7	Amikacin	8	16	98.1	0.0	Amikacin	16	16	100.0	0.0
Amikacin	≤4	8	96.7	2.4	Levofloxacin	≤0.5	≤0.5	100.0	0.0	Levofloxacin	≤0.5	1	100.0	0.0
Levofloxacin	≤0.5	≤0.5	98.6	1.4	TMP/SMX	≤0.5	≤0.5	100.0	0.0	TMP/SMX	≤0.5	>2	83.3	16.7
TMP/SMX	≤0.5	>2	86.3	13.7	Polymyxin B	≤1	2	92.5	3.8	Polymyxin B	≤1	>4	72.2	16.7
Polymyxin B	≤1	>4	66.8	30.3	Elizabethkingia meningos	septicun	ı (39)			Ralstonia pickettii (17)				
Burkholderia cepacia (19	8)				Tigecycline	2	>4	66.7 ^b	12.8 ^b	Tigecycline	0.25	0.5	100.0 ^b	0.0 ^b
Tigecycline	1	4	84.8 ^b	5.6 ^b	Meropenem	>8	>8	2.6	97.4	Meropenem	>8	>8	29.4	52.9
Meropenem	2	4	93.9	3.5	Ceftazidime	>16	>16	5.1	94.9	Ceftazidime	8	>16	52.9	11.8
Ceftazidime	4	16	88.4	5.6	Piperacillin/tazobactam	4	32	89.7	0.0	Piperacillin/tazobactam	8	>64	70.6	11.8
Ticarcillin/clavulanate	>128	>128	4.5	89.9	Amikacin	>32	>32	12.8	59.0	Amikacin	>32	>32	41.2	58.8
Amikacin	>32	>32	_	-	Tobramycin	>16	>16	0.0	100.0	Levofloxacin	≤0.5	≤0.5	100.0	0.0
Levofloxacin	2	>4	74.7	12.6	Levofloxacin	≤0.5	4	87.2	7.7	TMP/SMX	≤0.5	1	94.1	5.9
TMP/SMX	≤0.5	2	93.9	6.1	TMP/SMX	1	>2	64.1	35.9	Polymyxin B	>4	>4	5.9	94.1
Polymyxin B	>4	>4	_	-	Polymyxin B	>4	>4	0.0	100.0	Ochrobactrum anthropi (*				
Pseudomonas fluorescer	ns/putid				Sphingomonas paucimol					Tigecycline	, 0.5	1	100.0 ^b	0.0 ^b
Tigecycline	2	>4	75.4 ^b	14.1 ^b	Tigecycline	0.12	0.5	100.0 ^b	0.0 ^b	Meropenem	0.5	2	100.0	0.0
Meropenem	2	8	89.5	0.0	Meropenem	≤0.12	2	93.5	3.2	Ceftazidime	>16	>16	7.1	92.9
Ceftazidime	2	8	100.0	0.0	Ceftazidime	4	_ >16	62.5	21.9	Piperacillin/tazobactam	>64	>64	7.1	85.7
Piperacillin/tazobactam	_ 16	32	73.7	8.9	Piperacillin/tazobactam	2	64	75.0	6.3	Amikacin	8	32	85.7	7.1
Amikacin	≤4	≤4	100.0	0.0	Amikacin	∠ ≤4	32	84.4	3.1	Levofloxacin	≤0.5	≤0.5	92.9	7.1
Levofloxacin	_⊣ ≤0.5	2	100.0	0.0	Levofloxacin	 ≤0.5	4	84.4	6.3	TMP/SMX	_0.0 ≤0.5	_0.0 ≤0.5	92.9	7.1
TMP/SMX	<u></u> 20.0	>2	21.1	78.9	TMP/SMX	<u></u> ≤0.5	- ≤0.5	93.8	6.3	Polymyxin B	<u>⊐</u> 0.5 2	⊒0.0 >4	71.4	14.3
Polymyxin B	≤0.5	>4	89.5	10.5	Polymyxin B	<u>⊐0.5</u> 2	<u>⊐0.0</u> >4	59.4	37.5		-	~ 7	11.7	
a. Breakpoint susceptibili						<u> </u>	~7	00.4	0.10					
		, ແລ ມ/U/U		A CH IVI I () () = ()										

Breakpoint susceptibility criteria as published by the CLSI M100-S20 document (2010).

Tigecycline susceptible breakpoints for Enterobacteriaceae (≤2 mg/L for susceptible and ≥8 mg/L for resistant) approved by the US-FDA were applied for comparison purposes only.

-, indicates no breakpoint criteria are available for the respective drug/organism combination.

TMP/SMX, trimethoprim/sulfamethoxazole

Includes A. haemolyticus (13), A. junii (47) and A. Iwoffii (216).

Includes A. caviae (34), A. hydrophila (118), A. salmonicida (1), A. sobria (15), A. veronii (6) and Aeromonas spp. (37).

Includes P. mendocina (14), P. oryzihabitans (27) and P. stutzeri (42).

CONCLUSIONS

- Tigecycline showed potent in vitro activity against many NEGNB for which there are very limited therapeutic options and susceptibility testing data to guide therapy.
- Against S. maltophilia, tigecycline activity was comparable to that of trimethoprim/sulfamethoxazole.
- The results of this study indicated that tigecycline may have an important role in the treatment of infections caused by these NEGNB species.



Includes A. caviae (34), A. hydrophila (118), A. salmonicida (1), A. sobria (15), A. veronii (6) and Aeromonas spp. (37). Includes P. mendocina (14), P. oryzihabitans (27), and P. stutzeri (42).