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VITEK2 Advanced Expert System β-Lactam Resistance Phenotyping Compared to Whole Genome Sequencing in Enterobacterales Isolates from European Medical Centres

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Introduction

- The rapid detection of β -lactam resistant phenotypes such as transferable AmpC (tAmpC), ESBL, and carbapenemase are important for appropriate antimicrobial therapy administration and infection control.
- The VITEK2 Advanced Expert System (AES) provides interpretations of β-lactam resistance phenotypes based on an extensive database of MIC distributions and prevalent resistance mechanisms in Enterobacterales isolates.
- In this study, the AES β-lactam resistance phenotypes were compared to whole genome sequencing results from 572

Table 1. VITEK2 AES β -lactam resistance phenotypes compared to whole genome sequencing genotypes.

β-lactam resistance	AES phenotypic report		
genotype (No. of isolates)	Accuracy	Sensitivity	Specificit
Carbapenemase (212)	96.5%	99.5%	94.6%
ESBL (161)	98.6%	97.5%	99.0%
tAmpC (51)	97.9%	84.3%	99.2%
WT (140)	99.8%	100.0%	99.8%

Abbreviations: ESBL, extended-spectrum β -lactamase; tAmpC, transferrable AmpC; WT, wildtype.

Table 2. List of discordances between VITEK2 AESphenotype and WGS genotype

WGS genotype	AES phenotype	No. of occurrences
Carbapenemase	ESBL	1
ESBL	AmpC	4
tAmpC	Carbapenemase	4
	ESBL	3
	WT	1

Abbreviations: ESBL, extended-spectrum β -lactamase; tAmpC, transferrable AmpC; WT, wildtype.

Materials and Methods

- Isolates were collected from 39 medical centres in 19 European countries as part of the SENTRY Antimicrobial Surveillance Program during 2017–2020 (Figure 1).
- A total of 251 *Klebsiella pneumoniae*, 174 *Escherichia coli*, 48 *Enterobacter cloacae* species complex, and 99 other Enterobacterales isolates were tested (Figure 2A).
- Antimicrobial susceptibility testing was performed by VITEK2 AES v9.02 under Global European + Phenotypic mode, using the N388 and XN11 susceptibility cards (Iberic cards) and these results were compared to reference broth microdilution outcomes.
- The following β -lactam and β -lactam/ β -lactamase inhibitor combination agents were tested:
 - Amoxicillin/clavulanate, ampicillin, cefepime, cefixime, cefotaxime, cefoxitin, ceftazidime, ceftazidime/avibactam, ceftolozane/tazobactam, ceftriaxone, cefuroxime, ertapenem, imipenem, and mecillinam.
- EUCAST breakpoints were applied, except for compounds where AES criteria were not compatible with most recent EUCAST criteria.
- Discordant results were repeated by both methods using the same bacterial inoculum.
- Whole genome sequencing (WGS) was performed on isolates that met the following criteria by BMD:
 - *E. coli* and *K. pneumoniae* isolates displaying MIC values $\geq 2 \text{ mg/L}$ for at least 2 of the following β -lactams: aztreonam, cefepime, ceftazidime, or ceftriaxone; and/or

Figure 1. Distribution of the 572 Enterobacterales isolates included in this study stratified by country.



Figure 2. Characterization of Enterobacterales isolates included in the study.

A. By organism (572 isolates)

B. By genotype (572 isolates)

Other Enterobacterales* —

- Enterobacterales isolates displaying meropenem and/or imipenem MIC values >1 mg/L.
- Enterobacterales isolates that did not meet the criteria for molecular characterization were considered wildtype.
- The accuracy, sensitivity, and specificity of AES reports for β-lactam resistant phenotypes were compared to resistant genotypes confirmed by WGS.



*Organisms included: Proteus spp. (25 isolates), Citrobacter spp. (24), Serratia marcescens (15), Klebsiella aerogenes (11), Klebsiella oxytoca (9), Providencia spp. (6), and Morganella morganii (3).

Results

- Figure 2B shows the distribution of the 572 Enterobacterales isolates displaying carbapenemase, ESBL, tAmpC-encoding genes and wildtype genotypes.
- AES provided phenotypic reports for 564 (98.6%) isolates, including isolates harbouring carbapenemase (212; 37.6%), extended-spectrum β-lactamase (ESBL; 161; 28.5%), and transferable AmpC (tAmpC; 51; 9.0%) genes as well as wildtype (WT; 140; 24.8%) isolates.
- Eight of 572 isolates (1.4%) failed to report an AES phenotype due to technical error or because the organism expressed a phenotype that was not present in the AES knowledge base.
- Overall, the AES report was accurate for 551/564 isolates (97.7%; Table 1).
- AES accurately reported carbapenemase, ESBL, and tAmpC phenotypes for 96.5%, 98.6%, and 97.9% of isolates, respectively.
- All but 1 (99.8%) WT isolate was correctly categorized by AES, including when isolates displayed intrinsic resistance or an acquired penicillinase.

Conclusions

- VITEK2 AES provided β-lactam resistance phenotypes for 98.6% (564/572) of isolates from a large and challenging collection of Enterobacterales from Europe.
- AES β -lactam resistance phenotypes were correctly reported for 97.7% (551/564) of Enterobacterales isolates harbouring a variety of β -lactamase genes, including 96.5%, 98.6%, and 97.9% of carbapenemase, ESBL, and tAmpC, respectively.
- The AES phenotypic report could significantly aid antimicrobial stewardship initiatives and improve patient care if it was used in clinical laboratories as a rapid tool for the detection of resistance mechanisms among Enterobacterales from Europe.

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- AES sensitivity/specificity rates were 99.5%/94.6%, 97.5%/99.0%, 84.3%/99.2%, and 100%/99.8% for reporting carbapenemase, ESBL, tAmpC genes, and WT isolates, respectively (Table 1).
- Table 2 displays the discrepancies between the AES phenotype and genotype, including 8 isolates carrying tAmpC, 1 carbapenemase, and 4 ESBL genes by WGS.
- Additionally, 4 isolates harbouring ESBL were reported as AmpC, and 1 VIM-1–producing *E. coli* isolate was misreported as displaying an ESBL phenotype.

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