

Activity of Manogepix and Comparator Antifungal Agents Against Yeast Isolates Collected from Invasive Infections in Intensive Care Unit (ICU) and Non-ICU Settings

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Introduction

- The occurrence of invasive candidiasis among patients hospitalized in Intensive Care Units (ICUs) can be as high as 50%.
- Despite its high clinical impact, relatively few studies describe the species distribution of *Candida* and the antifungal resistance profiles associated with invasive candidiasis in ICU settings compared to non-ICU settings, highlighting the need for surveillance data and comparative analyses.
- We evaluated the species distribution according to ICU versus non-ICU location in a global yeast collection, and the associated resistance profiles for the contemporary echinocandin and azole antifungal agents and compared them to the activity of manogepix.

Materials and Methods

- 7,266 non-duplicate yeast isolates deemed as cause of invasive infections were collected in 91 hospitals during 2017–2024.
- All isolates were identified by MALDI-TOF MS and/or ITS and 28S sequencing.
- All isolates were tested by the CLSI reference broth microdilution method following the M27 guidelines.
- Breakpoints used for antifungal susceptibility interpretation were applied according to the CLSI M27M44S document.
- Epidemiological cutoff values (ECVs) were obtained from the CLSI M57 document.

Table 1. Activity of clinically available antifungals against main yeast species in ICU and non-ICU settings

Antifungal agent ^a	ICU ^b		Non-ICU ^b	
	%R ^c	%NWT ^d	%R ^c	%NWT ^d
<i>Candida albicans</i> (N=828 ICU, N=1297 Non-ICU)				
Micafungin	0.2	0.7	0.4	0.8
Caspofungin	0.2	0	0.4	0
Fluconazole	0.1	2.2	0.4	2.8
Voriconazole	0.1	1.1	0	1.5
Amphotericin B		0		0
<i>Candida glabrata</i> (N=666 ICU, N=1017 Non-ICU)				
Micafungin	3	5	1.7	3
Caspofungin	2.3		1.3	
Fluconazole	7.5	11.6	6.2	11.2
Voriconazole		10.2		9.5
Amphotericin B		0		0
<i>Candida parapsilosis</i> (N=433 ICU, N=787 Non-ICU)				
Micafungin	0	0	0	0
Caspofungin	0	0.2	0	0
Fluconazole	15.2	18.9	8.8	11.6
Voriconazole	3	19.6	1.5	12.3
Amphotericin B		0.5		0.4
<i>Candida tropicalis</i> (N=276 ICU, N=497 Non-ICU)				
Micafungin	0	0.4	0	1.4
Caspofungin	0		0	
Fluconazole	5.1	8	2.6	4.6
Voriconazole	4	5.8	1.4	2.8
Amphotericin B		0		0
<i>Candida dubliniensis</i> (N=92 ICU, N=143 Non-ICU)				
Micafungin		2.2		0.7
Fluconazole		0		3.5
Amphotericin B		0		1.4
<i>Candida krusei</i> (N=87 ICU, N=131 Non-ICU)				
Micafungin	0	0	0.8	0.8
Caspofungin	0		0.8	
Voriconazole	1.1	3.4	0	1.5
Amphotericin B		0		0
<i>Candida lusitanae</i> (N=68 ICU, N=90 Non-ICU)				
Micafungin		0		0
Caspofungin		0		0
Fluconazole		10.3		13.3
Amphotericin B		0		0
<i>Candida auris</i> (N=61 ICU, N=72 Non-ICU)				
Micafungin	6.6	9.8	1.4	2.8
Caspofungin	13.1	13.1	6.9	8.3
Fluconazole	90.2		93.1	
Amphotericin B	45.9		37.5	
<i>Candida orthopsilosis</i> (N=34 ICU, N=46 Non-ICU)				
Micafungin		0		0
Caspofungin		0		0
Fluconazole		14.7		6.5
Voriconazole		14.7		2.2
Amphotericin B		0		0
<i>Candida guilliermondii</i> (N=10 ICU, N=30 Non-ICU)				
Micafungin	0	0	0	0
Caspofungin	0	0	0	3.4
Fluconazole		40		16.7
Amphotericin B		0		0

^a Representative antifungal agents with established breakpoints and/or ECV criteria are shown
^b Values in red display difference of ± 0.5 when comparing ICU and non-ICU isolates
^c Breakpoint criteria published by CLSI M27M44S (2022)
^d ECV criteria published in CLSI M57S (2022)

Results

- A total of 2,744 isolates from ICU and 4,522 from non-ICU were evaluated.
- The frequency of different species by ICU versus non-ICU displayed minor differences.
- In both cases 83–84% of the infections were caused by the top 5 species—*C. albicans*, *C. glabrata*, *C. parapsilosis*, *C. tropicalis* and *C. dubliniensis*.
- The 10 most common species are displayed in Figure 1.
- Resistance to echinocandins was uncommon and similar in ICU and non-ICU settings for *C. albicans*, *C. parapsilosis*, *C. tropicalis*, *C. orthopsilosis* and *C. guilliermondii* ($\leq 0.4\%$; Table 1).
- Echinocandin resistance rates were higher overall and increased in the ICU setting for *C. glabrata* (3.0% vs. 1.7% for micafungin in the ICU vs. non-ICU) and *C. auris* (6.6% vs. 1.4%).
- Fluconazole resistance was noted among *C. glabrata* (7.5% vs. 6.2%), *C. parapsilosis* (15.2% vs. 8.8%), *C. tropicalis* (5.1% vs. 2.6%) and *C. orthopsilosis* (14.7% vs. 6.5%) with higher rates in ICU vs. non-ICU isolates.
- C. auris* isolates displayed high fluconazole-resistance rates in both settings (90.2–93.1%) and resistance to amphotericin B was higher among ICU (45.9%) compared to non-ICU isolates (37.5%).
- Manogepix (MIC₅₀, 0.004–0.03 mg/L and MIC₉₀, 0.008–0.25 mg/L) was very active against the main *Candida* spp. isolates tested regardless of ICU status (Figure 2) for all species but *C. krusei* and *Trichosporon asahii* (MIC_{50/90}, $\geq 1/>2$ mg/L).

Conclusions

- Yeast species distributions were similar between ICU and non-ICU settings.
- Among species displaying resistance to echinocandins and/or fluconazole, rates were higher among ICU compared to the non-ICU isolates.
- Manogepix activity was unchanged by ICU status, and this evidence supports the further development of this agent.

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References

Pfaller MA, Messer SA, Moet GJ, Jones RN, Castanheira M. *Candida* bloodstream infections: comparison of species distribution and resistance to echinocandin and azole antifungal agents in Intensive Care Unit (ICU) and non-ICU settings in the SENTRY Antimicrobial Surveillance Program (2008–2009). *Int J Antimicrob Agents*. 2011 Jul;38(1):65-9.
 CLSI. 2017. M27 Ed4. Reference method for broth dilution antifungal susceptibility testing of yeasts. Clinical and Laboratory Standards Institute, Wayne, PA.
 CLSI. 2022. M27M44S Ed3. Performance standards for antifungal susceptibility testing of yeasts. Clinical and Laboratory Standards Institute, Wayne, PA.
 CLSI. 2022. M57S Ed4. Epidemiological cutoff values for antifungal susceptibility testing. Clinical and Laboratory Standards Institute, Wayne, PA.

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Figure 1. Distributions of 10 most common yeast species in ICU and non-ICU settings

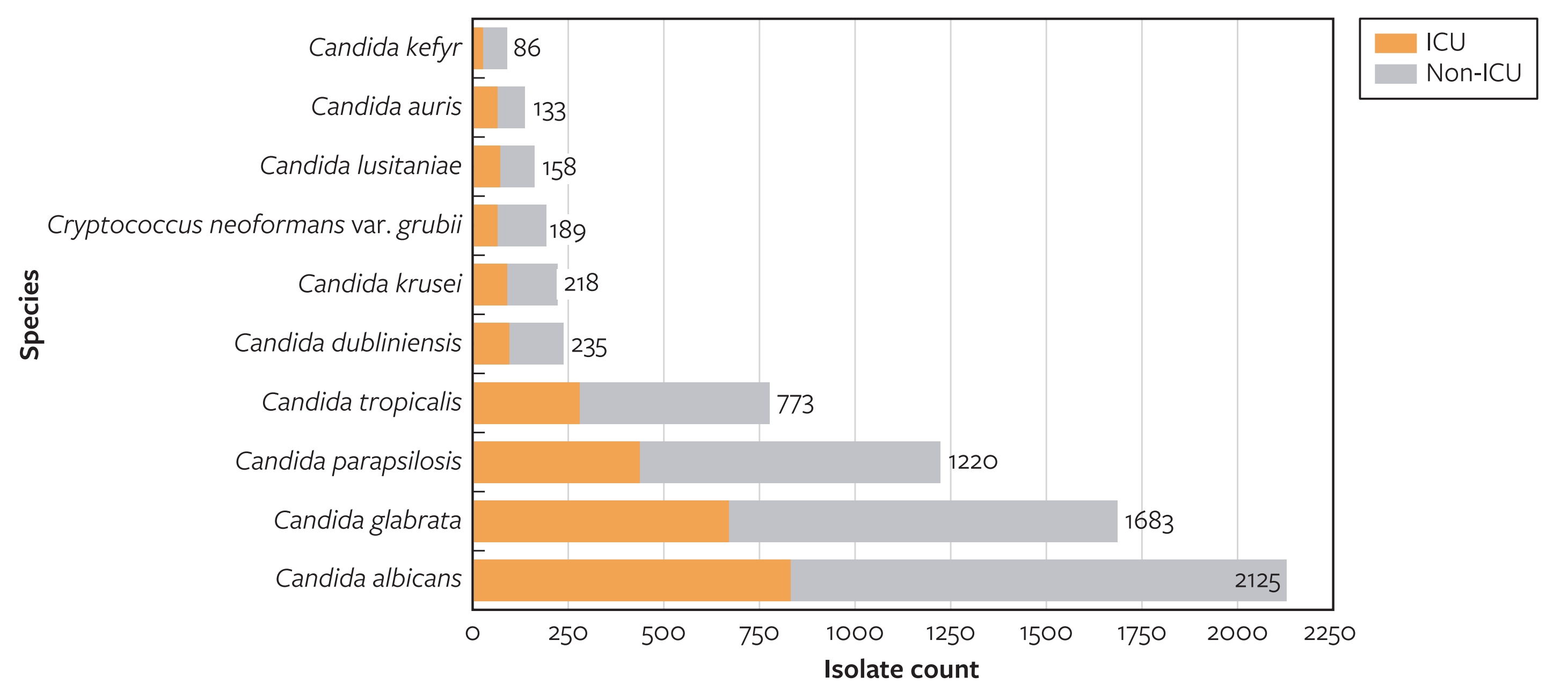


Figure 2. Activity of manogepix tested against main yeast species and *C. auris* from ICU vs non-ICU

