Induction of Resistance Against Antipseudomonal Agents: Comparison of Novel *β*-Lactam/*β*-Lactamase Inhibitor Combinations and Other **B-Lactam Agents**

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CONCLUSIONS



Our results demonstrate that exposure to meropenem, cefepime, and piperacillin-tazobactam terminal mutants displayed higher increases in MIC values compared to the isolates obtained after exposure to ceftolozanetazobactam, imipenem-relebactam, and ceftazidime-avibactam.



Mutations detected in isolates exposed to 10-day serial passaging involved various genes previously associated to BL resistance, including PBPs and AmpC, efflux, and porin regulators.



This data might indicate that these newer agents could help prevent the emergence of high-level resistance, but additional data is needed for confirmation.



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References

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- 1. Castanheira M, Doyle TB, Smith CJ, et al. (2019). Combination of MexAB-OprM overexpression and mutations in efflux regulators, PBPs and chaperone proteins is responsible for ceftazidime/avibactam resistance in Pseudomonas aeruginosa clinical isolates from US hospitals. J Antimicrob Chemother 74: 2588-2595 2. Castanheira M, Doyle TB, Hubler CM, et al. (2022). The Plethora of Resistance Mechanisms in Pseudomonas aeruginosa: Transcriptome Analysis Reveals a Potential Role of Lipopolysaccharide Pathway Proteins to Novel beta-lactam/beta-lactamase Inhibitor Combinations. J Glob Antimicrob Resist S2213-
- 7165(22)00188-6 3. Clinical and Laboratory Standards Institute (2018). M07Ed11. Methods for dilution antimicrobial susceptibility tests for bacteria that grow aerobically. Wayne, PA:
- 4. Mengin-Lecreulx D, Van Heijenoort J, Park JT (1996). Identification of the mpl gene encoding UDP-N-acetylmuramate: L-alanyl-gamma-D-glutamyl-mesodiaminopimelate ligase in *Escherichia coli* and its role in recycling of cell wall peptidoglycan. *J Bacteriol* 178: 5347-5352.

INTRODUCTION

- Mutation is the main driver of β -lactam (BL) resistance among P
- New β-lactam/β-lactamase inhibitor (BL/BLI) combinations, suc imipenem-relebactam, exhibit good activity against most P. aeru - Limited information is available regarding the potential for the resistance in *P. aeruginosa* compared to older agents.
- We subjected 7 P. aeruginosa isolates to a 10-day serial passage and mechanisms in terminal mutant strains.

MATERIALS AND METHODS

- Seven P. aeruginosa isolates susceptible to antipseudomonal (
- Serial passaging was performed in broth microdilution panels p The broth from the highest concentration of each antimicrobia
- was used to prepare a new 0.5 McFarland standard (5 x 10⁵ 0 This process was repeated for 9 days.
- Antimicrobial agents used for passaging were ceftazidime-avibation meropenem, cefepime, and piperacillin-tazobactam.
- All BLIs were added to wells at a 4 mg/L fixed concentration. The MIC of terminal mutants was determined by using the broth twice on drug-free agar.
- Parent strains and terminal mutants that displayed >2-fold chan genome sequencing at 100X coverage (MiSeq Sequencer; Illu
- Parent isolates were sequenced using long-read WGS (MinION) Data from short- and long-reads were combined using Unicyc
- Single nucleotide polymorphism (SNP) analysis was performed sequences and their follow-up pairs by employing MAUVE V2.4
- Insertion/deletion (INDEL) sites were realigned using IndelRealigned

RESULTS

- Final mutants had MIC increases of 2- to 8-fold for ceftazidime-32-fold for ceftolozane-tazobactam (Figure 1).
- The MIC increases for meropenem (1- to 128-fold), cefepime were slightly higher.
- Cefepime terminal mutants (n=7) exhibited
- Alterations upstream of nalC (n=2) and
- Missense alterations in *ampD*, *mexB*, and the TetR family trar amrR either alone or in combination with mutations in other of
- Three of 7 ceftazidime-avibactam mutants exhibited missense MexAB-OprM regulator, as well as 1 alteration in the intragenic nalC involved in the same regulatory pathway.
- Of the 6 sequenced ceftolozane-tazobactam mutants, 1 had a r ampG and another had a missense mutation in the penicillin-bir gene fts
- Mutations in genes not usually associated with BL resistance including 2 isolates with alterations in the HAMP domain, whi kinase
- Meropenem mutants (n=6) displayed alterations either in oprD (– Within or upstream of nalC (n=3) or its intragenic region, ftsl (
- Upon exposure to piperacillin-tazobactam, mutations in *merR* nalC, and ampD were observed in the terminal mutants.
- Among 2 imipenem-relebactam terminal mutants,
- 1 displayed a nonsense mutation in *pilF*, which encodes a pilu and
- 1 displayed a spermidine synthase that interacts with arnBCL increasing both the expression of MexXY/OprM and the stres
- Notably, ceftolozane-tazobactam and piperacillin-tazobactam r alterations in mpl and galU that assist the recycling of the cell

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<i>Pseudomonas aeruginos</i> ch as ceftazidime-avibact <i>ruginosa</i> isolates from US ese new β-lactam/β-lactar age with 6 antipseudomor	a isolates. am, ceftolozane-tazobactam, and hospitals. mase inhibitor combinations to induce hal agents to evaluate resistance levels	Parent Set #1
β-lactam agents were sub produced according to the al agent that displayed vis CFU/mL) bacterial inocule bactam, imipenem-relebad	ojected to passaging experiments. e CLSI M07 (2018) guidelines. sible growth after overnight incubation um.	Set #2
th microdilution reference	method after subculturing isolates ere subjected to short-read whole	
mina, San Diego, CA, US N; Oxford Nanopore Tech cler v0.4.8-beta and SPA d by comparing the baseli 4.0. aligner from the GATK too	SA). nologies Ltd, Oxford, UK). des. ine short- and long-read assembled lbox v3.8-1-0gf15c13ef and reported.	Set #3 Set #4
e-avibactam, 1- to 8-fold for e (2- to 32-fold), and piper	or imipenem-relebactam, and 2- to racillin-tazobactam (2- to 1024-fold)	Set #5
anscriptional regulator genes (Table 1). mutations in <i>nalD</i> , the c region upstream of	MIC results from the parent isolate to the terminal mutant	
missense mutation in inding protein (PBP) e were also detected, ich contained histidine		Set #6
(<i>n</i> =2); (<i>n</i> =1) <i>phoP</i> (<i>n</i> =1). (1 upstream mutation),		Set #7
lus-forming protein, <i>DTEF-ugd</i> and PmrAB, ss response in the cell. mutants exhibited wall peptidoglycan.	4 5 6 7	

2 4 8 16 >16

Fold increase in MIC

lable 1. SNP analysis of isolates exposed to							antimicropial agents			
	selection	Fold change	Initial MIC	Final MIC	Reference	Alteration	Annotation	Annotation effect	Gene	Function and upstream/downstream genes for intragenic mutation
T	cefepime	32	1	32	G	Т	Arg620Leu	missense	mexB	multidrug efflux RND transporter permease subunit
					Т	G	Thr163Pro	missense		HAMP domain-containing histidine kinase
					G	Т	Ala122Glu	missense	mlaE	lipid asymmetry maintenance ABC transporter permease subunit
					A	G		intragenic		efflux system transcriptional regulator nalC/ hypothetical protein
	ceftazidime-avibactam	8	1	8	Т	G	His56Pro	missense	nalD	efflux system transcriptional repressor
	ceftolozane-tazobactam	0.5	8	16	Т	G	Thr163Pro	missense		HAMP domain-containing histidine kinase
					С	G	Ala547Pro	missense	ftsl	peptidoglycan D,D-transpeptidase
	meropenem	64	0.25	16	G	С	Arg48Gly	missense	nalC	efflux system transcriptional repressor
	piperacillin-tazobactam	256	2	512	С	Т		intragenic		MarR family transcriptional repressor MexR/ multidrug efflux RND transporter periplasmic adaptor subunit MexA
					Α	G	Tyr207His	missense	galU	UTPglucose-1-phosphate uridylyltransferase
					A	G	Leu40Pro	missense	ampD	1,6-anhydro-N-acetylmuramyl-L-alanine amidase
	cefepime	4	2	8	Т	С	Asp155Gly	missense	amrR	TetR family transcriptional regulator
	ceftazidime-avibactam	8	2	16	Α	G	Thr254Ala	missense		glycosyltransferase family 4 protein
	imipenem-relebactam	1	0.5	0.5	С	Т	Gln153*	nonsense	pilF	type 4a pilus biogenesis protein
	meropenem	64	0.12	16	A	G		intragenic		efflux system transcriptional regulator nalC/ hypothetical protein
					Т	A	Thr79Ser	missense	phoP	two-component system response regulator
	piperacillin-tazobactam	8	4	32	С	Т	Arg36Cys	missense		MerR family transcriptional regulator
					С	Т	Gln306*	nonsense	pgi	glucose-6-phosphate isomerase
	cefepime	1	4	4	G	A	Gln357*	nonsense		inactive transglutaminase family protein
	ceftazidime-avibactam	8	0.12	1	Т	С	Ser239Pro	missense		carboxy terminal-processing peptidase
	ceftolozane-tazobactam	4	0.5	2	A	G	Leu365Pro	missense	secY	preprotein translocase subunit
	meropenem	1	0.03	0.03	Т	G	Asp54Ala	missense	uvrY	UvrY/SirA/GacA family response regulator transcription factor
	cefepime	32	1	32	A	C	Val10Gly	missense	ampD	1,6-anhydro-N-acetylmuramyl-L-alanine amidase
	ooftazidima avibaatam	2	1	2			ASIIZ 14501	missense		
		16	0.5	2	т	A		missonso	mpl	
	Celluluzane-lazubaciam	10	0.5	0		G	variz4Giy	1115561156	Πρι	glutamyl-meso-diaminopimelate ligase
	imipenem-relebactam	2	0.25	0.5	Α	G	Asp333Gly	missense		spermidine synthase
	meropenem	4	1	4	Т	C	lle224Val	missense		UDP-glucose 6-dehydrogenase
					С	A	Ala530Glu	missense	gacS	two-component system sensor histidine kinase
	piperacillin-tazobactam	64	2	128	G	Т	Tyr188*	nonsense	mupP	N-acetylmuramic acid 6-phosphate phosphatase
	cefepime	8	1	8	Т	C	Val179Ala	missense		N-acetyl sugar amidotransferase
	ceftazidime-avibactam	8	1	8	G	C	Tyr48*	nonsense	nalD	efflux system transcriptional repressor
	ceftolozane-tazobactam	8	0.5	4	Т	G	Thr163Pro	missense		HAMP domain-containing histidine kinase
					G	Т	Arg101Ser	missense	galU	UTP-glucose-1-phosphate uridylyltransferase
					A	C	His24Pro	missense		hypothetical protein
	meropenem	4	0.5	2	G	A	Ser40Asn	missense	oprD	outer membrane porin
					Т	C	Ser278Pro	missense	oprD	outer membrane porin
	piperacillin-tazobactam	16	2	32	Т	G	Thr24Pro	missense	nalC	efflux system transcriptional repressor
		16	1	16	A	G		intragenic		efflux system transcriptional regulator nalC/ hypothetical protein
	centazioime-avibaciam			2				missense	naiD	APC transcriptional repressor
	aaftalazana tazabaatam	2	0.5	1			Ser93Gly	missense		ABC transporter ATP-binding protein
		2 0	0.5	16	A		Trp4140rg	missense	ampG	
	piperaciiin-lazobaciam	0	2	10	A C	 т	Dho110Dho	synonymous		aliphatic sulfonato ABC transportor pormoaso
	cofonimo	ρ	2	16	C		Phe 119Phe	synonymous	ssuc	transporter permease
	ceftazidime-avibactam	4	2	8	A	G		intragenic	nusu	efflux system transcriptional regulator nalC/
					G	A	Gly544Ser	missense	clpA	ATP-dependent Clp protease ATP-binding
					-	_				subunit
	cettolozane-tazobactam	32	0.5	16	C G	T A	Phe119Phe Ala159Thr	synonymous missense	nusG mpl	transcription termination/antitermination protein UDP-N-acetylmuramate:L-alanyl-gamma-D- glutamyl-meso-diaminopimelate ligase
	meropenem	128	0.25	32	С	Т	Gly132Glu	missense	nalC	efflux system transcriptional repressor
					Α	С	Val471Gly	missense	ftsl	peptidoglycan D,D-transpeptidase
		4.0		0.4	•	•				

UDP-N-acetylmuramate:L-alanyl-gammaglutamyl-meso-diaminopimelate ligase