ICAAC 2015 C-1053





Abstract

Background: CoNS is a major cause of bloodstream infections, especially in patients with intravenous catheters and prosthetic devices. We evaluated the *in vitro* activity of ceftaroline against a large collection of CoNS from United States (USA) hospitals.

Methods: 1593 CoNS isolates considered clinically significant (multiple infection types) were collected from 71 USA medical centers in 2013-2014 and tested for susceptibility (S) by CLSI reference broth microdilution methods against ceftaroline and numerous comparators. Species identification was performed by MALDI-TOF.

Results: Overall, 59.7% of isolates were oxacillin-resistant (MRCoNS). Ceftaroline (MIC_{50/90}, 0.25/0.5 µg/ml) inhibited 99.2% of CoNS at $\leq 1 \mu g/ml$ (S breakpoint for S. aureus), including 98.7% of MRCoNS. Ceftaroline activity was 4-fold greater than that of vancomycin (MIC_{50/90}, $1/2 \mu g/ml$; 100.0% S) and similar to that of daptomycin (MIC_{50/90}, 0.25/0.5 µg/ml; 99.9% S). The highest ceftaroline MIC value was only $2 \mu g/ml$ (13 strains); which was observed only among S. cohnii (1 of 7; 14.3%), S. epidermidis (0.1%), S. haemolyticus (13.0%) and S. saprophyticus (2.9%). S. epidermidis represented 60.3% of the CoNS collection and was highly S to ceftaroline (MIC_{50/90}, 0.25/0.5 μ g/ml, 99.9% inhibited at $\leq 1 \mu g/ml$). S. lugdunensis and S. hominis (MIC_{50/90}, 0.25/0.5 μ g/ml for both) were the 2nd and 3rd most common CoNS species, respectively, and S. capitis $(MIC_{50/90}, 0.06/0.25 \mu g/ml)$ ranked 4th; all isolates from these three species were inhibited at ceftaroline MIC of $\leq 1 \mu g/ml$ (Table 1). S. haemolyticus represented only 4.8%, was atypically less S to ceftaroline (MIC_{50/90}, 0.5/2 μ g/ml, 87.0% inhibited at $\leq 1 \mu g/ml$) and accounted for 76.9% (10/13) of isolates with ceftaroline MIC >1 μ g/ml. Tigecycline and linezolid were also active against CoNS (≥99.3% S).

Conclusions: Ceftaroline exhibited potent *in vitro* activity against CoNS, including many uncommonly isolated species for which very limited susceptibility information is currently available to guide therapy. Ceftaroline may have a potential role in the treatment of CoNS infections as guided by reference MIC testing results.

Introduction

Ceftaroline fosamil, the prodrug of ceftaroline, is a broad-spectrum parenteral cephalosporin which was approved by the United States Food and Drug Administration (USA-FDA) for the treatment of acute bacterial skin and skin structure infections (ABSSSI) and communityacquired bacterial pneumonia (CABP), and by the European Medicines Agency (EMA) for the treatment of complicated skin and soft tissue infections (cSSTI) and community-acquired pneumonia (CAP). Ceftaroline has demonstrated potent in vitro bactericidal activity against resistant Gram-positive organisms, including methicillin-resistant Staphylococcus aureus (MRSA) and multidrugresistant (MDR) Streptococcus pneumoniae, as well as prevalent Gram-negative organisms.

Coagulase-negative staphylococci (CoNS) represent the most common cause of bacteremia related to indwelling devices and most of these infections are hospital-acquired. Other important infections caused by CoNS include central nervous system shunt infections, native or prosthetic valve endocarditis, urinary tract infections, and endophthalmitis. Resistance to oxacillin and other β -lactams is widespread among CoNS associated with human infections, and although CoNS are usually susceptible to glycopeptides, increased MIC values for teicoplanin ($\geq 4 \mu g/ml$) and/or vancomcyin ($\geq 2 \mu g/ml$) are frequently reported and may relate to poor clinical treatment outcomes. We evaluated the *in vitro* activity of ceftaroline tested against a large collection of CoNS from USA hospitals.

Methods

Organism collection: A total of 1,593 CoNS isolates considered clinically significant (multiple infection types) were collected from 71 USA medical centers in 2013-2014 (one/patient episode) through the AWARE (Assessing Worldwide Antimicrobial Resistance Evaluation) ceftaroline surveillance program. Isolates were submitted to a reference monitoring laboratory (JMI Laboratories, North Liberty, Iowa, USA) where species identifications were confirmed using MALDI-TOF-MS (Bruker Daltonics, Bremen, Germany).

Antimicrobial susceptibility testing: All isolates were tested for susceptibility by reference broth microdilution methods using the Clinical Laboratory Standards Institute recommendations (CLSI; M07-A10, 2015). Susceptibility testing was performed using validated broth microdilution panels manufactured by Thermo Fisher Scientific (Cleveland, Ohio, USA). Categorical interpretation of MIC values was performed according to CLSI (M100-S25, 2015) and validation of MIC values was performed by concurrent testing of CLSI-recommended quality control (QC) strains: S. aureus ATCC 29213 and Enterococcus faecalis ATCC 29212.

Antimicrobial Activity of Ceftaroline and Comparator Agents When Tested against Numerous Species of Coagulase-negative Staphylococcus (CoNS) HS SADER, DJ FARRELL, RK FLAMM, JM STREIT, RE MENDES, RN JONES JMI Laboratories, North Liberty, Iowa, USA

Results

- Among 1,593 CoNS isolates reported as clinically relevant; 602 (37.8%) were from bloodstream infections (BSI), 164 (10.3%) from urinary tract infections and 827 (51.9%) from other infection sites.
- The most frequently isolated species overall were *S. epidermidis* (960 isolates; 60.3%), S. lugdunensis (168 isolates; 10.5%), S. hominis (120 isolates; 7.5%) and *S. capitis* (103 isolates; 6.5%). Among isolates from BSI, the most common species were *S. epidermidis* (371; 61.6%), *S. hominis* (85; 14.1%) and S. capitis (103; 9.6%).
- Ceftaroline (MIC_{50/90}, 0.25/0.5 μ g/ml) inhibited 99.2% of CoNS at \leq 1 μ g/ml (susceptible breakpoint for S. aureus), including 98.7% of oxacillin-resistant CoNS (Table 1). Among isolates from BSI, 99.3% (598/602) were inhibited at ceftaroline MIC of $\leq 1 \mu g/ml$, and isolates with ceftaroline MIC $> 1 \mu g/ml$ were three S. haemolyticus and one S. cohnii isolates with ceftaroline MIC values of 2 µg/ml.
- Overall, 59.7% of isolates were oxacillin-resistant (MRCoNS). Oxacillinresistance rates varied from as low as 1.8% for S. lugdunensis and 27.2% for S. capitis to 100.0% for S. saprophyticus and 76.3% for S. warneri (Table 2). Among S. epidermidis, the oxacillin resistance rate was slightly higher for BSI isolates (76.0%), compared to non-BSI isolates (68.4%).
- Ceftaroline activity was four-fold greater than that of vancomycin ($MIC_{50/90}$, 1/2 μ g/ml; 100.0% susceptible) and similar to that of daptomycin (MIC_{50/90}, 0.25/0.5 µg/ml; 99.9% susceptible; data not shown).
- The highest ceftaroline MIC value was 2 µg/ml; which was observed only among S. cohnii (1 of 7; 14.3%), S. epidermidis (0.1%), S. haemolyticus (13.0%) and S. saprophyticus (2.9%).
- S. epidermidis was highly susceptible to ceftaroline (MIC_{50/90}, 0.25/0.5 μg/ml) 99.9% inhibited at $\leq 1 \mu g/ml$). S. lugdunensis and S. hominis (MIC_{50/90}, 0.25/0.5) µg/ml for both) were the 2nd and 3rd most common CoNS species, respectively, and *S. capitis* (MIC_{50/90}, 0.06/0.25 µg/ml) ranked 4th; all isolates from these 3 species were inhibited at ceftaroline MICs of $\leq 1 \mu g/ml$ (Tables 1 and 2).
- S. haemolyticus represented only 4.8% of CoNS, was atypically less susceptible to ceftaroline (MIC_{50/90}, 0.5/2 μ g/ml, 87.0% inhibited at ≤1 μ g/ml, 13.0% at 2 µg/ml) and accounted for 76.9% of isolates with ceftaroline MIC values of >1 μ g/ml (Tables 1 and 2)
- Highest tigecycline MIC value was 0.5 µg/ml and 99.3% of isolates were susceptible to linezolid (MIC₅₀ and MIC₉₀, 0.5 μ g/ml; Table 2). All linezolid-nonsusceptible isolates (n=11; 0.7%) were S. epidermidis, and seven of them (63.6%) were from BSI.

Table 1. Summary of ceftaroline activity tested against 1,593 clinical isolates of coagulase-negative staphylococci from USA medical centers (2013-2014).

| | | | | | | • | • | • | | | |
|-----------------------|---|------------|------------|------------|------------|-----------|------------|------|-------------|--|--|
| Organism / | No. of isolates (cumulative %) inhibited at MIC (µg/ml) of: | | | | | | | | MIC (µg/ml) | | |
| no. tested | ≤0.03 | 0.06 | 0.12 | 0.25 | 0.5 | 1 | 2 | 50% | 90% | | |
| All isolates (1,593) | 67 (4.2) | 265 (20.8) | 268 (37.7) | 570 (73.4) | 360 (96.0) | 50 (99.2) | 13 (100.0) | 0.25 | 0.5 | | |
| S. capitis (103) | 36 (35.0) | 43 (76.7) | 5 (81.6) | 10 (91.3) | 6 (97.1) | 3 (100.0) | - | 0.06 | 0.25 | | |
| S. epidermidis (960) | 28 (2.9) | 181 (21.8) | 124 (34.7) | 354 (71.6) | 257 (98.3) | 15 (99.9) | 1 (100.0) | 0.25 | 0.5 | | |
| S. haemolyticus (77) | - | 1 (1.3) | 14 (19.5) | 18 (42.9) | 21 (70.1) | 13 (87.0) | 10 (100.0) | 0.5 | 2 | | |
| S. hominis (120) | - | 6 (5.0) | 37 (35.8) | 26 (57.5) | 43 (93.3) | 8 (100.0) | - | 0.25 | 0.5 | | |
| S. lugdunensis (168) | - | 4 (2.4) | 24 (16.7) | 121 (88.7) | 17 (98.8) | 2 (100.0) | - | 0.25 | 0.5 | | |
| S. saprophyticus (35) | - | - | 7 (20.0) | 18 (71.4) | 6 (88.6) | 3 (97.1) | 1 (100.0) | 0.25 | 1 | | |
| S. warneri (38) | 1 (2.6) | 10 (28.9) | 19 (78.9) | 2 (84.2) | 5 (97.4) | 1 (100.0) | - | 0.12 | 0.5 | | |
| Other species (92) | 2 (2.2) | 20 (23.9) | 38 (65.2) | 21 (88.0) | 5 (93.5) | 5 (98.9) | 1 (100.0) | 0.12 | 0.5 | | |

Table 2. Activity of ceftaroline and comparator antimicrobial agents when tested

| Organism / | | | | | CLSI ^a | | | EUCAST ^a | |
|--------------------------------------|-------------------|-------------------|---------------------------|----------------------------|-------------------|--------------|----------------|---------------------|----------------|
| Antimicrobial Agent | MIC ₅₀ | MIC ₉₀ | Range | %S | %I | %R | %S | % | 9 |
| All (1,593) | | | | | | | | | |
| Ceftaroline | 0.25 | 0.5 | ≤0.015 — 2 | (99.2) ^b | - | - | - | - | |
| Oxacillin | 1 | >2 | ≤0.25 — >2 | 40.3 | - | 59.7 | 40.3 | - | 5 |
| Clindamycin | ≤0.25 | >2 | ≤0.25 — >2 | 68.6 | 1.4 | 30.0 | 66.1 | 2.5 | 3 |
| Levofloxacin | 0.25 | >4 | ≤0.12 — >4 | 58.3 | 1.4 | 40.3 | 58.3 | 1.4 | 4 |
| TMP/SMX ^c | ≤0.5 | >4 | ≤0.5 — >4 | 70.5 | - | 29.5 | 70.5 | 16.1 | 1 |
| Tigecycline | 0.06 | 0.12 | ≤0.015 — 0.5 | - | - | - | 100.0 | - | C |
| Linezolid | 0.5 | 0.5 | ≤0.12 — >8 | 99.3 | - | 0.7 | 99.3 | - | (|
| Vancomycin | 1 | 2 | 0.25 — 4 | 100.0 | 0.0 | 0.0 | 100.0 | - | C |
| S. capitis (103) | | | | | | | | | |
| Ceftaroline | 0.06 | 0.25 | ≤0.015 — 1 | (100.0) ^b | - | - | - | - | |
| Oxacillin | ≤0.25 | >2 | ≤0.25 — >2 | 72.8 | - | 27.2 | 72.8 | - | 2 |
| Clindamycin | ≤0.25 | >2 | ≤0.25 — >2 | 85.4 | 1.0 | 13.6 | 83.5 | 1.9 | 1 |
| Levofloxacin | 0.25 | >4 | ≤0.12 — >4 | 66.0 | 0.0 | 34.0 | 66.0 | 0.0 | 3 |
| TMP/SMX ^c | ≤0.5 | ≤0.5 | ≤0.5 — >4 | 98.1 | - | 1.9 | 98.1 | 1.0 | 1 |
| Tigecycline | 0.06 | 0.12 | 0.03 — 0.25 | - | - | - | 100.0 | - | C |
| Linezolid | 0.5 | 1 | 0.25 — 1 | 100.0 | - | 0.0 | 100.0 | - | C |
| Vancomycin | 1 | 1 | 0.5 — 2 | 100.0 | 0.0 | 0.0 | 100.0 | - | C |
| S. epidermidis (960) | 0.05 | 0.5 | <0.04F 0 | | | | | | |
| Ceftaroline | 0.25 | 0.5 | ≤0.015 — 2 | (99.9) ^b | - | - | - | - | _ |
| Oxacillin | 2 | >2 | ≤0.25 — >2 | 28.6 | - | 71.4 | 28.6 | - | 7 |
| Clindamycin | ≤0.25 | >2 | ≤0.25 — >2 | 59.0 | 1.5 | 39.6 | 56.3 | 2.7 | 4 |
| Levofloxacin | 4 | >4 | ≤0.12 — >4 | 46.5 60.0 | 1.8 | 51.8 40.0 | 46.5 60.0 | 1.8 22.2 | 5 1 |
| TMP/SMX ^c | | >4 | ≤0.5 — >4 ≤0.015 — 0.5 | 60.0 | - | 40.0 | 60.0 100.0 | 22.2 | 1 ⁻ |
| Tigecycline | 0.06 | 0.12 | | - | - | - | 100.0 98.9 | - | C 1 |
| Linezolid | 0.5 | 0.5 | ≤0.12 — >8 0.25 4 | 98.9 100.0 | - | 1.1 | 98.9 100.0 | - | 1 |
| Vancomycin S. haemolyticus (77) | 2 | 2 | 0.25 — 4 | 100.0 | 0.0 | 0.0 | 100.0 | - | C |
| Ceftaroline | 0.5 | 2 | 0.06 — 2 | (87.0) ^b | - | _ | _ | _ | |
| Oxacillin | 0.5 >2 | 2 >2 | 0.06 — 2 ≤0.25 — >2 | (87.0)* 35.1 | - | - 64.9 | - 35.1 | - | 6 |
| Clindamycin | ≥2 ≤0.25 | >2 | ≤0.25 — >2 ≤0.25 — >2 | 85.7 | - 3.9 | 04.9 10.4 | 80.5 | - 5.2 | 0. 1. |
| Levofloxacin | <u> </u> | >4 | ≤0.12 — >2 | 41.6 | 0.0 | 58.4 | 41.6 | 0.0 | 5 |
| TMP/SMX ^c | - ≤0.5 | >4 >4 | ≤0.12 — >4 ≤0.5 — >4 | 62.3 | - | 37.7 | 62.3 | 1.3 | 3 |
| Tigecycline | 0.06 | 0.25 | ≤0.015 — 0.25 | - | - | - | 100.0 | - | 0 |
| Linezolid | 0.5 | 1 | 0.25 — 1 | 100.0 | - | 0.0 | 100.0 | _ | C |
| Vancomycin | 1 | 2 | 0.25 — 2 | 100.0 | 0.0 | 0.0 | 100.0 | - | 0 |
| S. hominis (120) | • | _ | 0.20 2 | | 0.0 | 0.0 | | | |
| Ceftaroline | 0.25 | 0.5 | 0.06 — 1 | (100.0) ^b | - | - | - | - | |
| Oxacillin | 1 | >2 | ≤0.25 — >2 | 40.0 | - | 60.0 | 40.0 | - | 6 |
| Clindamycin | ≤0.25 | >2 | ≤0.25 — >2 | 76.7 | 0.8 | 22.5 | 75.8 | 0.8 | 2 |
| Levofloxacin | ≤0.12 | >4 | ≤0.12 — >4 | 66.7 | 0.0 | 33.3 | 66.7 | 0.0 | 3 |
| TMP/SMX ^c | ≤0.5 | 4 | ≤0.5 — >4 | 62.5 | - | 37.5 | 62.5 | 30.8 | 6 |
| Tigecycline | 0.06 | 0.12 | ≤0.015 — 0.25 | - | - | - | 100.0 | - | C |
| Linezolid | 0.5 | 1 | 0.25 — 4 | 100.0 | - | 0.0 | 100.0 | - | C |
| Vancomycin | 1 | 1 | 0.5 — 2 | 100.0 | 0.0 | 0.0 | 100.0 | - | C |
| S. lugdunensis (168) | | | | | | | | | |
| Ceftaroline | 0.25 | 0.5 | 0.06 — 1 | (100.0) ^b | - | - | - | - | |
| Oxacillin | 1 | 1 | ≤0.25 — >2 | 98.2 | - | 1.8 | 98.2 | - | 1 |
| Clindamycin | ≤0.25 | >2 | ≤0.25 — >2 | 87.5 | 0.0 | 12.5 | 86.9 | 0.6 | 1: |
| Levofloxacin | 0.25 | 0.25 | ≤0.12 — >4 | 98.2 | 0.6 | 1.2 | 98.2 | 0.6 | 1 |
| | ≤0.5 0.02 | ≤0.5 0.00 | ≤0.5 — >4 | 99.4 | - | 0.6 | 99.4 100.0 | 0.0 | C |
| Tigecycline | 0.03 | 0.06 | ≤0.015 — 0.12 | - | - | - | 100.0 | - | 0 |
| Linezolid | 0.25 1 | 0.5 1 | ≤0.12 — 1 0.5 — 2 | 100.0 100.0 | - 0.0 | 0.0 0.0 | 100.0 100.0 | - | C C |
| Vancomycin | I | I | 0.5 — 2 | 100.0 | 0.0 | 0.0 | 100.0 | - | C |
| S. saprophyticus (35) Ceftaroline | 0.25 | 1 | 0.12 — 2 | (97.1) ^b | | | | | |
| Oxacillin | 0.25 | >2 | 0.12 — 2 0.5 — >2 | (97.1) ⁵ 0.0 | | - 100.0 | - 0.0 | - | 10 |
| Clindamycin | ا ≤0.25 | >2 >2 | 0.5 — >2 ≤0.25 — >2 | 0.0 88.6 | - 0.0 | 100.0 | 0.0 85.7 | - 2.9 | 1 |
| Levofloxacin | ≤0.25 0.5 | >2 0.5 | ≤0.25 — >2 0.5 — 0.5 | 00.0 100.0 | 0.0 | 0.0 | 85.7 100.0 | 2.9 0.0 | ı C |
| TMP/SMX ^c | 0.5 ≤0.5 | 0.5 | 0.5 — 0.5 ≤0.5 — >4 | 94.3 | - | 0.0 5.7 | 94.3 | 0.0 | 5 |
| Tigecycline | ≤0.5 0.06 | 0.12 | ≤0.5 — >4 0.06 — 0.25 | - | _ | 5.7 | 94.3 100.0 | 0.0 | c C |
| Linezolid | 1 | 1 | 0.00 - 0.25 0.25 - 2 | - 100.0 | - | - 0.0 | 100.0 | _ | C |
| Vancomycin | 1 | 1 | 0.25 - 2 0.5 - 2 | 100.0 | 0.0 | 0.0 | 100.0 | - | C |
| S. warneri (38) | | | | 10010 | 0.0 | 0.0 | | | L |
| Ceftaroline | 0.12 | 0.5 | 0.03 — 1 | (100.0) ^b | - | - | - | - | |
| Oxacillin | 0.5 | >2 | ≤0.25 — >2 | 23.7 | - | 76.3 | 23.7 | - | 7 |
| Clindamycin | ≤0.25 | >2 | ≤0.25 — >2 | 81.6 | 2.6 | 15.8 | 81.6 | 0.0 | 18 |
| Levofloxacin | 0.25 | 0.25 | ≤0.12 — >4 | 94.7 | 0.0 | 5.3 | 94.7 | 0.0 | 5 |
| TMP/SMX ^c | ≤0.5 | ≤0.5 | ≤0.5 — 4 | 97.4 | - | 2.6 | 97.4 | 2.6 | C |
| Tigecycline | 0.06 | 0.12 | 0.015 — 0.12 | - | - | - | 100.0 | - | C |
| Linezolid | 0.5 | 1 | 0.25 — 1 | 100.0 | - | 0.0 | 100.0 | - | C |
| Vancomycin | 1 | 2 | 0.25 — 2 | 100.0 | 0.0 | 0.0 | 100.0 | - | C |
| Other species (92)d | | | | | | | | | |
| Ceftaroline | 0.12 | 0.5 | 0.03 — 2 | (98.9) ^b | - | - | - | - | |
| Oxacillin | 0.5 | >2 | ≤0.25 — >2 | 46.7 | - | 53.3 | 46.7 | - | 5 |
| Clindamycin | ≤0.25 | >2 | ≤0.25 — >2 | 77.2 | 2.2 | 20.7 | 72.8 | 4.3 | 22 |
| Levofloxacin | 0.25 | >4 | ≤0.12 — >4 | 72.8 | 4.3 | 22.8 | 72.8 | 4.3 | 2 |
| TMP/SMX ^c | ≤0.5 | ≤0.5 | ≤0.5 — >4 | 93.5 | - | 6.5 | 93.5 | 4.3 | 2 |
| Tigecycline | 0.06 | 0.12 | ≤0.015 — 0.25 | - | - | - | 100.0 | - | 0 |
| Linezolid | 0.5 | 1 | 0.25 — 1 | 100.0 | - | 0.0 | 100.0 | - | 0 |
| | | | 0.5 — 2 | 100.0 | | | 100.0 | | 0 |

Number in parenthesis indicate percentage inhibited at ≤1 µg/ml, the CLSI and USA-FDA susceptible breakpoint for *S. aureus*.

TMP/SMX = trimethoprim/sulfamethoxazole. Organisms include: S. arlettae (1), S. auricularis (5), S. capitis (103), S. schleiferi (4), S. cohnii (7), S. caprae (13), S. epidermidis (960), S. haemolyticus (77), S. hominis (120), S. intermedius (4), S. lentus (3), S. lugdunensis (168), S. pasteuri (2), S. pettenkoferi (10), S. pseudintermedius (6), S. pseudintermedius / ntermedius (5), S. saprophyticus (35), S. sciuri (2), S. simulans (27), Unspeciated Staphylococcus (2), S. warneri (38), S. xylosus (1).

Ceftaroline exhibited potent in vitro activity against CoNS, including many uncommonly isolated species for which very limited susceptibility information is available to guide contemporary therapy.

- 2015.

The authors would like to thank all participants of the Assessing Worldwide Antimicrobial Resistance Evaluation (AWARE) program for providing bacterial

This study was supported by Cerexa, Inc., an Allergan affiliate. Forest Laboratories, LLC, an Allergan affiliate provided financial support for the analysis of the data and was involved in the design and decision to present these results. Neither Cerexa, Inc. nor Forest Laboratories, LLC, had any involvement in the collection, analysis, and interpretation of data.

Conclusions

Ceftaroline may have a potential role in the treatment of CoNS infections as guided by reference MIC results.

References

1. Ahlstrand E, Svensson K, Persson L, Tidefelt U, Soderquist B (2011). Glycopeptide resistance in coagulase-negative staphylococci isolated in blood cultures from patients with hematological malignancies during three decades. Eur J Clin Microbiol Infect Dis 30: 1349-1354.

2. Clinical and Laboratory Standards Institute (2015). M07-A10. Methods for dilution antimicrobial susceptibility tests for bacteria that grow aerobically; approved standard- tenth edition. Wayne, PA: CLSI.

3. Clinical and Laboratory Standards Institute (2015). *M100-S25.* Performance standards for antimicrobial susceptibility testing: 25th informational supplement. Wayne, PA: CLSI.

4. Critchley IA, Eckburg PB, Jandourek A, Biek D, Friedland HD, Thye DA (2011). Review of ceftaroline fosamil microbiology: integrated FOCUS studies. J Antimicrob Chemother 66 Suppl. 3: iii45-iii51.

5. Sader HS, Jones RN, Stilwell MG, Flamm RK (2014). Ceftaroline activity tested against uncommonly isolated gram-positive pathogens: Report from the SENTRY Antimicrobial Surveillance Program (2008-2011). Int J Antimicrob Agents 43: 284-286.

6. TEFLARO® (2012). (Ceftaroline fosamil) injection for intravenous (IV) use. Available at: http://www.frx.com/pi/teflaro pi.pdf. Accessed January 1,

7. Widerstrom M, Wistrom J, Sjostedt A, Monsen T (2012). Coagulasenegative staphylococci: update on the molecular epidemiology and clinical presentation, with a focus on Staphylococcus epidermidis and Staphylococcus saprophyticus. Eur J Clin Microbiol Infect Dis 31: 7-20.

Acknowledgment