ICAAC 2012 E-189

Antimicrobial Spectrum and Potency of Ceftaroline-Avibactam Tested Against ESBL-Phenotype and Carbapenem-Resistant Enterobacteriaceae Collected from USA Hospitals (2009-2011) HS SADER, M CASTANHEIRA, RK FLAMM, RN JONES JMI Laboratories, North Liberty, Iowa, USA

Amended Abstract

Background: Ceftaroline is a new broadspectrum cephalosporin with bactericidal activity against Gram-positive bacteria (including MRSA) and Enterobacteriaceae (ENT). Avibactam is a novel non- β -lactam β -lactamase (BL) inhibitor that inhibits Ambler class A, C, and some D enzymes (eg, ESBL, KPC, and AmpC).

Methods: Ceftaroline-avibactam and comparators were tested for susceptibility (S) by CLSI broth microdilution methods against 7789 ENT, including ESBL-phenotype E. coli (285) and Klebsiella spp. (KSP; 411), AmpC derepressed *Enterobacter* spp. (ESP; 155) and meropenem (MER)-non-S KSP (most KPC-producing; 115) and ESP (14), among other resistance (R) phenotypes. The strains were consecutively collected in 2009-2011 from 72 USA medical centers.

Results: 97.9% of strains were inhibited at ceftaroline-avibactam MIC of $\leq 0.5 \,\mu g/mL(S)$ breakpoint for ceftaroline alone). Ceftarolineavibactam was very active against ESBLphenotype *E. coli* (MIC_{50/90}, 0.06/0.25 μ g/mL) and KSP (MIC_{50/90}, 0.12/1 µg/mL), MER-non-S KSP (MIC_{50/90}, 0.5/1 µg/mL) and ESP (MIC_{50/90}, 0.5/2 µg/mL), and ceftazidime (CAZ)-non-S ESP (MIC_{50/90}, 0.25/1 µg/mL). ESBL-phenotype strains showed low S to gentamicin (GEN; 62.0-69.1%) and levofloxacin (20.4-39.3%). Only 72.0 and 74.3% of ESBL-phenotype KSP were S to MER and amikacin, respectively. MER-non-S Klebsiella spp. strains exhibited low S to all agents, except tigecycline (MIC_{50/90}, 0.5/2 µg/mL; 95.7% S by CLSI criteria). 78.7% of CAZ-non-S ESP were S to GEN.

Conclusions: Avibactam can effectively lower ceftaroline MIC values for ENT producing the most clinically significant BLs found in USA hospitals. Ceftaroline-avibactam was highly active against ENT producing KPC, various ESBL types, and AmpC (chromosomally derepressed or plasmid-mediated) enzymes. Ceftarolineavibactam shows potent *in vitro* activity against pathogens associated with infections caused by multidrug-R ENT.

Introduction

Ceftaroline fosamil is the parenterally-administered prodrug form of ceftaroline, a new cephalosporin with potent activity against Gram-positive organisms, including methicillin-resistant Staphylococcus aureus (MRSA) and multidrug-resistant (MDR) Streptococcus pneumoniae. In Phase 3 trials (NCT00424190 [CANVAS 1], NCT00423657 [CANVAS 2], NCT00621504 [FOCUS 1] and NCT00509106 [FOCUS 2]), ceftaroline fosamil was shown to be non-inferior to comparator agents for the treatment of patients with acute bacterial skin and skin-structure infections (ABSSSIs) and community-acquired bacterial pneumonia (CABP) requiring hospitalization. Ceftaroline fosamil has been approved by the United States Food and Drug Administration (USA-FDA) for treatment of ABSSSIs and CABP.

Ceftaroline is also active against most Enterobacteriaceae species but, like other cephalosporins, has limited activity against isolates producing extended-spectrum β -lactamases (ESBL), cephalosporinases and carbapenemases. Avibactam (formerly, NXL104) is a novel non- β -lactam β -lactamase inhibitor currently in clinical development. Avibactam has very limited intrinsic antibacterial activity, but efficiently protects β-lactams from hydrolysis by a variety of strains producing Ambler class A and class C enzymes, including ESBL and KPC enzymes (carbapenemases), as well as some class D β -lactamases. We report the in vitro activity of ceftaroline combined with avibactam (fixed concentration of 4 µg/mL) when tested against ESBLphenotype and carbapenem-resistant Enterobacteriaceae collected from USA hospitals (2009-2011).

Methods

Organisms collection: A total of 7789 Enterobacteriaceae isolates were tested, including ESBL-phenotype E. coli (285) and Klebsiella spp. (411), AmpC stably derepressed Enterobacter spp. (155), meropenem-non-susceptible Klebsiella spp. (mostly KPCproducing; 115), and *Enterobacter* spp. (14). The strains were consecutively collected in 2009-2011 from 72 USA medical centers. Isolates were sent to the coordinating laboratory (JMI Laboratories, North Liberty, Iowa, USA) for reference susceptibility testing. Only one strain per patient infection episode was included in the surveillance program.

<u>Susceptibility testing</u>: Isolates were tested for susceptibility to ceftaroline-avibactam and multiple comparator agents by reference broth microdilution methods as described by Clinical and Laboratory Standards Institute (CLSI) M07-A9 (2012) document, and CLSI interpretations were based on M100-S22 (2012) breakpoints, while ceftaroline interpretations were based on the breakpoint criteria established by the USA-FDA. Ceftaroline was combined with avibactam at a fixed concentration of 4 µg/mL and concurrent testing of quality control (QC) strains assured proper testing conditions. All QC results were within published ranges.

Results

- Ceftaroline-avibactam was active against non-ESBL strains (MIC_{50/90} of 0.06/0.12 µg/mL; 99.8 and 100.0% inhibited at \leq 0.5 and \leq 1 µg/mL, respectively) as well as ESBLphenotype *Klebsiella* spp. (MIC_{50/90} of 0.12/1 µg/mL; 89.1 and 97.1% inhibited at ≤ 0.5 and $\leq 1 \mu g/mL$, respectively). Among ESBL-phenotype *Klebsiella* spp., resistance rates to "third-generation" cephalosporins were high (86.6 and 68.6% resistance to ceftriaxone and ceftazidime, respectively, according to CLSI breakpoints), and only 72.0 and 62.0% of strains were susceptible to meropenem and gentamicin, respectively (Tables 1 and 2)
- Ceftaroline-avibactam (MIC_{50/90} of 0.5/1 μg/mL; 97.4%) inhibited at $\leq 2 \mu g/mL$) and tigecycline (MIC_{50/90} of 0.5/2 µg/mL; 95.7% susceptible by USA-FDA breakpoint criteria) were the most active compounds tested against 115 meropenem-non-susceptible (MIC, ≥2 µg/mL) Klebsiella spp. isolates (Tables 1 and 2)
- *E. coli* isolates were highly susceptible to ceftarolineavibactam (MIC_{50/90.} ≤0.03/0.06 µg/mL) with >99.9% of strains inhibited at ceftaroline-avibactam MIC of ≤0.5 µg/mL. The highest ceftaroline-avibactam MIC was only 0.25 µg/mL among non-ESBL-phenotype strains (MIC_{50/90} ≤0.03/0.06 µg/mL)
- ESBL-phenotype *E. coli* strains exhibited high resistance rates to ceftriaxone (85.6%), ceftazidime (50.9%), gentamicin (30.2%) and levofloxacin (78.9%), but were very susceptible to ceftaroline-avibactam (MIC_{50/90} 0.06/0.25 μ g/mL; highest MIC, 1 μ g/mL [one strain]; Tables 1 and 2)
- Among 801 *Enterobacter* spp. strains, 95.9% were inhibited by ceftaroline-avibactam at $\leq 0.5 \ \mu g/mL$ (MIC_{50/90} 0.12/0.5 µg/mL). Meropenem (MIC_{90.} ≤0.12 µg/mL; 98.3% susceptible) and tigecycline (MIC_{50/90}, 0.25/1 μ g/mL; 98.9% susceptible) were also very active against Enterobacter spp. Among ceftazidime-non-susceptible (MIC, $\geq 8 \mu g/mL$) Enterobacter spp. strains, 80.0 and 96.1% of strains were inhibited at ceftaroline-avibactam MIC of ≤0.5 and ≤1 µg/mL, respectively (Tables 1 and 2)
- Overall, 97.9% of Enterobacteriaceae strains were inhibited at ceftaroline-avibactam MIC of $\leq 0.5 \,\mu g/mL$, which is the susceptibility breakpoint established for ceftaroline alone by the USA-FDA against indicated Enterobacteriaceae species (Tables 1 and 2); 99.5% at $\leq 1 \mu g/mL$. Meropenem and tigecycline covered 98.2 and 97.9% of all strains, respectively.

(2011)

	No. of isolates (cumulative %) inhibited at ceftaroline-avibactam MIC (µg/mL) of:									
Organism (no. tested)	≤0.03	0.06	0.12	0.25	0.5	1	2	≥4		
Klebsiella spp. (2,928)	710 (24.3)	1297 (68.6)	571 (88.1)	212 (95.3)	87 (98.3)	39 (99.6)	9 (99.9)	3 (100.0)		
Non-ESBL-phenotype (2517)	673 (26.7)	1226 (75.5)	458 (93.6)	125 (98.6)	29 (99.8)	6 (100.0)	-	-		
ESBL-phenotype (411)	37 (9.0)	71 (26.3)	113 (53.8)	87 (74.9)	58 (89.1)	33 (97.1)	9 (99.3)	3 (100.0)		
Meropenem-non-S (115)	7 (6.1)	6 (11.3)	14 (23.5)	19 (40.0)	31 (67.0)	28 (91.3)	8 (98.3)	2 (100.0)		
E. coli (2517)	1531 (60.8)	783 (91.9)	158 (98.2)	38 (99.7)	6 (>99.9)	1 (100.0)	-	-		
Non-ESBL-phenotype (2,232)	1443 (64.7)	676 (94.9)	97 (99.3)	16 (100.0)	-	-	-	-		
ESBL-phenotype (285)	88 (30.9)	107 (68.4)	61 (89.8)	22 (97.5)	6 (99.7)	1 (100.0)	-	-		
Enterobacter spp. (801)	116 (14.5)	212 (41.0)	249 (72.0)	139 (89.4)	52 (95.9)	27 (99.3)	5 (99.9)	1 (100.0)		
Ceftazidime-S (646)	114 (17.7)	193 (47.5)	227 (82.7)	93 (97.1)	17 (99.7)	2 (100.0)	-	-		
Ceftazidime-non-S (155)	2 (1.3)	19 (13.6)	22 (27.7)	46 (57.4)	35 (80.0)	25 (96.1)	5 (99.4)	1 (100.0)		
Meropenem-non-S (14)	-	-	1 (7.1)	3 (28.6)	3 (50.0)	4 (78.6)	2 (92.9)	1 (100.0)		
All Enterobactereaceae	2666 (34.2)	2759 (69.7)	1300 (86.3)	593 (94.0)	308 (97.9)	124 (99.5)	26 (99.8)	13 (100.0)		
Abbreviations: ESBL = extended-spectrum β-lactamase; S = susceptible.										

Table 2. Activity of Ceftaroline-Avibactam, Ceftaroline and Comparator Antimicrobial Agents When Tested against Enterobacteriaceae Collected from USA Hospitals (2009-2011)

Organism/antimicrobial	MIC (µg/mL)		%S /	%R ^a	Organism/antimicrobial		MIC (ug/mL)		%S/	%S / %Rª	
agent (no. tested)	MIC ₅₀	MIC ₉₀	Range	CLSI ^a	EUCAST ^a	agent (no. tested)	MIC ₅₀	MIC ₉₀	Range	CLSI ^a	EUCASTa
Klebsiella spp. ^b (2,928)						ESBL phenotype (285)	00				
Ceftaroline-avibactam	0.06	0.25	≤0.03 – 16	- / -	- / -	Ceftaroline-avibactam	0.06	0.25	≤0.03 – 1	- / -	- / -
Ceftaroline ^c	0.12	>16	≤0.015 – >16	82.6 / 14.3	- / -	Ceftarolinec	>16	>16	0.06 -> 16	7.7 / 89.8	- / -
Ceftriaxone	≤0.25	>8	≤0.25 – >8	87.3 / 12.2	87.3 / 12.2	Ceftriaxone	>8	>8	≤0.25 – >8	11.9 / 85.6	11.9 / 85.6
Ceftazidime	≤1	8	≤1 – >16	89.5 / 9.6	87.8 / 10.5	Ceftazidime	16	>16	≤1 – >16	37.2 / 50.9	13.3 / 62.8
Ampicillin/sulbactam	8	>16	≤2 – >16	73.2 / 16.3	- / 26.8	Ampicillin/sulbactam	>16	>16	≤2 – >16	11.6 / 69.1	- / 88.4
Piperacillin/tazobactam	2	32	≤0.5 – >64	89.3 / 8.7	84.1 / 10.7	Piperacillin/tazobactam	8	>64	≤0.5 – >64	73.7 / 10.5	59.3 / 26.3
Tigecyclined	0.25	1	0.06 ->4	98.5 / 0.2	95.0 / 1.5	Tigecyclined	0.12	0.25	0.06 – 1	100.0 / 0.0	100.0 / 0.0
Cefuroxime	≤2	>16	≤2 – >16	81.0 / 14.5	81.0 / 19.0	Gentamicin	≤2	>8	≤2 – >8	69.1 / 30.2	68.1/30.9
Gentamicin	≤2	≤2	≤2 ->8	93.6 / 4.9	92.9/6.4	Levofloxacin	>4	>4	≤0.5 - >4	20.4 / 78.9	20.4 / 79.6
Levofloxacin	≤0.5	4	≤0.5 - >4	89.7 / 9.2	88.2 / 10.3	Meropenem	≤0.12	≤0.12	≤0.12 – 4	99.6 / 0.4	99.6/0.0
Meropenem	≤0.12	≤0.12	≤0.12 ->8	96.1/3.7	96.3 / 2.8	Enterobacter spp. ^e (801)	0.40	0.5	<0.02 4	1	1
Coftereline evibertem)	0.12	<0.02 1	1	1		0.12	0.5	$\leq 0.03 - 4$	- / -	-/-
	0.00	0.12	$\leq 0.03 - 1$	-/-	-/-	Cettaionne	<0.25	>10	$\leq 0.013 - >10$	74.2/22.3	- / -
Cettaionne	<0.12	<0.5	$\leq 0.015 - >10$	90.7 / 1.1	100 0 / 0 0	Cettazidimo	≤0.25 <1	>0	<u> 1 - 16</u>	806/180	77 0 / 10 /
Ceftazidime	≤0.25 <1	≤0.25 <1	≤0.25 − 1 <1 _ 1	100.0 / 0.0	100.0 / 0.0	Ampicillin/sulbactam	_≤1 16	>10	≤1 - >10 <2 - >16	37 2 / 39 6	- / 62.8
Ampicillin/sulbactam	<u></u>	16	<u> </u>	842/50	- / 15 8	Piperacillin/tazobactam	2	>10 64	<u></u>	836/77	70 0 / 16 /
Piperacillin/tazobactam	2	8	<u>-</u> <0 5 - \64	987/06	94 1 / 1 3	Tigecyclined	0.25	1	-0.3 - 204	989/00	945/11
Tigecyclined	0.25	0.5	0.06 - 54	98.8/0.1	96.0/1.2	Gentamicin	<2	<2	<2 - 58	95 0 / <i>4 4</i>	94.0 / 1.1
Gentamicin	<2	<2	<2 - >8	98.7 / 1.1	987/13	Levofloxacin	- <u>-</u> <0.5	<0.5	<0 5 - >4	94.8/4.1	936/53
Levofloxacin	 <0.5	_∠ <0.5	<0.5 - >4	979/14	969/21	Meropenem	<0.12	=0.0 <0.12	<0.12 - >8	983/14	986/02
Meropenem	<0.12	<0.12	<0.12 - 0.25	100.0/0.0	100 0 / 0 0	ceftazidime-susceptible (64)	6)	=0.12	-0.12 20	00.07 1.4	00.07 0.2
ESBL phenotype (411)	-0.12	-0.12	-0.12 0.20				0.12	0.25	≤0.03 – 1	- / -	- / -
Ceftaroline-avibactam	0.12	1	≤0.03 – 16	- / -	- / -		0.12	0.5	≤0.015 - >16	92.0/4.0	- / -
	>16	>16	0.06 - >16	2.4 / 95.6	- / -	Ceftriaxone	≤0.25	0.5	≤0.25 ->8	95.8 / 2.8	95.8 / 2.8
Ceftriaxone	>8	>8	≤0.25 - >8	9.5 / 86.6	9.5 / 86.6	Ceftazidime	_ <u>_</u> ≤1	≤1	≤1 – 4	100.0 / 0.0	96.6 / 0.0
Ceftazidime	>16	>16	≤1 – >16	25.5 / 68.6	13.1 / 74.5	Ampicillin/sulbactam	16	>16	≤2 – >16	46.1 / 25.9	- / 53.9
Ampicillin/sulbactam	>16	>16	2 -> 16	5.6 / 84.9	- / 94.4	Piperacillin/tazobactam	2	4	≤0.5 – 64	99.1 / 0.0	96.7 / 0.9
Piperacillin/tazobactam	>64	>64	1 ->64	31.4 / 58.4	22.6 / 68.6	Tigecvclined	0.25	0.5	0.12 – 4	99.2 / 0.0	97.1 / 0.8
Tigecyclined	0.5	2	0.06 ->4	96.4 / 0.5	89.1 / 3.6	Cefuroxime	8	>16	≤2 – >16	68.3 / 15.3	68.3/31.7
Gentamicin	≤2	>8	≤2 – >8	62.0 / 28.2	56.9/38.0	Gentamicin	≤2	≤2	≤2 – >8	98.9 / 0.9	98.6 / 1.1
Levofloxacin	>4	>4	≤0.5−>4	39.3 / 57.3	34.6 / 60.7	Levofloxacin	≤0.5	≤0.5	≤0.5−>4	98.3 / 0.8	98.0 / 1.7
Meropenem	≤0.12	>8	≤0.12 – >8	72.0 / 26.0	74.0 / 20.0	Meropenem	≤0.12	≤0.12	≤0.12 – 2	99.8 / 0.0	100.0 / 0.0
meropenem-non-susceptible	e (115)					ceftazidime-non-susceptible	e (155)				
Ceftaroline-avibactam	0.5	1	≤0.03 – 16	- / -	- / -	Ceftaroline-avibactam	0.25	1	≤0.03 – 4	- / -	- / -
Ceftaroline ^c	>16	>16	4->16	0.0 / 100.0	- / -	Ceftaroline ^c	>16	>16	1 – >16	0.0 / 98.7	- / -
Ceftriaxone	>8	>8	8 -> 8	0.0 / 100.0	0.0 / 100.0	Ceftriaxone	>8	>8	2->8	0.0 / 98.1	0.0 / 98.1
Ceftazidime	>16	>16	2->16	1.7 / 93.9	0.0/98.3	Ceftazidime	>16	>16	8->16	0.0 / 92.9	0.0 / 100.0
Ampicillin/sulbactam	>16	>16	>16	0.0 / 100.0	- / 100.0	Ampicillin/sulbactam	>16	>16	16 – >16	0.0 / 96.8	- / 100.0
Piperacillin/tazobactam	>64	>64	8->64	0.9 / 98.3	0.9 / 99.1	Piperacillin/tazobactam	64	>64	1 – >64	19.4 / 40.0	9.7 / 80.6
Tigecyclined	0.5	2	0.06 ->4	95.7 / 0.9	88.7 / 4.3	Tigecyclined	0.25	2	0.12 – 4	97.4 / 0.0	83.9 / 2.6
Gentamicin	8	>8	≤2 – >8	49.6 / 34.8	42.6 / 50.4	Gentamicin	≤2	>8	≤2 – >8	78.7 / 18.7	76.8 / 21.3
Levofloxacin	>4	>4	≤0.5 – >4	9.6 / 90.4	7.8/90.4	Levofloxacin	≤0.5	>4	≤0.5−>4	80.0 / 18.1	75.5 / 20.0
Meropenem	>8	>8	2 – >8	0.0 / 93.0	7.0 / 71.3	Meropenem	≤0.12	0.5	≤0.12 – >8	91.6 / 7.1	92.9 / 1.3
Escherichia coli (2,517)					<u>.</u>	meropenem-non-susceptibl	le (14)				
Ceftaroline-avibactam	≤0.03	0.06	≤0.03 – 1	-/-	- / -	Ceftaroline-avibactam	0.5	2	0.12 – 4	-/-	- / -
Cettaroline ^c	0.12	8	≤0.015 - >16	84.3 / 12.8	-/-	Ceftaroline	>16	>16	>16	0.0 / 100.0	-/-
Cettriaxone	≤0.25	1	≤0.25 ->8	90.0/9.7	90.0/9.7	Ceftriaxone	>8	>8	8 -> 8	0.0 / 100.0	0.0 / 100.0
	≤1	≤1 40	≤1 — >16	92.9 / 5.8	90.2 / 7.1		>16	>16	4 -> 16	7.1/92.9	0.0/92.9
Ampicillin/sulbactam	8	>16	≤2 - >16	51.2/26.6	- / 48.8	Ampicillin/sulbactam	>16	>16	>16	0.0 / 100.0	- / 100.0
Piperacillin/tazobactam	2	8	≤0.5 - >64	94.2/3.0	91.6 / 5.8	Piperacillin/tazobactam	>64	>64	32 ->64	0.0 / 78.6	0.0 / 100.0
	0.12	0.25	≤0.03 – 2	100.0 / 0.0	99.9/0.0		1	2	0.25 – 2	100.0 / 0.0	78.6/0.0
Gentamicin	≤Z	>8	$\leq 2 - > 8$	87.6/11.8	86.7 / 12.4	Gentamicin	8	>8	$\leq 2 - > 8$	42.9/50.0	35.7 / 57.1
Levonoxacin	≤0.5 <0.12	>4	≤0.5 — >4	69.4 / 30.0 5 00 0 / 30.0	b9.3/30.b	Levolloxacin	>4	>4	≤0.5 – >4	35.7 / 57.1	21.4/04.3
non ESPL phonotype (2.222)	≥0.1Z	≤0.1Z	≤0.12 - 4	>99.97<0.1	>99.970.0	Enterobactoriacoao (7.780)	0	>0	2 - >0	0.0776.0	21.4/14.3
Coftaroline-avibactam	.) <0.03	0.06	<0.03 - 0.25	- / -	- / -	Coftarolino-avibactam	0.06	0.25	<0.03 - 16	- / -	- / -
	<u>≤0.05</u>	0.00	$\leq 0.03 - 0.23$	-/-	- / -		0.00	0.25	$\leq 0.03 - 10$	- / -	- / -
Cettaionne	<0.12	<0.5	$\leq 0.015 - >10$	94.1/2.9	100 0 / 0 0	Cettriaxono	<0.12	210	$\leq 0.013 - >10$	877/14.9	-/- 877/115
Cettazidime	<u>≤0.25</u> <1	<u>≤</u> 0.25 <1	<u>30.25 – 1</u> <1 <u>1</u>	100.0 / 0.0	100.0 / 0.0	Cettazidime	<u>⊐0.25</u> <1	0	<u>30.25 - >0</u>	00 2 / 8 7	880/98
Ampicillin/sulbactam	8	⊺ ∖16	≤1 = 1 <2 _ \16	56 3 / 21 2	- / 43 7	Ampicillin/sulbactam		- - 16	<u>-</u> - - - - - - - - - - - - - - - - - -	58 3 / 25 0	- / 41 7
Piperacillin/tazohactam	2		<0 5 - \64	96 8 / 2 N	957/22	Piperacillin/tazobactam	2	16	<0.5 - \64	915/55	881/85
Tigecyclined	0 12		<0.0 = 204 <0.03 = 2	100.072.0	999/00	Tigecyclined	0.25	1	≤0.03 - >4	979/02	932/21
Gentamicin	<2	4	_0.00 - ∠ <2 _ ∖8	90.0 / 0.0	89 1 / 10 0	Gentamicin	<2	<2	<2 - >8	915/74	903/85
Levofloxacin	_ <u>_</u> ≤0.5	т 54	<u> −</u>	757/237	756/243	Levofloxacin	<u>∽</u> ∠ ≤0.5	>4	$\leq 0.5 - >4$	829/157	81 4 / 17 1
Meropenem	_0.0 ≤0.12	≤0 12	≤0.12 – 0.25	100 0 / 0 0	100 0 / 0 0	Meropenem	<u>_</u> 0.0 ≤0.12	≤0.12	≤0.12 - >8	982/16	98 4 / 1 1
a Criteria as published by the (121				-0.16	-9.12		0012 / 110	
b. Includes: Klebsiella oxytoca (633 strains), K. ozaenae (2 strains), K. pneumoniae (2230 strains), and unspeciated Klebsiella (63 strains).											
c. USA-FDA breakpoints were applied [Teflaro® Prescribing Information, 2012].											
d. USA-FDA breakpoints were applied [Tygacil® Prescribing Information, 2011].											
e. Includes: Enterobacter aerog	genes (206 str	rains), <i>E. asburia</i>	e (4 strains), E. cancer	rogenus (1 strain), <i>E. cloa</i>	<i>cae</i> (583 strains), <i>E. g</i>	ergoviae (1 strain), <i>E. hormaechei</i> (1	l strain), <i>E. int</i>	ermedius (1 straii	n), <i>E. sakazakii</i> (1 strain),	and unspeciated Enter	robacter (3 strains

Table 1. Summary of Ceftaroline-Avibactam Activity Tested Against Organisms Collected from Patients Hospitalized in USA Medical Centers

Helio S. Sader, MD, PhD JMI Laboratories North Liberty, IA, USA www.jmilabs.com ph. 319.665.3370 fax 319.665.3371 helio-sader@jmilabs.com

Conclusions

- Avibactam can effectively lower ceftaroline MIC values for Enterobacteriaceae producing clinically significant β-lactamases.
- Ceftaroline-avibactam was highly active against Enterobacteriaceae-producing KPC, various ESBL types, and AmpC (chromosomally derepressed or plasmidmediated) enzymes.
- In summary, ceftaroline-avibactam shows potent in vitro activity against pathogens associated with infections caused by multidrug-resistant Enterobacteriaceae in USA hospitals (2009–2011).

References

- Castanheira M. Sader HS. Farrell DJ. Mendes RE. Jones RN (2012). Activity of ceftaroline-avibactam tested against gram-negative organism populations, including strains expressing one or more beta-lactamases and methicillinresistant Staphylococcus aureus carrying various SCCmec types. Antimicrob Agents Chemother in press.
- Clinical and Laboratory Standards Institute (2012). M07-A9. Methods for dilution antimicrobial susceptibility tests for bacteria that grow aerobically; approved standard: ninth edition. Wayne, PA: CLSI.
- Clinical and Laboratory Standards Institute (2012). M100-S22. Performance standards for antimicrobial susceptibility testing: 22nd informational supplement. Wayne, PA: CLSI.
- Endimiani A, Choudhary Y, Bonomo RA (2009). In vitro activity of NXL104 in combination with beta-lactams against Klebsiella pneumoniae isolates producing KPC carbapenemases. Antimicrob Agents Chemother 53: 3599-
- Louie A, Castanheira M, Liu W, Grasso C, Jones RN, Williams G, Critchley I, Thye D, Brown D, Vanscoy B, Kulawy R, Drusano GL (2012). Pharmacodynamics of betalactamase inhibition by NXL104 in combination with ceftaroline, examining organisms with multiple types of betalactamases. Antimicrob Agents Chemother 56: 258-270.
- Teflaro[®] Prescribing Information (2012). Available at http://www.frx.com/pi/Teflaro_pi.pdf. Accessed June 2012.
- Tygacil[®] Prescribing Information (2011). Available at www.tygacil.com. Accessed June 2012.

Acknowledgments

This study was supported by Forest Laboratories, Inc. Forest Laboratories, Inc., was involved in the study design and in the decision to present these results. Forest Laboratories, Inc., had no involvement in the analysis, collection, and interpretation of data. Scientific Therapeutics Information, Inc., provided editorial coordination, which was funded by Forest Research Institute, Inc.