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Activity of SPR206 and Comparator Agents Against Pseudomonas aeruginosa and Acinetobacter spp. Causing Infections in Europe and **Adjacent Regions**

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Introduction

- Non-fermentative Gram-negative bacilli (NF-GNB) are opportunistic organisms that have emerged as important healthcare-associated pathogens, mainly in immunocompromised patients.
- These organisms are innately less susceptible to many antimicrobial classes due to the presence of intrinsic genes encoding β -lactamases and efflux pumps.
- SPR206 is a next-generation polymyxin under clinical development to • treat pneumonia, bloodstream, and urinary tract infections caused by GNB multidrug-resistant (MDR) pathogens.
- The *in vitro* activity of SPR206 and comparator agents was monitored against GNB pathogens causing infection in US and European hospitals during 2021 as part of the SENTRY Antimicrobial Surveillance Program. This study reports the activity of SPR206 against Acinetobacter spp. and *Pseudomonas aeruginosa* recovered from patients hospitalized in European countries and adjacent regions.

Table 1. MIC distribution of SPR206 obtained against Acinetobacter spp. and P. aeruginosa and resistant subsets

Organism/Group	No. and cumulative % of isolates inhibited at MIC (mg/L) of:										MIC	MIC			
(no. of isolates)	≤0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	>64	50 State	90
Acinetobacter spp. (437) ^a	21 (4.8)	162 (41.9)	122 (69.8)	58 (83.1)	28 (89.5)	7 (91.1)	5 (92.2)	6 (93.6)	2 (94.1)	9 (96.1)	7 (97.7)	4 (98.6)	6 (100)	0.12	1
Non-MDR (168)	11 (6.5)	46 (33.9)	60 (69.6)	32 (88.7)	12 (95.8)	3 (97.6)	2 (98.8)	1 (99.4)	1 (100.0)					0.12	0.5
E-MDR (142)	2 (1.4)	46 (33.8)	42 (63.4)	15 (73.9)	5 (77.5)	3 (79.6)	3 (81.7)	3 (83.8)	0 (83.8)	7 (88.7)	6 (93.0)	4 (95.8)	6 (100)	0.12	32
W-MDR (127)	8 (6.3)	70 (61.4)	20 (77.2)	11 (85.8)	11 (94.5)	1 (95.3)	0 (95.3)	2 (96.9)	1 (97.6)	2 (99.2)	1 (100.0)			0.06	0.5
P. aeruginosa (448)	2 (0.4)	17 (4.2)	157 (39.3)	246 (94.2)	21 (98.9)	3 (99.6)	0 (99.6)	0 (99.6)	2 (100.0)					0.25	0.25
Non-MDR (378)	2 (0.5)	11 (3.4)	143 (41.3)	210 (96.8)	8 (98.9)	2 (99.5)	0 (99.5)	0 (99.5)	2 (100.0)					0.25	0.25
E-MDR (33)		3 (9.1)	5 (24.2)	16 (72.7)	8 (97.0)	1 (100.0)								0.25	0.5
W-MDR (37)		3 (8.1)	9 (32.4)	20 (86.5)	5 (100.0)									0.25	0.5

Materials and Methods

Bacterial organisms

- This study included 437 Acinetobacter spp. (see Table 1 for a list of species) and 448 *P. aeruginosa* recovered from patients hospitalized in 35 medical centres in 16 European countries plus Israel and Turkey.
- Only consecutive isolates (1 per patient infection episode) responsible for documented infections according to local criteria were included.
- Bacterial identification was confirmed by standard algorithms supported by matrix-assisted laser desorption ionization-time of flight mass spectrometry (Bruker Daltonics, Bremen, Germany).

Susceptibility testing

- Isolates were tested for susceptibility by broth microdilution following the Clinical and Laboratory Standards Institute (CLSI) M07 (2018) guidelines.
- Frozen-form broth microdilution panels were manufactured by JMI Laboratories (North Liberty, IA, USA) and contained cation-adjusted Mueller-Hinton broth per CLSI guidelines.
- Quality assurance was performed by sterility checks, colony counts, and testing CLSI- and/or EUCAST-recommended quality control reference strains. MIC interpretations for comparators were performed using EUCAST breakpoints. Clinical breakpoints are not available for SPR206, so values throughout indicate the percentage of isolates inhibited by SPR206 at $\leq 2 \text{ mg/L}$ for comparison purposes only.

Resistant subset definitions

• MDR was defined as any isolate resistant to ≥ 3 classes of antibiotics

Abbreviations: MDR, multidrug-resistant (resistant to \geq 3 classes); E-MDR, Eastern Europe MDR isolates; W-MDR, Western Europe MDR isolates.

^a Includes A. baumannii-calcoaceticus species complex (387), A. berezinae (7), A. calcoaceticus (2), A. gerneri (1), A. gyllenbergii (1), A. johnsonii (4), A. junii (9), A. lwoffii (4), A. radioresistens (4), A. soli (1), A. ursingii (13), A. vivianii (1), and Acinetobacter spp. (3).

Table 2. Antimicrobial activity of SPR206 and comparators against Acinetobacter spp. and resistant subsets

		MIC (EUCAST ^a			
Antimicrobial agent	50 %	90%	Range	% S	%	% R
Non-MDR (168)						
SPR206	0.12	0.5	≤0.03 to 8	98.8		
Colistin	0.5	1	0.12 to 8	97.6		2.4
Ampicillin-sulbactam	2	4	≤0.5 to 64			
Meropenem	0.25	1	≤0.015 to 16	99.4	0.0	0.6
Ceftazidime	4	8	0.25 to >32			
Ceftazidime-avibactam	4	16	≤0.015 to >32			
Piperacillin-tazobactam	≤0.06	8	≤0.06 to >128			
Amikacin	2	4	≤0.25 to >32	97.0		3.0
Tobramycin	0.5	1	≤0.12 to >16	97.6		2.4
Tigecycline	0.25	0.5	≤0.06 to 2			
Levofloxacin	0.12	0.25	≤0.015 to 16	95.8	1.8	2.4
E-MDR (142)						
SPR206	0.12	32	≤0.03 to >64	81.7		
Colistin	0.5	>8	0.12 to >8	79.6		20.4
Ampicillin-sulbactam	64	>64	4 to >64			
Meropenem	>32	>32	0.12 to >32	5.6	0.7	93.7
Ceftazidime	>32	>32	32 to >32			
Ceftazidime-avibactam	32	>32	1 to >32			
Piperacillin-tazobactam	>128	>128	32 to >128			
Amikacin	>32	>32	0.5 to >32	9.2		90.8
Tobramycin	>16	>16	≤0.12 to >16	13.4		86.6
Tigecycline	2	8	0.12 to >8			
Levofloxacin	16	>32	0.12 to >32	1.4	0.0	98.6
W-MDR (127)						
SPR206	0.06	0.5	≤0.03 to 32	95.3		
Colistin	0.5	2	0.25 to >8	90.6		9.4
Ampicillin-sulbactam	>64	>64	4 to >64			
Meropenem	>32	>32	0.5 to >32	3.1	0.0	96.9
Ceftazidime	>32	>32	4 to >32			
Ceftazidime-avibactam	>32	>32	4 to >32			
Piperacillin-tazobactam	>128	>128	16 to >128			
Amikacin	>32	>32	1 to >32	13.4		86.6
Tobramycin	>16	>16	0.25 to >16	11.8		88.2
Tigecycline	2	4	0.12 to >8			
Levofloxacin	16	>32	0.25 to >32	1.6	0.0	98.4

Table 3. Antimicrobial activity of SPR206 and comparators against *P. aeruginosa* and resistant subsets

Antimiershiel agent		MIC	(mg/L)	EUCAST ^a			
Antimicropial agent	50 %	90%	Range	% S	%	% R	
Non-MDR (378)							
SPR206	0.25	0.25	≤0.03 to 8	99.5			
Colistin	1	1	≤0.06 to 4	100.0		0.0	
Meropenem	0.5	4	≤0.015 to 32	89.7	9.0	1.3	
Ceftazidime	2	8	0.25 to >32		92.9 ^b	7.1	
Ceftazidime-avibactam	2	4	≤0.015 to 8	100.0		0.0	
Piperacillin-tazobactam	4	16	≤0.06 to >128		92.3 ^b	7.7	
Amikacin	4	8	≤0.25 to >32	98.9		1.1	
Tobramycin	0.5	1	≤0.12 to >16	97.6		2.4	
Levofloxacin	0.5	4	≤0.015 to >32		89.9 ^b	10.1	
Ceftolozane-tazobactam	0.5	1	≤0.12 to 4	100.0		0.0	
E-MDR (33)							
SPR206	0.25	0.5	0.06 to 1	100.0			
Colistin	1	1	0.25 to 2	100.0		0.0	
Meropenem	16	>32	0.5 to >32	12.1	36.4	51.5	
Ceftazidime	32	>32	8 to >32		3.0 b	97.0	
Ceftazidime-avibactam	4	>32	1 to >32	69.7		30.3	
Piperacillin-tazobactam	>128	>128	16 to >128		3.0 b	97.0	
Amikacin	8	>32	1 to >32	63.6		36.4	
Tobramycin	1	>16	0.25 to >16	57.6		42.4	
Levofloxacin	16	>32	0.06 to >32		33.3 ^b	66.7	
Ceftolozane-tazobactam	4	>16	1 to >16	60.6		39.4	
W-MDR (37)				<u> </u>			
SPR206	0.25	0.5	0.06 to 0.5	100.0			
Colistin	1	1	0.12 to 1	100.0		0.0	
Meropenem	16	>32	0.06 to >32	8.1	37.8	54.1	
Ceftazidime	32	>32	8 to >32		13.5 ^b	86.5	
Ceftazidime-avibactam	4	32	2 to >32	73.0		27.0	
Piperacillin-tazobactam	128	>128	2 to >128		5.4 ^b	94.6	
Amikacin	8	>32	1 to >32	78.4		21.6	
Tobramycin	1	>16	0.25 to >16	73.0		27.0	
Levofloxacin	2	32	0.25 to >32		54.1 ^b	45.9	
Ceftolozane-tazobactam	2	>16	1 to >16	67.6		32.4	

based on CLSI criteria.

Results

Acinetobacter spp.

- A total of 61.6% of all *Acinetobacter* spp. included exhibited an MDR phenotype.
 - Clinical isolates originating from hospitals located in Eastern — European countries plus Israel and Turkey had an MDR phenotype prevalence (77.6%) higher than those isolates from Western European hospitals (50.0%) (data not shown).
- Overall, SPR206 had MIC_{50/90} values of 0.12/1 mg/L against all Acinetobacter spp. (Table 1), whereas collistin had $MIC_{50/90}$ results of 0.5/4 mg/L (data not shown).
- Various agents were active (95.8%–99.4% susceptible) against non-MDR Acinetobacter spp., including SPR206 that inhibited all but 2 strains at $\leq 2 \text{ mg/L}$ (Table 2).
- SPR206 (MIC_{50/90}, 0.12/32 mg/L) and collistin (MIC_{50/90}, 0.5/>8 mg/L) MICs against the MDR subset from Eastern Europe were higher than those MICs obtained against isolates from Western Europe (MIC_{50/90}, 0.06/0.5 mg/L for SPR206 and MIC_{50/90}, 0.5/2 mg/L for colistin) (Table 2).
- Comparator agents showed limited activities against isolates _ from Eastern Europe (Table 2).
- SPR206 (MIC_{50/90}, 0.06/0.5 mg/L) exhibited the lowest MIC values when tested against isolates from Western Europe, followed by collistin (MIC_{50/90}, 0.5/2 mg/L). Other agents had limited activities (Table 2).

P. aeruginosa

- Overall, 15.6% of all *P. aeruginosa* showed an MDR phenotype (Table 1).
- Those clinical isolates originating from hospitals located in Eastern European countries plus Israel and Turkey had an MDR phenotype prevalence (24.8%) higher than those isolates from Western European hospitals (11.7%) (data not shown). • In general, MIC_{50/90} values of 0.25/0.25 mg/L were obtained for SPR206 against all *P. aeruginosa* (Table 1), a 4-fold lower result than achieved for collistin (MIC_{50/90}, 1/1 mg/L; 100% susceptible) (data not shown). • Various agents were active against non-MDR *P. aeruginosa*, although SPR206 (MIC_{50/90}, 0.06/0.5 mg/L) exhibited, respectively, MIC₅₀ and MIC₉₀ values at least 2-fold and 4-fold lower than the comparators (Table 3). SPR206 (MIC_{50/90}, 0.25/0.5 mg/L) and collistin (MIC_{50/90}, 1/1 mg/L) showed the lowest MIC₅₀ and MIC₉₀ results against the *P. aeruginosa* MDR subsets, whereas the other comparators had limited activities (Table 3).

Abbreviations: MDR, multidrug-resistant (resistant to \geq 3 classes); E-MDR, Eastern Europe MDR isolates; W-MDR, Western Europe MDR isolates

^a Criteria as published by EUCAST (2022). A susceptible breakpoint of $\leq 2 \text{ mg/L}$ was used for SPR206 for comparison purposes.

Abbreviations: MDR, multidrug-resistant (resistant to \geq 3 classes); E-EU, Eastern Europe MDR isolates; W-EU, Western European MDR isolates.

^a Criteria as published by EUCAST (2022). Clinical breakpoints are not available for SPR206, so values indicate the percentage of isolates inhibited by SPR206 at $\leq 2 \text{ mg/L}$ for comparison purposes only. ^b Isolates can be considered as susceptible if a higher drug concentration can be achieved by increasing the dosing regimen or increased concentration is achieved in the infection site.

Conclusions

- SPR206 showed potent *in vitro* activity against these recent collections of Acinetobacter spp. and P. aeruginosa from Europe and inhibited 92.2% and 99.6% of all isolates, respectively.
- In addition, the potency of SPR206 was consistently greater than clinically available in-class and other comparator agents.
- These SPR206 results, plus favourable safety and tolerability profiles obtained during Phase 1 studies, support the clinical development of SPR206 for difficult-to-treat infections caused by these pathogens and their resistant subsets.

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Contact

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References

1. Bruss J, Lister T, Gupta VK, Stone E, Morelli L, Lei Y, Melnick D (2021). Single- and multiple-ascending-dose study of the safety, tolerability, and pharmacokinetics of the polymyxin derivative SPR206. Antimicrob. Agents Chemother. 65 (10): e0073921.

2. Brown P, Abbott E, Abdulle O, Boakes S, Coleman S, Divall N, Duperchy E, Moss S, Rivers D, Simonovic M, Singh J, Stanway S, Wilson A, Dawson MJ (2019). Design of next generation polymyxins with lower toxicity: The discovery of SPR206. ACS Infect Dis. 5 (10): 1645-1656.

3. Clinical and Laboratory Standards Institute (2018). Methods for dilution antimicrobial susceptibility tests for bacteria that grow aerobically. MO7Ed11. Wayne, PA, USA.

4. Clinical and Laboratory Standards Institute (2022). Performance standards for antimicrobial susceptibility testing. M100Ed32. Wayne, PA, USA.

5. EUCAST. The European Committee on Antimicrobial Susceptibility Testing. Breakpoint tables for interpretation of MICs and zone diameters, v. 12.0, 2022. http://www.eucast.org.

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