

Ceftobiprole Activity Tested against Bacterial Isolates from Hospitalized Patients with Pneumonia in European Hospitals and Israel (2013)

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ABSTRACT

Objective: To evaluate the activity of ceftobiprole and comparator agents against pathogens isolated during 2013 from patients hospitalized with pneumonia in Europe and Israel. Ceftobiprole medoclaril is a parenteral broad-spectrum, anti-MRSA cephalosporin with activity against Gram-positive and -negative pathogens including *Pseudomonas aeruginosa*. It is the first anti-MRSA cephalosporin approved for both the treatment of hospital-acquired pneumonia (excluding ventilator-associated pneumonia) and community-acquired pneumonia in adults in 12 European Union countries.

Methods: Non-duplicate consecutive isolates from patients hospitalized with pneumonia were collected during 2013 from 26 medical centres in 13 European countries and Israel. Ceftobiprole, the active form of the prodrug ceftobiprole medoclaril and comparator agents were susceptibility tested according to Clinical and Laboratory Standards Institute (CLSI) methods in validated minimum inhibitory concentration (MIC) panels manufactured by ThermoFisher Inc., formerly TREK Diagnostics (Cleveland, Ohio, USA) to determine the antimicrobial susceptibility of ceftobiprole and comparator agents. *S. pneumoniae* strains were tested in CA-MHB supplemented with 2.5-5% lysed horse blood and *Haemophilus influenzae* were tested in *Haemophilus* test medium (M07-A9, 2012). *Moraxella catarrhalis* strains were tested in cation-adjusted Mueller-Hinton broth (CA-MHB; M45-A2). β -lactamase production was characterized by the nitrocefin disk test (Remel; Lenexa, Kansas, USA).

Results: The ceftobiprole MIC_{50/90} values for 336 *S. aureus* were 0.5/2 mg/L, respectively. For methicillin-resistant *S. aureus* (MRSA, 28.3% of all *S. aureus*) MIC_{50/90} values were 2/2 mg/L, respectively. The highest MIC value for methicillin-susceptible *S. aureus* (MSSA) was 0.5 mg/L and for MRSA it was 2 mg/L. Ceftobiprole was highly potent against all 47 *S. pneumoniae* isolates (100.0% susceptibility) with 48.9% of isolates with a MIC₅₀ value of ≤ 0.008 mg/L (highest ceftobiprole MIC, 0.5 mg/L). These included three penicillin-resistant isolates (6.4%), all with a penicillin MIC of 4 mg/L and a ceftobiprole MIC of 0.5 mg/L. These three isolates were also ceftazidime-non-susceptible (ceftazidime MIC, 1-2 mg/L). Ceftobiprole susceptibility for MRSA and methicillin-susceptible *S. aureus* (MSSA) were 2/2 and 0.25/0.5 mg/L, respectively (Tables 1 and 2). The highest MIC for MSSA was 0.5 mg/L.

Conclusions: Ceftobiprole demonstrated potent broad-spectrum *in vitro* activity against pathogens from European and Israeli patients hospitalized with pneumonia including MRSA, penicillin-resistant/ceftazidime-non-susceptible *S. pneumoniae*, Enterobacteriaceae and *P. aeruginosa*.

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METHODS

Isolates were consecutively collected (one isolate per infection episode) from patients with lower respiratory tract infections with an emphasis on obtaining isolates from invasive sampling (tracheal aspirates, bronchoalveolar lavage, protected brush samples, etc.) at 26 medical centres in 13 European countries and Israel during 2013. Broth microdilution testing was performed according to Clinical and Laboratory Standards Institute (CLSI) methods in validated minimum inhibitory concentration (MIC) panels manufactured by ThermoFisher Inc., formerly TREK Diagnostics (Cleveland, Ohio, USA) to determine the antimicrobial susceptibility of ceftobiprole and comparator agents. *S. pneumoniae* strains were tested in CA-MHB supplemented with 2.5-5% lysed horse blood and *Haemophilus influenzae* were tested in *Haemophilus* test medium (M07-A9, 2012). *Moraxella catarrhalis* strains were tested in cation-adjusted Mueller-Hinton broth (CA-MHB; M45-A2). β -lactamase production was characterized by the nitrocefin disk test (Remel; Lenexa, Kansas, USA).

Quality control (QC) testing was performed using the following QC strains: *S. aureus* ATCC 29213, *S. pneumoniae* ATCC 49619, *E. coli* ATCC 25922 and ATCC 35218, *P. aeruginosa* ATCC 27853, and *H. influenzae* ATCC 49247 and 49766. QC results were within published CLSI guidelines (M100-S24 and EUCAST, 2014) except for ceftobiprole (MHRSA, M00-S24). Isolates with an ESBL phenotype were defined as a MIC ≥ 2 mg/L for one of ceftazidime, or ceftazidime, or aztreonam [CLSI, 2014].

Results: The ceftobiprole MIC_{50/90} values for 336 *S. aureus* were 0.5/2 mg/L, respectively. For methicillin-resistant *S. aureus* (MRSA, 28.3% of all *S. aureus*) MIC_{50/90} values were 2/2 mg/L, respectively. The highest MIC value for methicillin-susceptible *S. aureus* (MSSA) was 0.5 mg/L and for MRSA it was 2 mg/L. Ceftobiprole was highly potent against all 47 *S. pneumoniae* isolates (100.0% susceptibility) with 48.9% of isolates with a MIC₅₀ value of ≤ 0.008 mg/L (highest ceftobiprole MIC, 0.5 mg/L). These included three penicillin-resistant isolates (6.4%), all with a penicillin MIC of 4 mg/L and a ceftobiprole MIC of 0.5 mg/L. These three isolates were also ceftazidime-non-susceptible (ceftazidime MIC, 1-2 mg/L). Ceftobiprole susceptibility for MRSA and methicillin-susceptible *S. aureus* (MSSA) were 2/2 and 0.25/0.5 mg/L, respectively (Tables 1 and 2). The highest MIC for MSSA was 0.5 mg/L.

Conclusions: Ceftobiprole demonstrated potent broad-spectrum *in vitro* activity against pathogens from European and Israeli patients hospitalized with pneumonia including MRSA, penicillin-resistant/ceftazidime-non-susceptible *S. pneumoniae*, Enterobacteriaceae and *P. aeruginosa*.

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CONCLUSIONS

Ceftobiprole exhibited potent *in vitro* activity against a broad range of Gram-positive and -negative pathogens isolated from European and Israeli patients hospitalized with pneumonia during 2013. All *S. pneumoniae* (including isolates that were not susceptible to ceftazidime nor penicillin) and *S. aureus* (including MRSA) were susceptible to ceftobiprole. Ceftobiprole also exhibited potent activity against Gram-negative bacteria including *H. influenzae*, non-ESBL-phenotype *E. coli* and *K. pneumoniae*. Additionally against *P. aeruginosa*, ceftobiprole showed similar activity to cefepime and ceftazidime. The spectrum of activity of ceftobiprole supports the role in the treatment of pulmonary infections caused by these organisms.

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Table 1. Summary of ceftobiprole activity tested against pathogens from patients hospitalized with bacterial pneumonia in Europe and Israel (2013).

Organism group (no. tested)	No. of organisms (cumulative %)										MIC ₅₀	MIC ₉₀	
	≤ 0.008	0.015	0.03	0.06	0.12	0.25	1	2	4	≥ 8			
<i>Staphylococcus aureus</i> (336)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	1 (0.3)	123 (36.9)	127 (37.7)	37 (86.7)	48 (100.0)	0	0	0.5	2
MSSA (241)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	1 (0.4)	123 (51.5)	117 (100.0)	0	0	0	0	0.25	0.5
MRSA (95)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	10 (10.5)	37 (49.5)	48 (100.0)	0	0	0	0.015	0.5
<i>Streptococcus pneumoniae</i> (47)	23 (48.9)	10 (70.2)	17 (72.3)	17 (72.3)	17 (72.3)	17 (72.3)	17 (72.3)	17 (72.3)	17 (72.3)	17 (72.3)	17 (72.3)	0.008	0.015
Pen ^r -S (MIC ≤ 0.06 mg/L; 32)	23 (71.9)	8 (96.9)	1 (100.0)	0	0	0	0	0	0	0	0	0.008	0.015
Pen ^r -N (MIC 0.12-1 mg/L; 7)	0 (0.0)	2 (28.6)	0 (28.6)	0 (28.6)	0 (28.6)	0 (28.6)	0 (28.6)	0 (28.6)	0 (28.6)	0 (28.6)	0 (28.6)	0.25	0.5
Pen ^r -N (MIC ≥ 2 mg/L; 8)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0.25	0.5
CRO-non-S (MIC ≤ 1 mg/L; 11)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0.25	0.5
CRO-non-S (MIC ≥ 2 mg/L; 2)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0.5	0.25
<i>Haemophilus influenzae</i> (54)	0 (0.0)	0 (0.0)	21 (38.9)	18 (72.3)	18 (72.3)	18 (72.3)	18 (72.3)	18 (72.3)	18 (72.3)	18 (72.3)	18 (72.3)	0.06	0.25
β -lactamase-positive (8)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0.03	0.03
β -lactamase-negative (45)	0 (0.0)	0 (0.0)	15 (33.3)	16 (68.9)	17 (84.4)	17 (84.4)	17 (84.4)	17 (84.4)	17 (84.4)	17 (84.4)	17 (84.4)	0.06	0.25
Enterobacteriaceae (743)	1 (0.1)	72 (8.9)	205 (26.5)	148 (65.4)	127 (17.1)	27 (74.7)	12 (77.9)	8 (79.0)	10 (80.3)	146 (100.0)	0.06	0.25	
<i>Escherichia coli</i> (221)	0 (0.0)	21 (9.5)	102 (45.7)	40 (73.8)	10 (73.8)	5 (80.5)	2 (81.4)	1 (81.9)	1 (81.9)	39 (100.0)	0.03	0.8	
ESBL-phenotype (46)	0 (0.0)	0 (0.0)	1 (2.2)	0 (2.2)	2 (6.5)	1 (8.7)	1 (10.9)	0 (10.9)	1 (13.0)	1 (15.2)	39 (100.0)	0.8	0.8
non-ESBL-phenotype (175)	0 (0.0)	21 (12.0)	101 (69.7)	40 (62.6)	2 (8.1)	4 (9.4)	1 (10.0)	0	0	0	0.03	0.06	
<i>Klebsiella</i> spp. (224)	0 (0.0)	24 (10.7)	72 (49.9)	20 (51.8)	14 (65.4)	14 (65.4)	3 (62.9)	7 (66.1)	1 (67.9)	72 (100.0)	0.06	0.8	
non-ESBL-phenotype (83)	0 (0.0)	0 (0.0)	11 (21.2)	1 (21.2)	0 (21.2)	0 (21.2)	0 (21.2)	0 (21.2)	0 (21.2)	71 (100.0)	0.03	0.8	
non-ESBL-phenotype (141)	0 (0.0)	24 (17.0)	77 (61.4)	19 (69.8)	14 (65.4)	14 (65.4)	3 (62.9)	7 (66.1)	1 (67.9)	72 (100.0)	0.03	0.25	
<i>Klebsiella pneumoniae</i> (172)	0 (0.0)	23 (13.4)	62 (49.4)	10 (55.2)	3 (57.0)	5 (59.9)	0 (59.9)	5 (62.8)	3 (64.5)	1 (65.1)	60 (100.0)	0.06	0.8
ESBL-phenotype (71)	0 (0.0)	0 (0.0)	11 (15.4)	1 (25.8)	0 (25.8)	0 (25.8)	0 (25.8)	0 (25.8)	0 (25.8)	60 (100.0)	0.8	0.8	
non-ESBL-phenotype (101)	0 (0.0)	23 (22.8)	51 (68.2)	9 (62.1)	3 (69.0)	5 (70.0)	0 (70.0)	0 (70.0)	0 (70.0)	0 (70.0)	0.03	0.06	
<i>Enterobacter</i> spp. (151)	0 (0.0)	15 (9.9)	54 (45.7)	35 (68.9)	10 (75.5)	1 (76.2)	3 (78.1)	1 (78.8)	4 (81.5)	7 (86.1)	21 (100.0)	0.06	0.8
<i>Citrobacter</i> spp. (23)	0 (0.0)	0 (0.0)	11 (47.8)	5 (69.6)	0 (69.6)	0 (69.6)	0 (69.6)	0 (69.6)	0 (69.6)	0 (69.6)	0 (69.6)	0.06	1
<i>Proteus mirabilis</i> (24)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0.03	0.06
<i>Pseudomonas aeruginosa</i> (419)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	1 (0.2)	12 (3.1)	116 (30.8)	89 (52.0)	59 (66.1)	142 (100.0)	2	0.8	

Table 2. Activity of ceftobiprole and comparator antimicrobial agents when tested against Gram-positive pathogens from patients hospitalized with pneumonia in Europe and Israel (2013).

Organism group (no. tested)	MIC (mg/L)					%Susc. / %Resistant	Organism group (no. tested)	MIC (mg/L)					%Susc. / %Resistant		
	MIC ₅₀	MIC ₉₀	Range	CLSP ^a	EUCAST ^b			MIC ₅₀	MIC ₉₀	Range	CLSP ^a	EUCAST ^b			
<i>Staphylococcus aureus</i> (336)	0.5	2	0.12-2	-	100.0/0.0	0.5	2	0.12-2	-	100.0/0.0	0.5	2	0.12-2	-	100.0/0.0
Cefepime	0.5	2	0.12-2	-	100.0/0.0	0.5	2	0.12-2	-	100.0/0.0	0.5	2	0.12-2	-	100.0/0.0
Ceftazidime	0.25	0.5	0.12-1	71.1/93.3	100.0/0.0	0.5	2	0.12-2	-	100.0/0.0	0.5	2	0.12-2	-	100.0/0.0
Daptomycin	0.25	0.5	0.12-1	71.1/93.3	100.0/0.0	0.5	2	0.12-2	-	100.0/0.0	0.5	2	0.12-2	-	100.0/0.0
Levofloxacin	0.25	0.5	0.12-1	71.1/93.3	100.0/0.0	0.5	2	0.12-2	-	100.0/0.0	0.5	2	0.12-2	-	100.0/0.0
Linezolid	1	1	0.5-2	100.0/0.0	100.0/0.0	0.5	2	0.12-2	-	100.0/0.0	0.5	2	0.12-2	-	100.0/0.0
Tetracycline	0.12	0.25	0.06-0.5	94.6/100.0	100.0/0.0	0.5	2	0.12-2	-	100.0/0.0	0.5	2	0.12-2	-	100.0/0.0
Trimethoprim/sulfamethoxazole	0.05	0.05	0.05-0.5	100.0/0.0	100.0/0.0	0.5	2	0.12-2	-	100.0/0.0	0.5	2	0.12-2	-	100.0/0.0
Vancomycin	1	1	0.25-2	100.0/0.0	100.0/0.0	0.5	2	0.12-2	-	100.0/0.0	0.5	2	0.12-2	-	100.0/0.0
MSSA (241)	0.25	0.5	0.12-0.5	-	100.0/0.0	0.25	0.5	0.12-0.5	-	100.0/0.0	0.25	0.5	0.12-0.5	-	100.0/0.0
Cefepime	0.25	0.5	0.12-0.5	-	100.0/0.0	0.25	0.5	0.12-0.5	-	100.0/0.0	0.25	0.5	0.12-0.5	-	100.0/0.0
Daptomycin	0.25	0.5	0.12-0.5	-	100.0/0.0	0.25	0.5	0.12-0.5	-	100.0/0.0	0.25	0.5	0.12-0.5	-	100.0/0.0
Levofloxacin	0.12	0.25	0.12-0.4	97.5/125	97.5/125	0.5	1	0.5-2	100.0/0.0	100.0/0.0	0.5	1	0.5-2	100.0/0.0	
Linezolid	0.5	1	0.5-2	100.0/0.0	100.0/0.0	0.5	1	0.5-2	100.0/0.0	100.0/0.0	0.5	1	0.5-2	100.0/0.0	
Tetracycline	0.12	0.25	0.06-0.5	94.6/100.0	94.6/100.0	0.5									