Epidemiologic Analysis of a Worldwide Collection of Escherichia coli ST131 Using the 1928D Core Genome Multilocus Sequence Typing **Reveals Country Specific and Globally Disseminated Clades**

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Introduction

- Extraintestinal lineage of pathogenic Escherichia coli belonging to multilocus sequence type (ST) 131 has emerged as an international multidrug-resistant (MDR) high-risk clone
- Increasing antimicrobial resistance among E. coli isolates worldwide can be associated with the expansion of ST131 isolates that harbor virulence factors and cause more severe infections when compared to other antimicrobialresistant *E. coli* isolates
- Analyzing core genomic sequencing data using >2,500 genes instead of multilocus sequence typing (MLST) that evaluates only 7 genes enables further discrimination of this global clone
- We evaluated the epidemiology of a global ST131 E. coli collection and unrelated STs using the core genome MLST (cgMLST) and resistance gene profiles generated by the bioinformatics tool 1928D to better understand the evolution and dissemination of ST131 isolates and compared them to a small set of non-ST131 E. coli

Materials and Methods

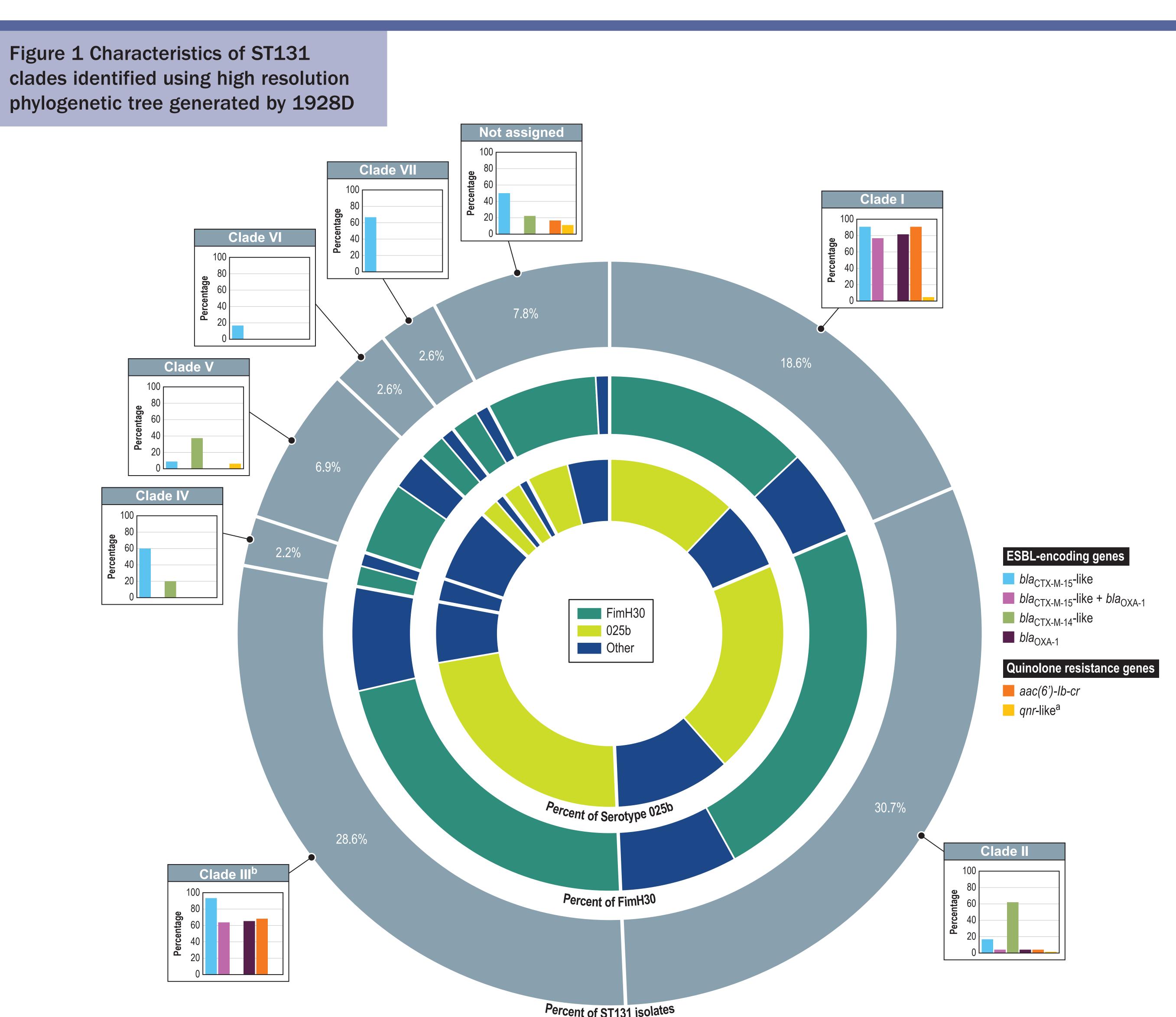
- A total of 259 *E. coli* clinical isolates belonging to ST131 (n=206), ST131single loci variants (SLV; n= 25), and non-ST131 (n=28) collected from 27 countries during 2016-2018 were selected for analysis
- All isolates were subjected to whole genome sequencing on MiSeq (Illumina, San Diego, CA, USA)
- High-quality input DNA was extracted and purified using the KingFisher Cell and Tissue DNA kit (Thermo Scientific, Waltham, Massachusetts, USA) in a robotic workstation KingFisher[™] Flex Magnetic Particle Processor (Thermo Scientific)
- DNA libraries were prepared using the Nextera XT DNA Library Preparation Kit (Illumina) and sequenced with a target depth of coverage >70X - Each raw data set was quality assured, error corrected, and assembled using SPAdes v. 3.11.0
- An *in-silico* PCR with zero mismatches as described in the literature was used to identify serotype 025b, and Fimtyper (https://cge.cbs.dtu.dk /services/FimTyper/) was used to identify isolates belonging to FimH30 type
- Raw reads (FASTQ files) were uploaded onto the 1928 Diagnostics *E. coli* platform (1928D v. 2019-07.5) for analysis
- MLST, cgMLST, and resistance gene prediction outputs were generated
- Custom analysis was performed comparing >2,500 genes to determine allele differences and core genome MLST analysis among isolates
- Nodes on the phylogenetic tree with distances of ≤ 50 alleles were selected for designating clades, between 51 and 60 were considered outliers of the clade, and >60 were considered distinct clades
- Allelic distance among the major clades assigned was analyzed to identify alleles that distinguish these clades to add further granularity

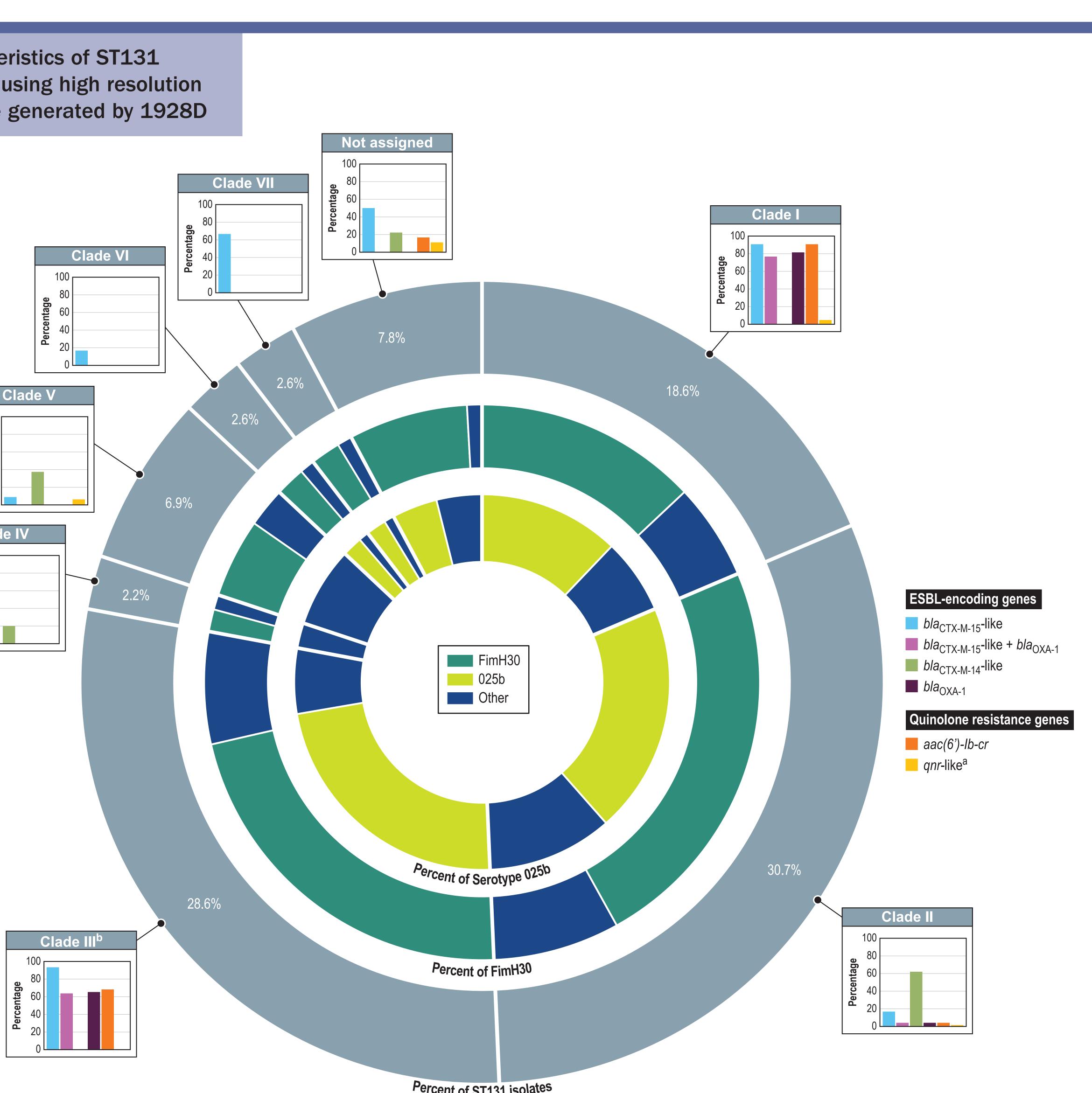
Results

- Among 231 ST131 and SLV E. coli isolates, 7 clades were identified applying cgMLST allele distance of \leq 50 as a cutoff (Table 1, Figure 1)
- The majority of ST131 isolates were categorized into clades I, II, or III (77.9%; 180/231)
- Isolates within the 3 major clades were observed on multiple continents (Table 1)
- A total of 18 isolates were not assigned to clades since their allelic distance from ST131 clades was >60
- The most allelic distance between the 7 ST131 clades was 216, while unrelated STs showed variable allelic distance among isolates within that ST Isolates belonging to ST1193 were closely related genetically and displayed an allelic distance of 30 despite originating from different continents (countries of Australia, United States, and Brazil)
- >2,000 differences were observed among ST131 and unrelated types
- Clades I, II, and III were very similar and only showed <60 allelic distance Based on >95% concordance, 11 alleles differentiated clades II and III from
- clade I, while 6 alleles separated clades I and III from clade II (Table 2) - Outliers of clade III were not included in this analysis
- Isolates in clades I to IV were ciprofloxacin resistant (MIC, $\geq 4 \text{ mg/L}$) and the majority carried an extended-spectrum β -lactamase gene (164/185, 91.3%; Figure 1)
- Isolates in clades I and III predominantly carried bla_{CTX-M-15} (39/43 and 62/66), bla_{0XA-1} (35/43 and 43/66), and aac(6')-lb-cr (39/43, 45/66) The majority of isolates in clade II carried bla_{CTX-M-14}-like (44/71) and less frequently carried bla_{CTX-M-15}-like (11/71) and/or aac(6')lb-cr (3/71)
- Other resistance genes prevalent among 231 ST131 isolates were dfrA17 (118 isolates), tetA (119), sul1 (138), and efflux pumps qacE (138) and/or yphE
- Transferrable cephalosporinases (AmpCs) were present in a minority of isolates (*bla*_{CMV}-like in 4 isolates and *bla*_{DHA-1} in 2)
- The prevalence of type 1 fimbrin D-mannose specific adhesin, FimH30, was similar among isolates belonging to ST131 (173/231; 74.9%) and non-ST131 (20/28; 71.4%) types (data not shown)
- Overall, a smaller number of isolates belonged to FimH41 (n=22) and FimH27 (n=13)
- A significant number of isolates belonging to ST131 clades I, II, III, VI, and VII (64.8-80.3%) were identified as ST131-025b serotype, while only 1 of 28 (3.6%) non-ST131 belonged to this serotype (Figure 1)

- Other STs showed more variability among isolates (ST167, allelic distance 552; ST38, allelic distance 150; and ST69, allelic distance 179)

Figure 1 Characteristics of ST131





gnr-like includes gnrA, gnrB, gnrE, and gnrS and subtype ncludes outliers of the clade III-a and b: 6 isolates ea

Table 1 *E. coli* ST131 isolates distributed among clades based on the cutoff (≤50 allelic distance) criteria and relevant characteristics

| | Allele | Geographic | No. of | Predominant resistance mechanism (no positive) | | | | | | |
|---|-----------------------|-------------------------------|----------|---|----------------------------------|--|--|--|--|--|
| Clade | distance ^a | distribution | isolates | ESBL | Quinolone R | | | | | |
| I | 43 | 9 countries including USA | 43 | bla _{ctx-M-15} (39), bla _{oxa-1} (35) | aac(6')-lb-cr (39) | | | | | |
| П | 47 | 12 countries including USA | 71 | bla _{ctx-M-14} -like ^b (44) | aac(6')-lb-cr (3), qnrB19 (1) | | | | | |
| Ш | 49 (53) | 13 countries including USA | 66 | bla _{CTX-M-15} (61), bla _{OXA-1} (43) | aac(6')-lb-cr (45) | | | | | |
| IV | 40 | Australia and USA | 5 | <i>bla</i> _{CTX-M-15} (3), <i>bla</i> _{CTX-M-14} -like ^b (1) | qnrS1 (1) | | | | | |
| V | 45 (55) | 8 countries including USA | 16 | bla _{ctx-M-14} -like (6) | | | | | | |
| VI | 49 | Costa Rica and USA | 6 | $bla_{\text{CTX-M-211}}$ and $bla_{\text{CTX-M-3}}$ (1) | | | | | | |
| VII | 28 | Costa Rica and USA | 6 | <i>bla</i> _{CTX-M-15} (4) | | | | | | |
| ^a Number in parentheses is allele distance (cgMLST) for outliers of the clade. | | | | | | | | | | |

^bIncludes *bla*_{CTX-M-14}, *bla*_{CTX-M-24}, *bla*_{CTX-M-27}, and *bla*_{CTX-M-134}. ESBL, extended-spectrum β -lactamase; R, resistant

Table 2 Allelic differences between ST131 clades I, II, and III

| Gene | Clade I | Occurrence | Clade II | Occurrence | Clade III | Occurrence % | Different |
|------------|---------|--------------|----------|---------------|-----------|---------------------|-----------|
| identifier | allele | % in clade I | allele | % in clade II | allele | in clade III | clade |
| RS00050 | 22 | 100 | 25 | 100 | 25 | 100 | Clade I |
| RS00065 | 45 | 100 | 46 | 100 | 46 | 98.1 | Clade I |
| RS00070 | 45 | 97.7 | 44 | 98.6 | 44 | 100 | Clade I |
| RS00090 | 59 | 100 | 57 | 100 | 57 | 98.1 | Clade I |
| RS00100 | 42 | 100 | 44 | 100 | 44 | 100 | Clade I |
| RS00105 | 33 | 100 | 35 | 100 | 35 | 100 | Clade I |
| RS00110 | 53 | 100 | 58 | 98.6 | 58 | 98.1 | Clade I |
| RS00120 | 68 | 100 | 63 | 98.6 | 63 | 98.1 | Clade I |
| RS05065 | 41 | 100 | 38 | 100 | 38 | 100 | Clade I |
| RS15715 | 19 | 95.3 | 6 | 98.6 | 6 | 98.1 | Clade I |
| RS23945 | 27 | 100 | 17 | 100 | 17 | 100 | Clade I |
| RS02515 | 61 | 97.7 | 59 | 100 | 61 | 98.1 | Clade II |
| RS02720 | 52 | 100 | 44 | 100 | 52 | 98.1 | Clade II |
| RS17410 | 62 | 100 | 64 | 100 | 62 | 100 | Clade II |
| RS23535 | 55 | 97.7 | 56 | 100 | 55 | 98.1 | Clade II |
| RS24430 | 78 | 100 | 77 | 100 | 78 | 100 | Clade II |
| RS25820 | 82 | 97.7 | 81 | 98.6 | 82 | 100 | Clade II |

Rows highlighted in orange represent alleles that distinguish clade I from clades II and III. Blue rows represent alleles that distinguish clade II from clades I and III. Allele numbers are consecutive numbers assigned by the software for more than 2,500 gene sequences compared.

Allele occurrence of \geq 95.0% was selected to differentiate clades.

Conclusions

- The cgMLST 1928D is a robust platform for epidemiological analysis of E. coli isolates, providing additional granularity when compared to MLST
- A threshold of \leq 50 cgMLST distance was useful for classifying isolates into clades
- Clades II and III were closely related, but carried different bla_{CTX-M} genes, while clades I and III were not as closely related, but both carried bla_{CTX-M-15}, bla_{OXA-1}, and aac(6')-lb-cr
- These findings suggest that these clades might have acquired resistance genes at different points in their genetic evolution
- E. coli ST131 clone has a global presence with bla_{CTX-M-15} and bla_{CTX-M-14} as the major β -lactamases and *aac*(6')-*lb-cr* as the aminoglycoside/quinolone resistance mechanism
- FimH30 and serotype 025b are predominant in ST131 isolates

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References

Clinical and Laboratory Standards Institute (2019). M100Ed29. Performance standards for antimicrobial susceptibility testing: 29th informational supplement. Wayne, PA: CLSI.

EUCAST (2019). Breakpoint tables for interpretation of MICs and zone diameters. Version 9.0, January 2019. Available at: http://www.eucast.org/fileadmin/src/media/PDFs /EUCAST_files/Breakpoint_tables/v_9.0_Breakpoint_Tables.pdf. Accessed January 2019.

Mathers AJ, Peirano G, Pitout JD (2015). Escherichia coli ST131: The quintessential example of an international multiresistant high-risk clone. Adv Appl Microbiol 90: 109–154.

Petty NK, Ben Zakour NL, Stanton-Cook M, et al. (2014). Global dissemination of a multidrug resistant Escherichia coli clone. Proc Natl Acad Sci U S A 111: 5694–5699.

Stoesser N, Sheppard AE, Pankhurst L, et al. (2016). Evolutionary history of the global emergence of the Escherichia coli epidemic clone ST131. MBio 7: e02162.

Woksepp H, Ryberg A, Berglind L, et al. (2017). Epidemiological characterization of a nosocomial outbreak of extended spectrum beta-lactamase Escherichia coli ST-131 confirms the clinical value of core genome multilocus sequence typing. APMIS 125: 1117–1124.

Johnson JR, Clermont O, Johnston B, Clabots C, Tchesnokova V, Sokurenko E, Junka AF, Maczynska B, Denamur E. Rapid and specific detection, molecular epidemiology, and experimental virulence of the 016 subgroup within Escherichia coli sequence type 131. J Clin *Microbiol*. 2014 May; 52(5):1358-65. doi: 10.1128/JCM.03502-13. Epub 2014 Feb 5

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