In Vitro Antibacterial Spectrum of BC-3205, a Novel Pleuromutilin Derivative for Oral Use in Humans

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Abstract

Background: BC-3205 [14-O-[(N-((R)-ValyI)-piperidin-3(S)-yI)sulfanyI)-acetyI]-mutilinhydrochloride] is a novel semi-synthetic pleuromutilin derivative that inhibits prokaryotic protein synthesis. BC-3205 is in early stage of clinical development for oral treatment of skin and skin structure infections (SSSI) and community-acquired pneumonia (CAP). We assessed BC-3205 antimicrobial activity against a wide range of clinical isolates.

Methods : BC-3205 and comparators were susceptibility (S) tested by CLSI broth microdilution methods (M07-A8, 2009) against 1063 recent clinical isolates (mainly from 2006-2007) from the USA and Europe that were collected from the SENTRY Antimicrobial Surveillance Program.

Results: BC-3205 was highly active against Staphylococcus spp. and Streptococcus spp. including methicillin-resistant Staphylococcus aureus (MRSA) (MIC₂₀ 0.12 µg/ml), communityacquired (CA-MRSA) (MICon 0.06 µg/ml) and penicillin-resistant Streptococcus pneumoniae (PEN-R SPN) (MICon 0.12 µg/ml). Against S. aureus, BC-3205 was 16- to 32-fold more potent than linezolid (LZD). Against PEN-R SPN, BC-3205 was 8-fold more active than levofloxacin (LEV) or LZD. BC-3205 exhibited significant activity against vancomycin (VAN)-resistant (R) E. faecium (Efm) (MIC₉₀ 0.5 µg/ml), S. pyogenes (MIC₉₀ 0.03 µg/ml), S. agalactiae (MIC₉₀ 0.06 µg/ml), H. influenzae (MICon 4 µg/ml) and M. catarrhalis (MICon range, 0.25 µg/ml).

Conclusions: BC-3205 exhibited potent antimicrobial activity against the most prevalent Gram-positive pathogens involved in SSSI and pathogens causing CAP, being more active against staphylococci and streptococci than many antimicrobials currently available for oral use.

Introduction

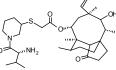
BC-3205 is a novel semi-synthetic pleuromutilin derivative (Figure 1). Pleuromutilin-derived antibiotics are inhibitors of ribosomal protein synthesis interacting with the rRNA of the ribosomal peptidyl transferase cavity. More specifically, foot-printing analysis revealed that pleuromutilins interfere with the central part of domain V of the 23S rRNA, where they prevent the correct positioning of the CCA-ends of tRNAs for peptide transfer and where they are thought to block translation initiation 1-5

BC-3205 is being developed for oral treatment of skin and skin structure infections (SSSI) and community-acquired pneumonia (CAP) as it exhibits potent antibacterial activity against common bacterial pathogens causing SSSI and CAP including Staphylococcus aureus (MSSA, MRSA and CA-MRSA), Streptococcus pyogenes, S. agalactiae, S. pneumoniae, Haemophilus influenzae, Moraxella catarrhalis, Chlamydophila pneumoniae, Mycoplasma pneumoniae and Legionella pneumophilia.

Of increasing concern is the rising frequency of SSSI caused by MRSA. Being highly endemic in certain geographic regions, especially multi-resistant phenotypes and the high prevalence of community-acquired MRSA (CA-MRSA), in particular the highly virulent clone USA300, are considered to be a substantial threat to public health.

In this study we present the antibacterial activity of BC-3205 against predominant pathogens causing SSSI and CAP including multi-resistant clinical isolates and CA-MRSA.

Figure 1: Structure of BC-3205



Methods

Organism Collection: The activity of BC-3205 was determined against bacterial pathogens causing SSSI and CAP. Organisms included strains isolated from patients in either the United States (USA, 52.3%) or European (47.6%) medical centers (n=67) during 2006 to 2007. Less commonly isolated species and those organisms with characterized resistance mechanism were included from 2005 (7.8%), 2004 (4.7%) and 2000-2003 (7.2%). Most of the isolates were from patients with bloodstream infections (63.1%), respiratory tract infections (29.6%) and skin and skin structure infections (5.6%)

Organisms examined (n=1013) included methicillin-susceptible S. aureus (MSSA, 151), methicillin-resistant S. aureus (MRSA, 102), community-acquired MRSA (CA-MRSA, 50) S. pneumoniae (SPN, 157), penicillin-resistant S. pneumoniae (PEN-R SPN, 52). S. progenes (50), Vancomycin-susceptible E. faecium (VAN-S Efm. 51), vancomycin-resistant E. faecium (VAN-R Efm, 51), H. influenzae (102), M. catarrhalis (50), C. pneumoniae (2), Mycoplasma spp. (6) and L. pneumophilia (30).

Susceptibility test methods: MIC values of SSSI and typical CAP pathogens were determined according to CLSI guidelines (M07-A8, 2009 and M100-S19, 2009).⁶ 96-well format panels were produced by JMI Laboratories using Mueller Hinton broth, cation adjusted MHB supplemented with 2-5% lysed horse blood for testing Streptococcus spp. or Haemophilus Test Medium for testing H. influenzae. Interpretive criteria for comparator antibiotics were used as published by CLSI (M100-S19, 2009).7 Susceptibility of C. pneumoniae, Mycoplasma spp. and L. pneumophilia was determined by Nabriva Therapeutics and as described earlier. 8, 9, 7

Quality control was performed as recommended by the CLSI using the following strains: S. aureus ATCC 29213, S. pneumoniae ATCC 49619, H. influenzae ATTC 49247 and E. faecalis ATCC 29212.

Table 1. Antibacterial spectrum of BC-3205 and comparator antibiotics

Species	MIC₀₀ [µg/mi]									
opecies	n BC-3205		LZD	AZI	CLI	LEV				
MSSA	151	0.06	2	8	0.12	0.25				
MRSA	102	0.12	2	>16	>16	>16				
CA-MRSA ^a	50	0.06	2	>16	0.12	0.5				
CoNS	105	0.12	2	>16	>16	>16				
SPN	157	0.12	1	>16	>16	1				
PEN-R SPN	52	0.12	1	>16	>16	1				
S. pyogenes	50	0.03	1	0.12	0.06	0.5				
VAN-S Efm	51	4	2	>16	>16	>16				
VAN-REfm	51	0.5	2	>16	>16	>16				
H. influenzae	102	4	-	2	-	≤0.06				
M. catarrhalis	50	0.25	-	≤0.5	-	≤0.06				
C. pneumoniae b	2	0.01-0.04	-	0.08-0.16	-	-				
Mycoplasma spp.b.c	6	≤0.0003-0.04	-	0.00015-6.4	0.2-0.8	-				
L. pneumophilia d	30	0.5	-	0.12	-	-				

Abbreviations: AZI, azithromycin; CLI, clindamycin; LEV, levofloxacin; LZD, linezolid;

MSSA, methicillin-susceptible S. aureus; MRSA, methicillin-resistant S. aureus; CA-MRSA, community-acquired MRSA; CoNS, coagulase negative Staphylococcus spp.

SPN, S. pneumoniae; PEN-R SPN, penicillin-resistant SPN; VAN-S Efm, vancomycin-susceptible E. faecium, VAN-R Efm, vancomycin-resistant E. faecium

* CA-MRSA include USA 300 clone (45) and USA 400 clone (5); * Range is shown instead of MICm · M. pneumoniae (n=4), M. hominis (n=1), M. genitalium (n=1); dMICs determined by microbroth dilution using charcoal supplemented BCYEg medium

Results

- The antibacterial activity of BC-3205 and comparator antibiotics is presented in Table 1 and the cumulative percentage of strains inhibited by BC-3205 is shown in Table 2.
- With a MIC₅₀ value of 0.06 µg/ml and a MIC₉₀ value of 0.12 µg/ml, BC-3205 was the most potent compound tested against S. aureus; 100.0% of isolates were inhibited by ≤0.12 µg/ml
- The CA-MRSA isolates (USA300 and USA400 clones) resistant to macrolides and susceptible to the other tested antimicrobial classes were inhibited at BC-3205 concentrations of 0.06 -0.12 µg/ml, identical to that of MSSA isolates.
- S. pyogenes and other ß-hemolytic streptococci were highly susceptible to BC-3205 (S. pyogenesMIC₉₀, 0.03 µg/ml) with 100% being inhibited at 0.12 µg/ml

· BC-3205 was also the most potent compound tested against CoNS with MIC90 (0.12 µg/ml) values being the same as for S. aureus.

Table 2, MIC frequency distributions of BC-3205 against 1013 bacterial isolates

Organism (no. tested)	Cumulative percentage of strains inhibited at each MIC [µg/m1]												
	≤0.008	0.015	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	>16
S. aureus (303)		-	0.3	87.5	100.0	-	-	-	-	-	-	-	-
Oxacillin-susceptible (151)	-	-	-	90.7	100.0	-	-	-	-	-	-	-	-
Oxacillin-resistant (102)	-	-	1.0	78.4	100.0	-	-	-	-	-	-	-	-
CA-Oxacillin-resistant (50)	-	-	-	96.0	100.0	-	-	-	-	-	-	-	-
oagulase-negative staphylococci (105)	-	-	3.8	71.4	90.5	93.3	95.2	96.2	96.2	96.2	99.1	99.1	100.
Oxacillin-susceptible (50)	-	-	6.0	88.0	98.0	98.0	98.0	98.0	98.0	98.0	100.0	-	-
Oxacillin-resistant (55)	-	-	1.8	56.4	83.6	89.1	92.7	94.6	94.6	94.6	98.2	98.2	100.
. faecium (102)	-	1.0	6.9	65.7	79.4	83.3	85.3	87.3	89.2	95.1	98.0	100.0	-
Vancomycin-susceptible (51)	-	-	2.0	54.9	70.6	78.4	78.4	78.4	82.4	92.2	98.0	100.0	-
Vancomycin-resistant (51)	-	2.0	11.8	76.5	88.2	88.2	92.2	96.1	96.2	98.0	98.0	100.0	-
Interococcus spp. (22)	4.6	4.6	9.1	31.8	31.8	31.8	40.9	40.9	40.9	59.1	77.3	90.9	100.0
S. pneumoniae (157)	0.6	1.9	20.4	80.3	100.0	-	-	-	-	-	-	-	-
Penicillin-susceptible (54)	-	1.9	13.0	57.4	100.0	-	-	-	-	-	-	-	-
Penicillin-intermediate (51)	2.0	3.9	35.3	96.1	100.0	-	-	-	-	-	-	-	-
Penicillin-resistant (52)	-	-	13.5	88.5	100.0	-	-	-	-	-	-	-	-
'iridans group streptococci. (20)	5.0	15.0	55.0	85.0	100.0	-		-	-	-	-	-	-
Beta-hemolytic streptococci (152)	-	3.3	68.4	98.0	100.0	-	-	-	-	-	-	-	-
I. influenzae (102)	-	-	-	-	-	1.0	2.9	48.0	87.3	100.0	-	-	-
Beta-lactamase negative (51)	-	-	-	-	-	-	-	43.1	92.2	100.0	-	-	-
Beta-lactamase positive (51)	-	-	-	-	-	2.0	5.9	52.9	82.4	100.0	-	-	-
1. catarrhalis (50)	-	-	2.0	32.0	84.0	100.0		-	-	-	-	-	-

- potent against penicillin-susceptible (MIC₉₀, 0.12 µg/mI), -intermediate (MIC₉₀, 0.06 µg/ml) and -resistant (MIC₉₀, 0.12 µg/ml) isolates. Overall, BC-3205 was the most potent agent tested against this species BC-3205 was also shown to be active against the fastidious Gram-negative
- respiratory pathogens H. influenzae (MIC₉₀, 4 µg/mI) and M. catarrhalis (MIC₉₀, 0.25 µg/ml) irrespective of ß-lactamase production.
- BC-3205 exhibited potent activity against C. pneumoniae (MIC range,0.01-0.04) μ g/ml), Mycoplasma spp. (MIC range $\leq 0.0003 - 0.04 \mu$ g/ml) and L. pneumophilia (MIC₉₀, 0.12 µg/mI), .
- · BC-3205 also exhibited potent activity against vancomvcin-susceptible and -resistant E. faecium with >90% of isolates being inhibited at ≤4 µg/ml whereas it was less active against E. faecalis (MIC₉₀, >16 µg/mI). Resistance to vancomycin had no effect on the MIC values obtained for BC-3205.
- BC-3205 was fully active against bacterial strains resistant to macrolides, clindamycin, fluoroquinolones and vancomycin.

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All S. pneumoniae were inhibited by ≤0.12 µg/ml of BC-3205, which was equally

Conclusions

- · BC-3205 shows promising activity against the most prevalent Grampositive pathogens producing skin and skin structure infections, and also Gram-negative pathogens causing community-acquired respiratory tract infections.
- BC-3205 was very active against S. aureus. methicillin-susceptible and -resistant isolates including CA-MRSA, S. pyogenes, S. pneumoniae and other streptococcal species with 100% of strains being inhibited $\leq 0.12 \,\mu g/ml$.
- Modest activity was found against H. influenzae and E. faecium.
- · BC-3205 exhibited potent activity against the atypical pathogens: C. pneumoniae. M. pneumoniae and L. pneumophilia.
- No cross-resistance was observed with macrolides. lincosamides. fluoroquinolones and vancomycin.
- · Overall, BC-3205 showed an activity profile being superior to many antibiotics currently in use for oral treatment of SSSI and CAP.

Selected References

- 1. Bosling, J., Poulsen, S. M., Vester, B., and Long, K. S. Resistance to the peptidyl transferase inhibitor tiamulin caused by mutation of ribosomal protein 13. Antimicrob. Agents Chemother. 47(9), 2892 (2003).
- 2. Davidovich, C., Bashan, A., uerbach-Nevo, T., Yaggie, R. D., Gontarek, R. R., and Yonath, A. Induced-fit tightens pleuromutilins binding to ribosomes and remote interactions enable their selectivity. Proc. Natl. Acad. Sci. U. S. A 104(11), 4291 (2007)
- 3. Long, K. S., Hansen, L. H., Jakobsen, L., and Vester, B. Interaction of pleuromutilin derivatives with the ribosomal peptidyl transferase center. Antimicrob. Agents Chemother. 50(4), 1458 (2006),
- 4. Poulsen, S. M., Karlsson, M., Johansson, L. B., and Vester, B. The pleuromutilin drugs tiamulin and valnemulin bind to the RNA at the peptidyl transferase centre on the ribosome. Mol. Microbiol. 41(5), 1091 (2001).
- 5. Schlunzen, F., Pyetan, E., Fucini, P., Yonath, A., and Harms, J. M. Inhibition of peptide bond formation by pleuromutilins; the structure of the 50S ribosomal subunit from Deinococcus radiodurans in complex with tiamulin. Mol. Microbiol. 54(5), 1287 (2004).
- 6. Clinical and Laboratory Standards Institute. (2009) M100-S19, Performance standards for antimic robial susceptibility testing, 19th informational supplement. Wayne PA: CLSI
- 7. Clinical and Laboratory Standards Institute. (2009) M07-A8, Methods for dilution antimicrobial susceptibility tests for bacteria that grow aerobically, approved standard seventh edition, Wavne, PA: CLSI
- 8. Hammerschlag, M. R. 1999, Activity of guinolones against Chlamydia pneumoniae. Drugs 58 Suppl 2:78-81.
- 9. Ridgw ay, G. 2001. Antimicrobial susceptibility testing of intracellular and cell-associated pathogens. EUCAST Discussion document E.Dis 6.1.